

EXTENDED REPORT

Significant associations of antidrug antibody levels with serum drug trough levels and therapeutic response of adalimumab and etanercept treatment in rheumatoid arthritis

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ABSTRACT

Objective To evaluate the associations between (1) antidrug antibody (ADAb) and therapeutic response, (2) ADAb and serum drug trough levels and (3) serum drug levels and therapeutic responses in rheumatoid arthritis (RA) patients receiving adalimumab or etanercept. Secondly, we aim (1) to evaluate the concordance between radioimmunoassay and bridging ELISA for ADAb assessment and to evaluate the correlation between two different ELISA methods for detecting drug levels, and (2) to determine the optimal cut-off drug levels for good European League Against Rheumatism (EULAR) response.

Methods ADAb levels were determined by bridging ELISA and radioimmunoassay, and drug levels evaluated using sandwich ELISA among 36 adalimumab-treated patients and 34 etanercept-treated patients at the 6th and 12th month. The optimal cut-off drug levels for EULAR responses were determined by receiver-operating characteristic curve analysis.

Results ADAb was detected in 10 (27.8%) and 13 (36.1%) of adalimumab-treated patients after 12-month therapy using bridging ELISA and radioimmunoassay respectively, but not detected in any of etanercept-treated patients. The presence of ADAb was associated with lower EULAR response and lower drug levels compared with those without ADAb (both $p < 0.001$). Drug trough levels were positively associated with DAS28 decrement (Δ DAS28) (all $p < 0.001$). The optimal cut-off trough levels for adalimumab were 1.274 μ g/mL and 1.046 μ g/mL, and those for etanercept were 1.242 μ g/mL and 0.800 μ g/mL for good EULAR response assessed at the 6th and 12th month, respectively.

Conclusions ADAb levels were inversely correlated with therapeutic response and drug levels. The positive correlation between drug levels and Δ DAS28 indicates that drug monitoring would be useful to evaluate therapeutic response of TNF- α inhibitors.

BACKGROUND

Rheumatoid arthritis (RA) is characterised by synovial inflammation and hyperplasia, cartilage degradation and bone erosions.¹ Tumour necrosis factor (TNF)- α is a crucial inflammatory mediator in synovitis and subsequent tissue damage in RA.^{2,3} Although anti-TNF- α therapy can be effective and well tolerated for RA patients,^{4–6} not all patients respond to them from the start (primary failure)

and the effectiveness diminishes in some patients over time (secondary failure).⁷ The exact mechanism of the inadequate response to anti-TNF- α therapy has not been fully explored.^{8–10}

TNF- α inhibitors can elicit immunogenic responses, including the emergence of antidrug antibodies (ADAb), which results in changes of pharmacokinetics.^{11–12} Several studies indicate that the immunogenicity may be associated with low or undetectable drug levels and reduced therapeutic response to TNF- α inhibitors.^{13–18} Concomitant administration of immunosuppressive agents, such as methotrexate (MTX), reduces immunogenicity.^{14–17,18} Because ADAb to one TNF- α inhibitor are specific and do not cross-react with another different TNF- α inhibitor, RA patients with secondary response failure to one TNF- α inhibitor may benefit from a switch to another TNF- α inhibitor.^{15,16}

Several different methods have been used to assess ADAb, but these assays have their own advantages and disadvantages,^{19–21} and scanty information on the clinical potential of these assays has been reported. Moreover, there are limited data regarding the association of ADAb levels with serum drug levels and therapeutic response.^{14,22,23} There have also been few studies that examined the optimal cut-off trough levels of etanercept and adalimumab in terms of therapeutic response.^{24,25}

The main objective of this study was to evaluate the associations between (1) ADAb and therapeutic response, including low-disease activity and European League Against Rheumatism (EULAR) responses; (2) ADAb levels and MTX dosages or serum drug trough levels; and (3) serum drug trough levels and therapeutic responses in RA patients receiving adalimumab or etanercept. Secondly, we aim (1) to evaluate the concordance grade between radioimmunoassay (RIA) and bridging ELISA for ADAb assessment and to evaluate the correlation between two different sandwich ELISA methods for detecting drug levels, and (2) to determine the optimal cut-off drug trough level for a good EULAR response.

PATIENTS AND METHODS

Patients

Seventy biologic-naïve patients (62 women and 8 men; mean age \pm SD, 56.9 \pm 12.3 years) who fulfilled the 1987 revised criteria of the American College of



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Rheumatology for RA²⁶ started anti-TNF- α therapy according to the guidelines of the British Society for Rheumatology.²⁷ Thirty-four patients received etanercept at a dose of 25 mg twice weekly and 36 patients received adalimumab at a dose of 40 mg every other week, with or without concomitant MTX at a stable dose of 7.5–15 mg weekly. Disease activity was assessed by the 28-joint disease activity score (DAS28) at baseline, and at the 6th and 12th month of anti-TNF- α therapy, respectively.²⁸ Low-disease activity was defined as DAS28 \leq 3.2. The therapeutic response was evaluated after 6 and 12 months of anti-TNF- α therapy using the EULAR response criteria.²⁹ Patients were categorised as good, moderate or poor responders based on the amount of change in the DAS28 and the level of DAS28 reached. Good responders were defined as patients who had a decrease in DAS28 from baseline (Δ DAS28) $>$ 1.2 and a DAS28 \leq 3.2 at evaluation time; moderate responders had either Δ DAS28 $>$ 1.2 and a DAS28 $>$ 3.2 or Δ DAS28 of 0.6–1.2 and a DAS28 \leq 5.1 at evaluation time; and poor responders were those who had either Δ DAS28 $<$ 0.6 or a DAS28 $>$ 5.1 at evaluation time. Blood samples were obtained immediately before etanercept or adalimumab injection at 6 and 12 months of anti-TNF- α therapy. The Ethics Committee of Taichung Veterans General Hospital approved this study, and the written consent of each participant was obtained.

Assessments of antibodies against adalimumab or etanercept

Antibodies against adalimumab and etanercept were detected by bridging ELISA (Progenika Biopharma SA, Derio, Spain) at 6 and 12 months of anti-TNF- α therapy. This assay measures serum levels of free ADAb but lacks sensitivity towards IgG4-ADAb because only the bivalent fraction will be detected.³⁰ The details on this assay are added as online supplementary text.

Antibodies against adalimumab or etanercept were also detected by RIA (Sanquin Diagnostic Services, Amsterdam, The Netherlands) as described previously^{19 20} at 12 months of anti-TNF- α therapy. This assay measures both IgG1 and IgG4 ADAb using an antigen binding test (ABT). Test results were converted into arbitrary units per millilitre (AU/mL) by comparison with dilutions of a reference serum. Patients were defined as positive for anti-adalimumab antibodies if the levels were greater than 12 AU/mL in combination with serum adalimumab levels less than 5.0 mg/L assessed by an ELISA from Sanquin Diagnostic.^{14 20} Similarly, patients were defined as positive for anti-etanercept antibodies if the levels exceeded 12 AU/mL.

Determination of serum trough levels of adalimumab or etanercept

Serum trough levels of adalimumab and etanercept were determined using sandwich ELISA according to the manufacturer's instructions (Progenika Biopharma SA, Derio, Spain) at 6 and 12 months of anti-TNF- α therapy. Serum levels of adalimumab and etanercept were also detected at 12 months of anti-TNF- α therapy by using another ELISA method (Sanquin Diagnostic Services, Amsterdam, The Netherlands) as described previously³¹ based on the principle that adalimumab or etanercept is captured through their capacity to bind TNF- α . The details on these assays are added as online supplementary text.

Determination of the optimal cut-off trough levels for a good EULAR response

Because serum drug trough levels are positively associated with therapeutic response, a search for the optimal cut-off level for

TNF- α inhibitors is important for clinical practice. The optimal drug cut-off level for a good EULAR response was determined using receiver-operating characteristic (ROC) curve analysis.

Statistical analysis

Results are presented as the mean \pm SD or median (IQR). A Fisher's exact test was used for between-group comparisons of ADAb positivity and therapeutic responses. The Kruskal–Wallis test was used for among-group comparison of drug levels in patients with negative or positive ADAb and in EULAR good, moderate and poor responses. When this test showed significant differences, the exact p values were then determined using the Mann–Whitney U test. We assessed the concordance grade between bridging ELISA and RIA for detecting ADAb using the χ^2 tests. The nonparametric Spearman's correlations were determined between (1) ADAb levels and improvement of activity scores (Δ DAS28), MTX dosages, or drug trough levels; (2) two different sandwich ELISA methods for detecting drug levels; and (3) drug trough levels and Δ DAS28. The diagnostic sensitivity, specificity and area under ROC curve (AUC) were determined using MedCalc statistical software V9.3 (MedCalc Software, Belgium, China). A probability of less than 0.05 was considered significant.

RESULTS

Clinical characteristics of RA patients

The majority of RA patients were women, and all patients had active disease (DAS28, mean \pm SD, 6.02 \pm 0.69). Although etanercept-treated patients were older, there were no significant differences in the positive rate of rheumatoid factor and anticyclic citrullinated peptide antibodies, daily dose of corticosteroids or proportion of the used DMARDs between the adalimumab-treated and etanercept-treated patients (table 1).

ADAb levels and their relation to therapeutic response

In 36 adalimumab-treated patients, 8 (22.2%) and 10 (27.8%) were positive for ADAb detected by bridging ELISA at 6 and 12 months of therapy, respectively, while 13 (36.1%) were positive for ADAb using RIA at 12 months of therapy (table 2). After 12 months of adalimumab treatment, ADAb levels in three patients (38, 64 and 180 AU/mL) could be detected by RIA but not by bridging ELISA. The overall agreement between bridging ELISA and RIA was excellent, with a Cohen's kappa coefficient of 0.810 (p $<$ 0.001). After 6 and 12 months of etanercept therapy, no patient developed anti-etanercept antibodies.

When compared with adalimumab-treated patients with negative ADAb (bridging ELISA), those with positive ADAb had significantly higher rates of a poor EULAR response (75% vs 0% at the 6th month and 70% vs 11.5% at the 12th month, both p $<$ 0.001), and lower rates of achieving low-disease activity (DAS28 \leq 3.2) (10% vs 38.5% at the 12th month, p=0.127; figure 1A–C). There was an inverse correlation between ADAb levels and Δ DAS28 (correlation coefficients, r=−0.683, p $<$ 0.001; r=−0.528, p $<$ 0.005, detected by bridging ELISA at the 6th and 12th month, respectively; and r=−0.517, p $<$ 0.005, detected by RIA at the 12th month; figure 1D–F). In addition, MTX dosages were inversely correlated with ADAb levels (r=−0.503, p $<$ 0.005; r=−0.591, p $<$ 0.001, detected by bridging ELISA and RIA, respectively, at 12 months of anti-TNF- α therapy).

ADAb levels and their relation to serum drug trough levels

Patients with negative ADAb (bridging ELISA) had significantly higher drug levels at the 6th and 12th month (median=4.25 μ g/mL, IQR 2.03–11.20 μ g/mL and median=4.19 μ g/mL, IQR

Table 1 Baseline demographic data, clinical characteristics and laboratory findings in rheumatoid arthritis (RA) patients receiving therapy with adalimumab or etanercept*

	Adalimumab-treated patients (n=36)	Etanercept-treated patients (n=34)	p Value
Mean age, years	52.9±15.0	58.0±5.6	0.012
Female (%)	32 (88.9%)	30 (88.2%)	1.000
Disease duration, years	5.44±2.42	5.37±2.37	0.638
RF positivity (%)	30 (83.3%)	27 (79.4%)	0.909
Anti-CCP positivity (%)	28 (77.8%)	25 (73.5%)	0.892
ESR (mm/1st hour)	48.0±20.9	49.5±25.0	0.939
DAS-28	6.08±0.81	5.95±0.54	0.445
Daily steroid dose (mg)	5.8±2.2	6.1±2.0	0.579
DMARDs at baseline			
Methotrexate	32 (88.9%)	30 (88.2%)	1.000
Sulfasalazine	30 (83.3%)	27 (79.4%)	0.763
Hydroxychloroquine	29 (80.6%)	25 (73.5%)	0.574
DMARDs combined with biologic			
Methotrexate	32 (88.9%)	30 (88.2%)	1.000
Sulfasalazine	8 (22.2%)	6 (17.6%)	0.768
Hydroxychloroquine	7 (19.4%)	6 (17.6%)	1.000

Mann-Whitney U test was used for between-group comparison of numerical variables. The χ^2 test with Yates's continuity correction or Fisher's exact test was used to compare binary variables.

*Data are presented as mean±SD or number (percentage).

Anti-CCP, anticyclic citrullinated peptide antibodies; DAS28, disease activity score for 28 joints; DMARDs, disease-modifying antirheumatic drugs; ESR, erythrocyte sedimentation rate; RF, rheumatoid factor.

0.58–9.87 µg/mL, respectively) compared with those with positive ADAb (median=0.51 µg/mL, IQR 0.10–0.95 µg/mL and median=0.35 µg/mL, IQR 0.02–0.68 µg/mL, both $p<0.001$, respectively; figure 2A,B). Similarly, patients with negative ADAb (RIA) had significantly higher drug levels at the 12th month (median=3.90 µg/mL, IQR 0.80–6.60 µg/mL) compared with those with positive ADAb (median=0.00 µg/mL, IQR 0.00–0.07 µg/mL, $p<0.001$; figure 2C). Moreover, there was an inverse correlation between ADAb levels (bridging ELISA) and drug trough levels (Progenika) ($r=-0.667$, $p<0.001$; $r=-0.575$, $p<0.001$, at the 6th and 12th month, respectively; figure 2D,E). Similarly, there was an inverse correlation between ADAb levels (RIA) and drug trough levels (Sanquin) at the 12 months of anti-TNF- α therapy ($r=-0.612$, $p<0.001$; figure 2F). In the present study, there was a significant and high

correlation between two different ELISA methods (Progenika and Sanquin) for detecting adalimumab trough levels ($r=0.875$, $p<0.001$) and etanercept trough levels ($r=0.703$, $p<0.001$).

Serum drug trough levels and their relation to EULAR response

As illustrated in table 2 and figure 3A,B, serum adalimumab trough levels were significantly higher in good EULAR responders than in moderate or poor EULAR responders. Similarly, serum etanercept trough levels were significantly higher in good EULAR responders than in moderate or poor EULAR responders. There was a positive correlation between drug levels and Δ DAS28 in both adalimumab-treated and etanercept-treated patients (figure 3C–F). In five etanercept-treated patients who had detectable drug levels and a good EULAR response assessed

Table 2 Antidrug antibodies (ADAb) against adalimumab or etanercept and serum drug trough levels in rheumatoid arthritis patients receiving 6 months and 12 months of anti-tumour necrosis factor- α therapy according to therapeutic responses

	Good EULAR response		Moderate EULAR response		Poor EULAR response	
	6th month	12th month	6th month	12th month	6th month	12th month
Ada.-Rx (n=36)	24 (66.6%)	20 (55.5%)	6 (16.7%)	6 (16.7%)	6 (16.7%)	10 (27.8%)
Eta.-Rx (n=34)	26 (76.4%)	21 (61.7%)	4 (11.8%)	6 (17.7%)	4 (11.8%)	7 (20.6%)
ADAb(+),† bridging ELISA						
Eta.-Rx	0 (0.00%)	0 (0.00%)	0 (0.00%)	0 (0.00%)	0 (0.00%)	0 (0.00%)
Ada.-Rx	1 (12.5%)#	1 (10.0%)#	1 (12.5%)	2 (20.0%)	6 (75.0%)	7 (70.0%)
ADAb(+),† RIA						
Eta.-Rx	ND	0 (0.00%)	ND	0 (0.00%)	ND	0 (0.00%)
Ada.-Rx	ND	2 (15.4%)*	ND	4 (30.8%)	ND	7 (53.8%)
Ada level(µg/ml)	6.5 (3.0–11.5)***##	5.9 (1.4–9.9)***	1.0 (0.9–1.0)#	0.5 (0.4–0.7)	0.4 (0.02–0.6)	0.3 (0.02–0.6)
Eta. Level (µg/mL)	2.3 (1.4–3.3)***##	1.3 (0.4–2.8)#	1.0 (0.7–1.2)	0.5 (0.1–1.0)	0.2 (0.07–0.6)	0.1 (0.05–0.3)

Data are presented as number (%) or median (25th–75th centile).

*** $p<0.001$, ** $p<0.01$, * $p<0.05$, versus moderate EULAR responders; ## $p<0.001$, # $p<0.01$, versus poor EULAR responders, determined by Mann-Whitney U test.

†Data are presented as number (the proportion of ADAb-positive patients).

Ada., adalimumab; Eta., etanercept. EULAR, European League Against Rheumatism; Good EULAR responders are defined as patients who have a decrease in DAS28 from baseline (Δ DAS28) >1.2 and a DAS28 ≤ 3.2 ; moderate responders have either Δ DAS28 >1.2 and a DAS28 >3.2 or Δ DAS28 0.6–1.2 and a DAS28 ≤ 5.1 ; and poor responders are those who have either Δ DAS28 <0.6 or a DAS28 >5.1 ; ND, not done RIA, radioimmunoassay; Rx, treatment.

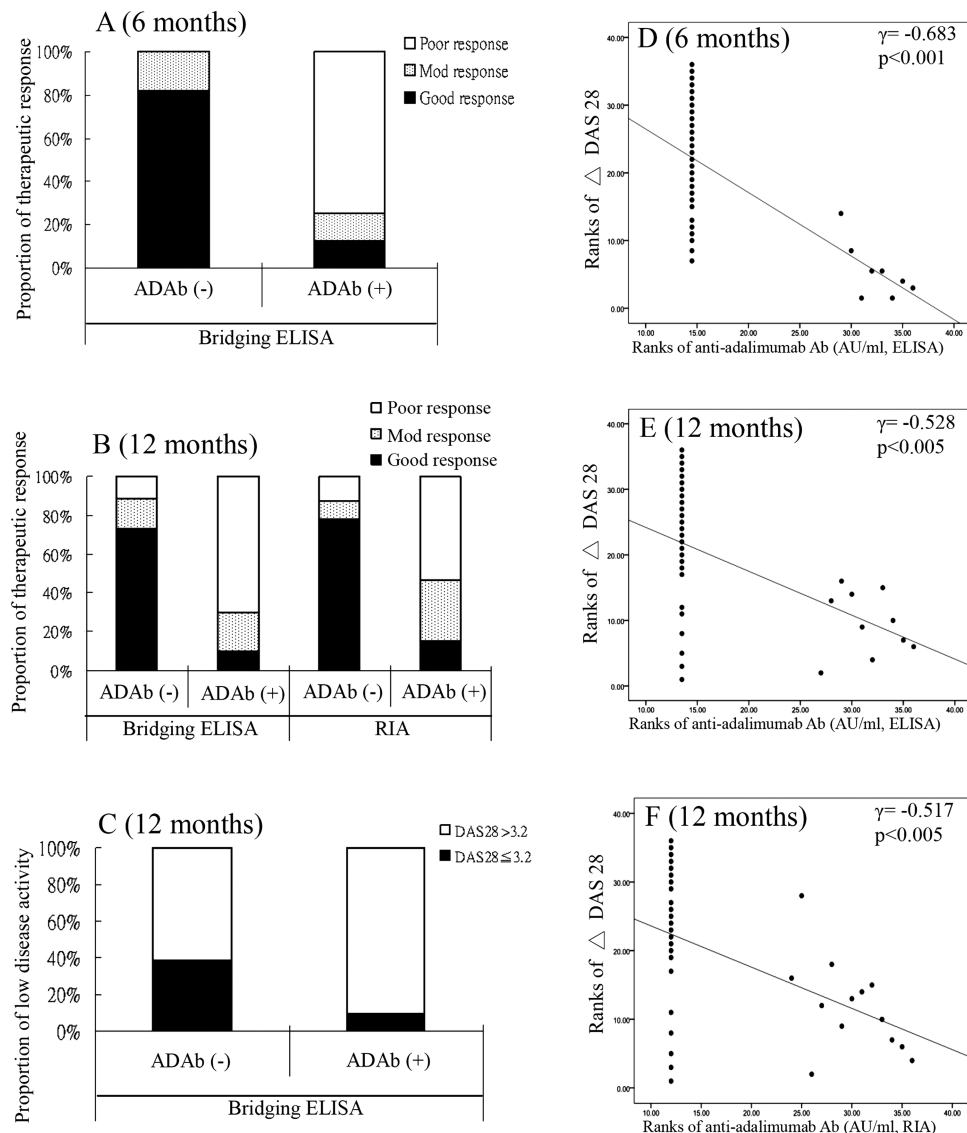


Figure 1 The association of antidrug antibody (ADAb) levels with EULAR response assessed at the 6th month (A) and at 12th month (B) and with the proportion of achieved low-disease activity at the 12th month (C) was determined by Fisher's exact test. The correlation between ADAb levels and therapeutic response (Δ DAS28) assessed at the 6th month (D) and 12th month (E and F) was obtained by the Spearman rank correlation test. EULAR, European League Against Rheumatism; good EULAR responders are defined as patients who have a decrease in DAS28 from baseline (Δ DAS28) > 1.2 and a DAS28 ≤ 3.2 ; moderate responders have either Δ DAS28 $\Delta 1.2$ and a DAS28 > 3.2 or Δ DAS28 $0.6-1.2$ and a DAS28 ≤ 5.1 ; and poor responders are those who have either Δ DAS28 < 0.6 or a DAS28 > 5.1 . Low-disease activity was defined as DAS28 ≤ 3.2 ; RIA, radioimmunoassay.

at the 6th month, their drug levels significantly declined with a loss of initial response after an extended interval of therapy at a dose of 25 mg once weekly or 25 mg every other week. In three adalimumab-treated patients who had detectable drug levels and a good EULAR response assessed at the 6th month, their drug levels declined with a loss of initial response after an interval extension to adalimumab once monthly, which was confirmed by asking these patients.

Optimal drug cut-off levels for a good EULAR response using ROC curve analysis

With ELISA (Progenika) for detecting drug trough levels, the optimal adalimumab cut-off level for good EULAR responders at 6 months was 1.274 μ g/mL with high sensitivity (90%) and specificity (100%), and the optimal etanercept cut-off level was 1.242 μ g/mL with high sensitivity (80.8%) and specificity (100%) (figure 3G,H). The optimal adalimumab cut-off level for good EULAR responders at 12 months was 1.046 μ g/mL

with high sensitivity (100%) and specificity (100%), and the etanercept cut-off level was 0.800 μ g/mL with sensitivity (85.7%) and specificity (84.6%) (figure 3I,J). With ELISA (Sanquin) for detecting serum drug levels at 12 months of therapy, the optimal adalimumab cut-off level was 0.801 μ g/mL with high sensitivity (95%) and specificity (100%) for good EULAR responders, and the etanercept cut-off level was 0.700 μ g/mL with high sensitivity (84%) and specificity (100%) for good EULAR responders.

DISCUSSION

In the present study, ADAb was detected by bridging ELISA in 8 (22.2%) and 10 (27.8%) of adalimumab-treated patients at 6 and 12 months of therapy, respectively, and in 13 (36.1%) at 12 months of therapy using RIA. However, ADAb was not detected in any of etanercept-treated patients. Patients with positive ADAb had significantly lower rates of a good EULAR response as well as achieving low-disease activity, and had lower

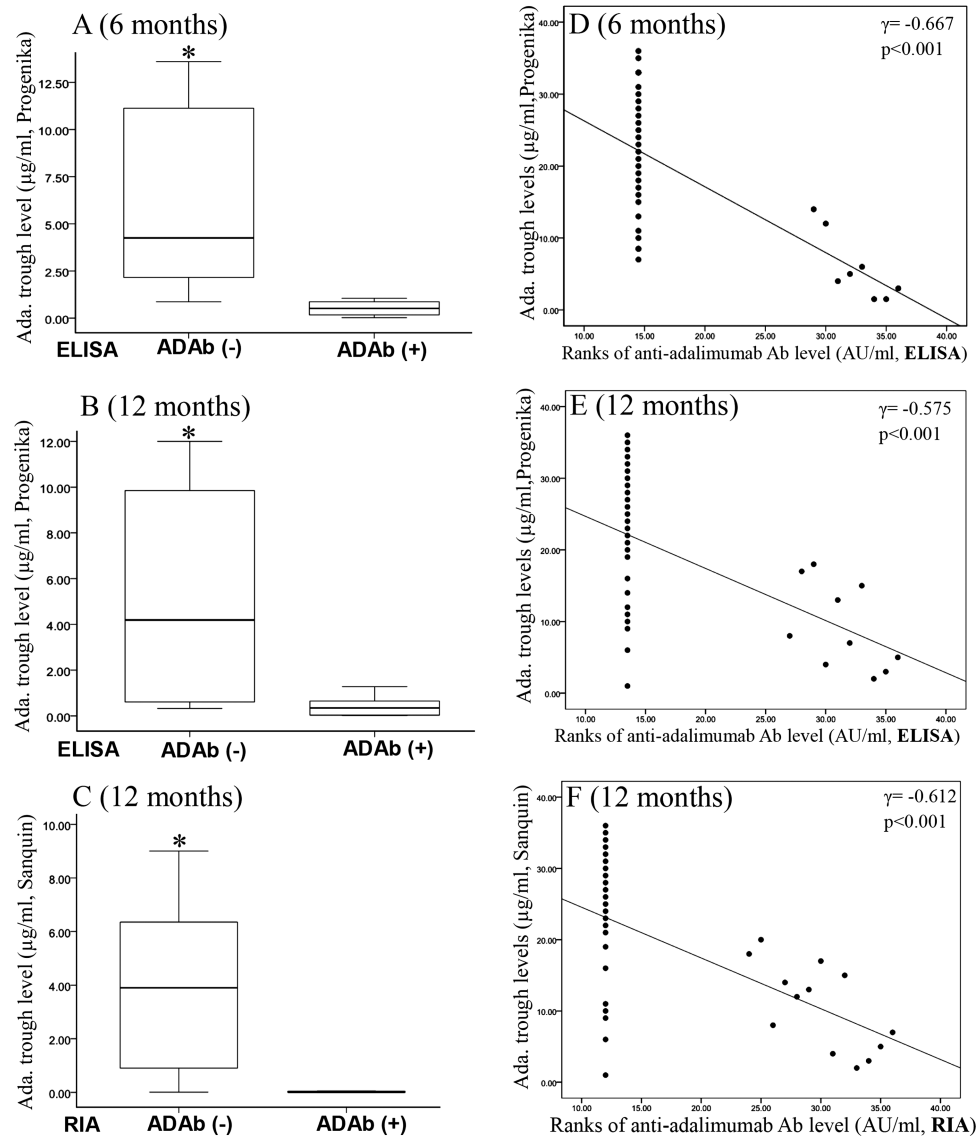


Figure 2 Comparison of drug trough levels between adalimumab-treated patients with negative ADAb (ADAb (-)) and positive ADAb (ADAb (+)) examined at the 6th month (A) and 12th month (B, C). The data are presented as box-plot diagrams, with the box encompassing the 25th centile (lower bar) to the 75th centile (upper bar). The horizontal line within the box represent the maximum and minimum values respectively for each group. *p<0.001, vs patients with ADAb (+), determined by the Mann-Whitney U test. The correlation between ADAb levels and serum drug trough levels examined at the 6th month (D) and 12th month (E and F) was determined by the Spearman rank correlation test. Ada., adalimumab.

drug trough levels than those with negative ADAb. Serum drug trough levels were positively associated with therapeutic responses in patients receiving anti-TNF- α therapy. With ELISA (Progenika) for detecting drug levels, the optimal adalimumab cut-off level for a good EULAR response assessed at 6 and 12 months of therapy was 1.274 µg/mL and 1.046 µg/mL, respectively, and that for etanercept were 1.242 µg/mL and 0.800 µg/mL, respectively. With ELISA (Sanquin) for detecting drug levels at 12 months of therapy, the optimal adalimumab cut-off level was 0.801 µg/mL and the etanercept cut-off level was 0.700 µg/mL for a good EULAR response. Although there was a difference in cut-off level between the two different ELISA methods used for drug levels, a good correlation between drug trough levels assessed by both methods was observed.

Although various rates of ADAb positivity have been reported in adalimumab-treated patients,^{9 14 18 23 32 33} the proportion of ADAb-positive patients in this study was consistent with the results of some other studies.^{9 14 33} The variability reported in previous

studies might be related to the methods used. Although there is a good concordance between the two different assays used in this study, ADAb levels in three patients were detected by RIA but not by bridging ELISA, one of whom even had high ADAb level (180 AU/mL). The possible cause might be that this ADAb was IgG4 isotype, which could only be verified by IgG4-specific ABT³⁰ but not assessed in our study. These observations support the proposition that RIA seems to be a more precise assay than bridging ELISA in detecting ADAb.^{9 13 14 34} Besides, MTX dosages were inversely correlated with ADAb levels in our patients, as also demonstrated by other studies in which MTX reduced immunogenicity in a dose-dependent manner.^{32 35}

In contrast to the high proportion of adalimumab-treated patients developing ADAb, anti-etanercept antibodies were not detected in any of our etanercept-treated patients. This finding is consistent with previous studies that anti-etanercept antibodies were measured in less than 5% of RA patients³⁶⁻³⁹ and supports the findings of a higher drug survival for etanercept

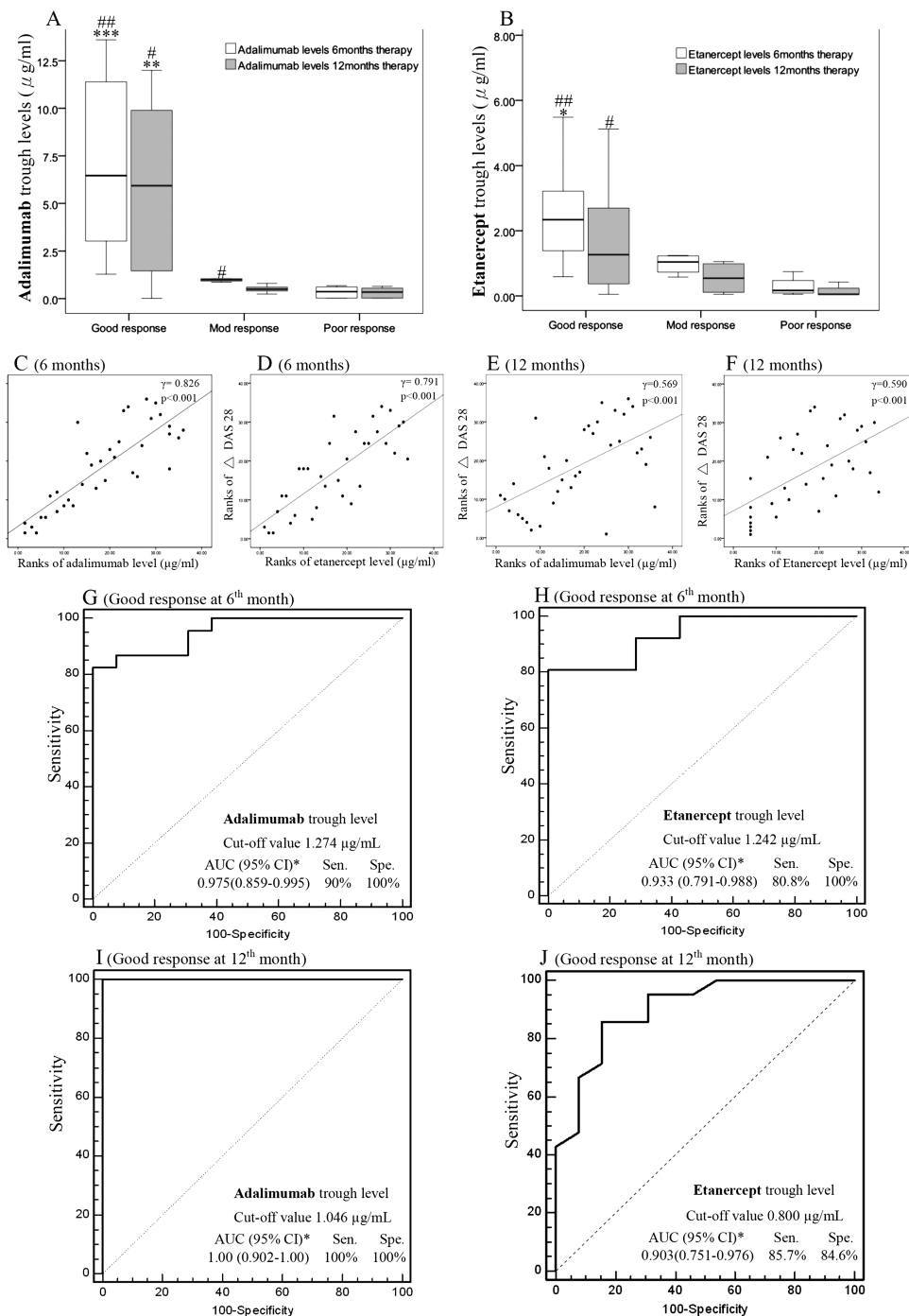


Figure 3 Comparison of serum drug trough levels among different European League Against Rheumatism (EULAR) responses assessed at the 6th and 12th month in adalimumab-treated patients (A) and in etanercept-treated patients (B). The data are presented as box-plot diagrams, with the box encompassing the 25th centile (lower bar) to the 75th centile (upper bar). The horizontal line within the box indicates median value, and the horizontal lines above and below the box represent the maximum and minimum values respectively for each group. *** $p < 0.001$, ** $p < 0.01$, * $p < 0.05$, vs moderate EULAR responders; ### $p < 0.001$, # $p < 0.01$, vs poor EULAR responders, determined by the Mann–Whitney U test. The correlation between serum drug levels and therapeutic response (Δ DAS28) assessed at the 6th and 12th month in adalimumab-treated patients (C and E, respectively) and in etanercept-treated patients (D and F, respectively) was determined by the Spearman rank correlation test. Receiver-operating characteristic (ROC) curves analysis for the determination of the optimal drug cut-off levels for good EULAR response assessed at the 6th month (G and H) and 12th month (I and J) in adalimumab-treated patients and etanercept-treated patients, respectively. p value was determined by the χ^2 test with Yate's correction of contingency. AUC, area under ROC curve; sen., sensitivity; spe., specificity.

than for adalimumab.³⁹ The disparity in the rates of ADAb induction between both drugs may be explained by their differences in pharmacokinetics, chemical structures and the stability of the TNF- α /anti-TNF- α complex.^{40–41}

Consistent with previous studies,^{9 14 18 42} the presence of ADAb was associated with a reduced therapeutic response assessed at the 6th and 12th month in our patients. When compared with adalimumab-treated patients with negative ADAb,

those with positive ADA_b had significantly higher rates of a poor EULAR response and lower rates of achieving low-disease activity ($\text{DAS28} \leq 3.2$) (figure 1A–C). Moreover, our results showed an inverse correlation between ADA_b levels and improvement of disease activity (ΔDAS28). The mechanisms through which the emergence of ADA_b hampers therapeutic response may include the formation of immune complexes leading to acceleration of drug clearance^{10 43} or functional neutralisation of the drug through blockage of its binding to the target.¹⁰

As found in recent reports,^{9 14 23 44} our patients with ADA_b had significantly lower adalimumab trough levels compared with those without ADA_b. We also demonstrated an inverse correlation between ADA_b levels and drug trough levels (figure 2). Five ADA_b-negative patients have low/undetectable drug levels assessed at 12 months of therapy. Possible contributing factors include therapeutic noncompliance in three patients and immune complexes formed by drug/ ADA_bs not detected by current assays. Based on the data of previous reports,^{22 34} a bridging ELISA used for detecting ADA_b is also susceptible to drug interference and only measures ADA_b in the absence of detectable drug levels.

Despite the absence of ADA_b in any etanercept-treated patient, there remained a substantial proportion (20.6%, 7 of 34) of patients for whom etanercept failed to have therapeutic response, as assessed at 12 months of therapy. Among them, five patients initially had adequate drug levels and good response assessed at 6 months of therapy, reflecting the poor therapeutic compliance that was confirmed by asking the patients. In this study, we also observed that three adalimumab-treated patients had therapeutic noncompliance. Therefore, if patients have low or undetectable drug levels in the absence of ADA_b, therapeutic noncompliance should be considered. A higher proportion of therapeutic noncompliance seen in etanercept-treated patients (14.7%) than in adalimumab-treated patients (8.3%) might explain our findings that there were no significant differences in EULAR response assessed at the 12th month between adalimumab-treated patients and etanercept-treated patients in spite of the negligible immunogenicity of etanercept.

In agreement with the results of recent studies,^{13–16 23–25 33 45} drug trough levels in both adalimumab-treated and etanercept-treated patients in our study were positively associated with therapeutic response and DAS28 decrement (figure 3A–F).

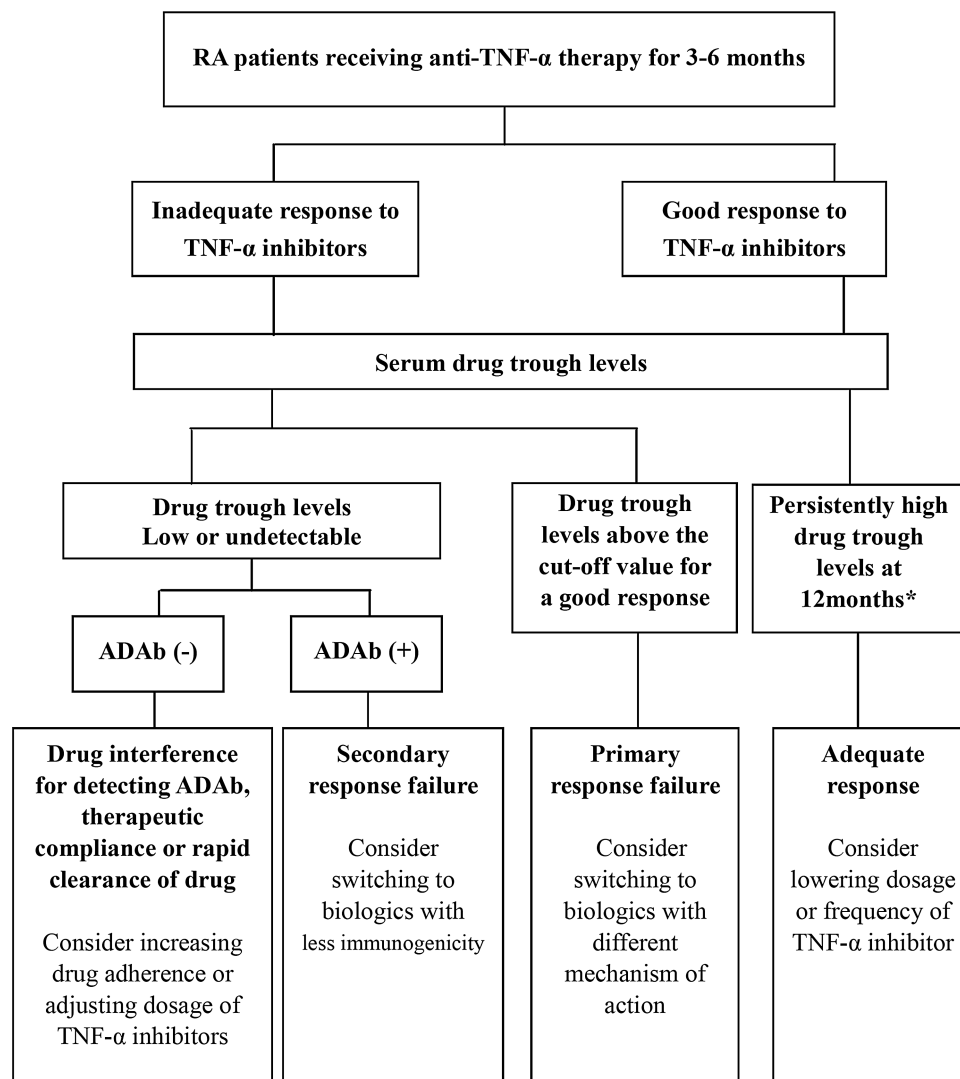


Figure 4 Algorithm for therapeutic strategies based on serum drug trough levels and antidrug antibodies (ADAb) levels in rheumatoid arthritis (RA) patients who have different therapeutic responses to tumour necrosis factor (TNF)- α inhibitor. The cut-off drug trough level for a good EULAR response was determined by receiver-operating characteristic curve analysis. *The optimal drug trough levels that call for dose reduction in patients with a good response to TNF- α inhibitors will be determined in the future studies. EULAR, European League Against Rheumatism.

Therefore, a search for the optimal cut-off level for TNF- α inhibitors is important. Using ROC analysis, we demonstrated patients with an adalimumab trough level above 1.274 $\mu\text{g/mL}$ or an etanercept level above 1.242 $\mu\text{g/mL}$ cut-off value would have a good EULAR response as assessed at 6 months of therapy. Although the data suggest that drug trough levels above the cut-off value may predict a good EULAR response in our patients, it is too early to recommend this for clinical practice until the external validity of those cut-off values can be confirmed in other cohorts. The etanercept cut-off level for predicting clinical response in our study was different from that of prior studies,²⁵ possibly due to the differences in detection methods or patient characteristics such as baseline DAS28, a high proportion of therapeutic noncompliance in our study and genetics associated with varied drug metabolism.

Based on the findings of previous studies,^{9 10 14–16 23–25} meta-analyses as well as recommendations^{18 32 44 46–48} and our results, we recommend an algorithm (figure 4) with an emphasis on therapeutic strategies based on drug trough levels and ADAb positivity for RA patients with different responses to anti-TNF- α therapy. Also, recent reports^{24 25} have proposed that determination of the drug trough level may serve as the front-line evaluation and help to predict therapeutic response. Among patients with an inadequate response to anti-TNF- α therapy, those with drug trough levels above the cut-off value for a good EULAR response and an absence of ADA are more likely to have primary response failure and may benefit from biologics with different mechanisms of action.^{48–50} For patients with low/undetectable drug levels and positive ADAb, secondary response failure should be suspected, and switching to biologics with less immunogenicity would be more appropriate.^{16 18 32 46–48} Patients who have low/undetectable drug levels and negative ADAb, drug interference for detecting ADAb, therapeutic non-compliance or rapid clearance of drug should be considered. For those who have an initial good response, but a low/undetectable drug level in the absence of ADAb at 12 months, increasing compliance might be cost effective. Some patients who have inadequate therapeutic responses and undetectable drug levels in the absence of ADAb may benefit from adjusting the dosage of TNF- α inhibitor.^{45 51} However, the merit of increasing the dosage of TNF- α inhibitor requires more evaluation.^{5 14 36} Among our patients who had a good EULAR response, 18.4% of adalimumab-treated patients and 14.7% of etanercept-treated patients had persistently high drug levels above the 75th centile of the detectable drug levels at 12 months. Those patients may be eligible for dose reduction, considering the high costs and dose-dependent adverse effects of biologics.^{48 52}

Some limitations in this study should be addressed. The determination of ADAb may be confounded by the circulating drug levels. In this study, we obtained blood samples immediately before drug injection to reduce drug interference. Although the duration of anti-TNF- α therapy may be related to the emergence of ADAb,¹⁴ our results were compatible with the positive rate of ADAb in a long-term study of 272 adalimumab-treated patients.¹⁴ The sample size may be too small to draw a definitive conclusion regarding the optimal cut-off level for predicting a good therapeutic response. In addition, there is no specific design in this study to define the high drug levels that call for dose reduction in patients with a good response to TNF- α inhibitors. Therefore, further prospective and longitudinal investigation in a large number of subjects is needed.

In conclusion, a positive correlation was shown between drug trough levels and improvement of disease activity in patients

receiving anti-TNF- α therapy, indicating that monitoring of drug trough levels would be useful for evaluation of therapeutic response. We also demonstrated an inverse correlation between ADAb levels and drug levels or therapeutic response in adalimumab-treated patients, suggesting the beneficial role of ADAb level in making therapeutic decisions for those with low or undetectable drug levels. The data combining drug levels and ADAb positivity would be a useful guide for physicians to optimise dose regimens and prevent prolonged use of inadequate therapy or overtreatment for those receiving anti-TNF- α therapy.^{46–50}

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Contributors All authors made substantive intellectual contributions to the present study and approved the final manuscript. D-YC conceived of the study, generated the original hypothesis, designed the study, acquired clinical data, analysed data, drafted and revised the manuscript. Y-MC, W-CT and J-CT contributed equally on this work, conceived of the study, generated the original hypothesis and designed the study. Y-HC, C-WH, W-TH and J-LL performed clinical assessments and data acquisition, statistical analysis and revised the manuscript.

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REFERENCES

- Christodoulou C, Choy EH. Joint inflammation and cytokine inhibition in rheumatoid arthritis. *Clin Exp Med* 2006;6:13–19.
- Choy EH, Panayi GS. Cytokine pathways and joint inflammation in rheumatoid arthritis. *N Engl J Med* 2001;344:907–16.
- Feldmann M, Maini RN. TNF defined as a therapeutic target for rheumatoid arthritis and other autoimmune diseases. *Nat Med* 2003;9:1245–50.
- van der Heijde D, Klareskog L, Rodriguez-Valverde V, *et al*. TEMPO Study Investigators: Comparison of etanercept and methotrexate, alone and combined, in the treatment of rheumatoid arthritis: Two-year clinical and radiographic results from the TEMPO study, a double-blind, randomized trial. *Arthritis Rheum* 2006;54:1063–74.
- Breedveld FC, Weisman MH, Kavanaugh AF, *et al*. The PREMIER study: a multicenter, randomized, double-blind clinical trial of combination therapy with adalimumab plus methotrexate versus methotrexate alone or adalimumab alone in patients with early, aggressive rheumatoid arthritis who had not had previous methotrexate treatment. *Arthritis Rheum* 2006;54:26–37.
- Kievit W, Fransen J, Adang EMM, *et al*. Long-term effectiveness and safety of TNF-blocking agents in daily clinical practice: results from the Dutch rheumatoid arthritis monitoring register. *Rheumatology (Oxford)* 2011;50:196–203.
- Rubbert-Roth A, Finckh A. Treatment options in patients with rheumatoid arthritis failing initial TNF inhibitor therapy: a critical review. *Arthritis Res Ther* 2009;11 (Suppl 1):S1.
- Alzabin S, Abraham SM, Taher TE, *et al*. Incomplete response of inflammatory arthritis to TNF α blockade is associated with the Th17 pathway. *Ann Rheum Dis* 2012;71:1741–8.
- Radstake TR, Svenson M, Eijsbouts AM, *et al*. Formation of antibodies against infliximab and adalimumab strongly correlates with functional drug levels and clinical response in rheumatoid arthritis. *Ann Rheum Dis* 2009;68:1739–45.

- 10 van Schouwenburg PA, van de Stadt LA, de Jong RN, *et al.* Adalimumab elicits a restricted anti-idiotypic antibody response in autoimmune patients resulting in functional neutralization. *Ann Rheum Dis* 2013;72:104–9.
- 11 Schellekens H. Immunogenicity of therapeutic proteins. *Nephrol Dial Transplant* 2003;18:1257–9.
- 12 Anderson P, Louie J, Lau A, *et al.* Mechanisms of differential immunogenicity of tumor necrosis factor inhibitors. *Curr Rheumatol Rep* 2005;7:3–9.
- 13 Svenson BM, Geborek P, Saxne T, *et al.* Monitoring patients treated with anti-TNF- α biopharmaceuticals: assessing serum infliximab and anti-infliximab antibodies. *Rheumatology (Oxford)* 2007;46:1828–34.
- 14 Bartelds GM, Kriekkaert CL, Nurmohamed MT, *et al.* Development of antidrug antibodies against adalimumab and association with disease activity and treatment failure during long-term follow-up. *JAMA* 2011;305:1460–68.
- 15 Hansen KE, Hildebrand JP, Genovese MC, *et al.* The efficacy of switching from etanercept to infliximab in patients with rheumatoid arthritis. *J Rheumatol* 2004;31:1098–102.
- 16 Jamnitski A, Bartelds GM, Nurmohamed MT, *et al.* The presence or absence of antibodies to infliximab or adalimumab determines the outcome of switching to etanercept. *Ann Rheum Dis* 2011;70:284–8.
- 17 Kriekkaert CL, Bartelds GM, Lems WF, *et al.* The effect of immunomodulators on the immunogenicity of TNF-blocking therapeutic monoclonal antibodies: a review. *Arthritis Res Ther* 2010;12:217.
- 18 Garcés S, Demengeot J, Benito-García E. The immunogenicity of anti-TNF therapy in immune-mediated inflammatory diseases: a systematic review of the literature with a meta-analysis. *Ann Rheum Dis* 2013;72:1947–55.
- 19 Aarden L, Ruuls SR, Wolbink GJ. Immunogenicity of anti-tumor necrosis factor antibodies: toward improved methods of anti-antibody measurement. *Curr Opin Immunol* 2008;20:431–5.
- 20 Wolbink GJ, Aarden LA, Dijkmans BA. Dealing with immunogenicity of biological: assessment and clinical relevance. *Curr Opin Rheumatol* 2009;21:211–15.
- 21 van Schouwenburg PA, Bartelds GM, Hart MH, *et al.* A novel method for the detection of antibodies to adalimumab in the presence of drug reveals “hidden” immunogenicity in rheumatoid arthritis patients. *J Immunol Methods* 2010;362:82–8.
- 22 Pascual-Salcedo D, Plasencia C, Ramiro S, *et al.* Influence of immunogenicity on the efficacy of long-term treatment with infliximab in rheumatoid arthritis. *Rheumatology (Oxford)* 2011;50:1445–52.
- 23 Bartelds GM, Wijbrandts CA, Nurmohamed MT, *et al.* Clinical response to adalimumab: relationship to anti-adalimumab antibodies and serum adalimumab concentrations in rheumatoid arthritis. *Ann Rheum Dis* 2007;66:921–6.
- 24 Daien CI, Daien V, Parussini E, *et al.* Etanercept concentration in patients with rheumatoid arthritis and its potential influence on treatment decisions: a pilot study. *J Rheumatol* 2012;39:1533–8.
- 25 Jamnitski A, Kriekkaert CL, Nurmohamed MT, *et al.* Patients non-responding to etanercept obtain lower etanercept concentrations compared with responding patients. *Ann Rheum Dis* 2012;71:88–91.
- 26 Arnett FC, Edworthy SM, Bloch DA, *et al.* The American Rheumatism Association 1987 revised criteria for the classification of rheumatoid arthritis. *Arthritis Rheum* 1988;31:315–24.
- 27 Ledingham J, Deighton C: British Society for Rheumatology Standards, Guidelines and Audit Working Group. Update on the British Society for Rheumatology guidelines for prescribing TNF α blockers in adults with rheumatoid arthritis (update of previous guidelines of April 2001). *Rheumatology (Oxford)* 2005;44:157–63.
- 28 Prevoo MLL, van’t Hof MA, Kuper HH, *et al.* Modified disease activity scores that include twenty-eight-joint counts: development and validation in a prospective longitudinal study of patients with rheumatoid arthritis. *Arthritis Rheum* 1995;38:44–8.
- 29 Van Gestel AM, Prevoo ML, van’t Hof MA, *et al.* Development and validation of the European League Against Rheumatism response criteria for rheumatoid arthritis. Comparison with the preliminary American College of Rheumatology and the World Health Organization/International League Against Rheumatism Criteria. *Arthritis Rheum* 1996;39:34–40.
- 30 Hart MH, de Vrieze H, Wouters D, *et al.* Differential effect of drug interference in immunogenicity assays. *J Immunol Methods* 2011;372:196–203.
- 31 de Vries MK, van der Horst-Bruinsma IE, Nurmohamed MT, *et al.* Immunogenicity does not influence treatment with etanercept in patients with ankylosing spondylitis. *Ann Rheum Dis* 2009;68:531–5.
- 32 Vincent FB, Morand EF, Murphy K, *et al.* Antidrug antibodies (ADAb) to tumor necrosis factor (TNF)-specific neutralizing agents in chronic inflammatory diseases: a real issue, a clinical perspective. *Ann Rheum Dis* 2013;72:165–78.
- 33 Bartelds GM, Wijbrandts CA, Nurmohamed MT, *et al.* Anti-infliximab and anti-adalimumab antibodies in relation to response to adalimumab in infliximab switchers and anti-TNF naive patients: a cohort study. *Ann Rheum Dis* 2010;69:817–21.
- 34 Francisca LT, de Salazar JRG, Gallego JMS, *et al.* Analytical and clinical evaluation of a new immunoassay for therapeutic drug monitoring of infliximab and adalimumab. *Clin Chem Lab Med* 2012;50:1845–7.
- 35 Kriekkaert CL, Nurmohamed MT, Wolbink GJ. Methotrexate reduces immunogenicity in adalimumab treated rheumatoid arthritis patients in a dose dependent manner. *Ann Rheum Dis* 2012;71:1914–15.
- 36 Weinblatt ME, Schiff MH, Ruderman EM, *et al.* Efficacy and safety of etanercept 50 mg twice a week in patients with rheumatoid arthritis who had a suboptimal response to etanercept 50 mg once a week: results of a multicenter, randomized, double-blind, active drug–controlled study. *Arthritis Rheum* 2008;58:1921–30.
- 37 Hoshino M, Yoshio T, Onishi S, *et al.* Influence of antibodies against infliximab and etanercept on the treatment effectiveness of these agents in Japanese patients with rheumatoid arthritis. *Mod Rheumatol* 2011;22:532–40.
- 38 Klareskog L, Gaubitz M, Rodriguez-Valverde V, *et al.* Assessment of long-term safety and efficacy of etanercept in a 5-year extension study in patients with rheumatoid arthritis. *Clin Exp Rheumatol* 2011;29:238–47.
- 39 Hetland ML, Christensen IJ, Tarp U, *et al.* Direct comparison of treatment responses, remission rates, and drug adherence in patients with rheumatoid arthritis treated with adalimumab, etanercept, or infliximab: results from eight years of surveillance of clinical practice in the nationwide Danish DANBIO registry. *Arthritis Rheum* 2010;62:22–32.
- 40 Lobo ED, Hansen RJ, Balthasar JP. Antibody pharmacokinetics and pharmacodynamics. *J Pharm Sci* 2004;93:2645–68.
- 41 Tracey D, Klareskog L, Sasso EH, *et al.* Tumor necrosis factor antagonist mechanisms of action: a comprehensive review. *Pharmacol Ther* 2008;117:244–79.
- 42 Kriekkaert CL, Jamnitski A, Nurmohamed MT, *et al.* Comparison of long-term clinical outcome with etanercept treatment and adalimumab treatment of rheumatoid arthritis with respect to immunogenicity. *Arthritis Rheum* 2012;64:3850–5.
- 43 van der Laken CJ, Voskuyl AE, Roos JC, *et al.* Imaging and serum analysis of immune complex formation of radiolabelled infliximab and anti-infliximab in responders and non-responders to therapy for rheumatoid arthritis. *Ann Rheum Dis* 2007;66:253–6.
- 44 van schouwenburg PA, Rispens T, Wolbink GJ. Immunogenicity of anti-TNF biologic therapies for rheumatoid arthritis. *Nat Rev Rheumatol* 2013;9:164–72.
- 45 Mulleman D, Chu Miow Lin D, Ducourau E, *et al.* Trough infliximab concentrations predict efficacy and sustained control of disease activity in rheumatoid arthritis. *Ther Drug Monit* 2010;32:232–6.
- 46 Emi Aikawa N, de Carvalho JF, Artur Almeida Silva C, *et al.* Immunogenicity of Anti-TNF- α agents in autoimmune diseases. *Clin Rev Allergy Immunol* 2010;38:82–9.
- 47 Bendtzen K, Geborek P, Svenson M, *et al.* Individualized monitoring of drug bioavailability and immunogenicity in rheumatoid arthritis patients treated with the tumor necrosis factor inhibitor Infliximab. *Arthritis Rheum* 2006;54:3782–9.
- 48 Garcés S, Antunes M, Benito-García E, *et al.* A preliminary algorithm introducing immunogenicity assessment in the management of patients with RA receiving tumor necrosis factor inhibitor therapies. *Ann Rheum Dis* 2013. Published Online First: 11 May 2013. 10.1136/annrheumdis-2013-203296
- 49 Du Pan SM, Scherer A, Gabay C, *et al.* Differential drug retention between anti-TNF agents and alternative biological agents after inadequate response to an anti-TNF agent in rheumatoid arthritis. *Ann Rheum Dis* 2012;71:997–9.
- 50 Schoels M, Aletaha D, Smolen JS, *et al.* Comparative effectiveness and safety of biological treatment opinions after tumor necrosis factor- α inhibitor failure in rheumatoid arthritis: systematic review and indirect pairwise meta-analysis. *Ann Rheum Dis* 2012;71:1303–8.
- 51 Méric JC, Mulleman D, Ducourau E, *et al.* Therapeutic drug monitoring of infliximab in spondyloarthritis: an observational open-label study. *Ther Drug Monit* 2011;33:411–16.
- 52 den Broeder AA, van der Maas A, van den Bemt B. Dose de-escalation strategies and role of therapeutic drug monitoring of biologics in RA. *Rheumatology* 2010;49:1801–3.



Significant associations of antidrug antibody levels with serum drug trough levels and therapeutic response of adalimumab and etanercept treatment in rheumatoid arthritis

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