Use of Serum Procalcitonin to Detect Bacterial Infection in Patients With Autoimmune Diseases

A Systematic Review and Meta-Analysis

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Objective. To systematically review evidence of the accuracy of the procalcitonin test for diagnosis of bacterial infection in patients with autoimmune disease.

Methods. The major databases Medline, EMBase, and the Cochrane Library were searched for studies published between January 1966 and October 2011 that evaluated procalcitonin, alone or in comparison with other laboratory markers such as C-reactive protein (CRP), as a diagnostic marker for bacterial infection in patients with autoimmune disease and provided sufficient data to permit construction of 2×2 tables.

Results. Nine studies were included in the final meta-analysis. The area under the summary receiver operating characteristic curve values were 0.91 (95% confidence interval [95% CI] 0.88–0.93) for procalcitonin and 0.81 (95% CI 0.78–0.84) for CRP. In general, testing for procalcitonin was highly specific for identifying infectious complications, although it was not as

sensitive as testing for CRP. Pooled sensitivity was 0.75 (95% CI 0.63–0.84) for procalcitonin tests and 0.77 (95% CI 0.67–0.85) for CRP tests. Pooled specificity was 0.90 (95% CI 0.85–0.93) for procalcitonin tests and 0.56 (95% CI 0.25–0.83) for CRP tests. The positive likelihood ratio for procalcitonin (7.28 [95% CI 5.10–10.38]) was sufficiently high to qualify procalcitonin testing as a rule-in diagnostic tool, while the negative likelihood ratio (0.28 [95% CI 0.18–0.40]) was not sufficiently low to qualify procalcitonin testing as a reliable rule-out diagnostic tool.

Conclusion. Procalcitonin has higher diagnostic value than CRP for the detection of bacterial sepsis in patients with autoimmune disease, and the test for procalcitonin is more specific than sensitive. A procalcitonin test is not recommended to be used in isolation as a rule-out tool.

It is critically important but can be challenging to distinguish infection from disease flare in febrile patients with autoimmune diseases. Administration of additional or higher dosages of immunosuppressive agents can ameliorate disease flare but can exacerbate infection. Disease flare and infection have similar clinical manifestations and are identified by similar laboratory markers, such as leukocytosis with or without a left shift and elevation of the serum C-reactive protein (CRP) level (1,2). Moreover, the leukocyte count and the serum CRP level can also be affected by systemic corticosteroid and immunosuppressive therapy (1,2). Thus, there is an urgent need for a reliable biomarker that provides high sensitivity and specificity for the early discrimination of infection from disease flare in febrile patients with autoimmune diseases.

Procalcitonin is a precursor of calcitonin that is

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primarily produced by parafollicular or calcitonin cells of the thyroid gland. Serum procalcitonin is normally undetectable (<0.05 ng/ml) in healthy individuals, but its level increases rapidly following bacterial infection (3). In contrast to CRP, expression of procalcitonin is not elevated following noninfectious inflammation or non-bacterial infections, so it may be useful as a marker to distinguish bacterial infection from disease flare in patients with autoimmune diseases (4–12). However, previous studies have demonstrated elevated serum levels of procalcitonin in patients who have certain autoimmune diseases (granulomatosis with polyangiitis [Wegener's] [13], adult-onset Still's disease [AOSD] [5], and Kawasaki disease during acute exacerbation [14]) but no evidence of bacterial infection.

We conducted a systematic review and metaanalysis to quantitatively summarize current evidence regarding the use of serum procalcitonin as a marker to distinguish bacterial infection from disease flare in patients with systemic autoimmune diseases.

MATERIALS AND METHODS

Systematic meta-analysis guideline adherence. We followed standard guidelines and used standard methods for systematic reviews and meta-analyses of diagnostic tests (15).

Literature search strategy. We performed a comprehensive literature search to identify studies that examined the diagnostic accuracy of procalcitonin in the evaluation of patients with autoimmune diseases, by searching the Medline, EMBase, and Cochrane Library databases for the period of time from 1966 to October 2011. "Procalcitonin" is not listed as a MeSH term, so we initially searched for "procalcitonin" as a text word with no language restriction and then combined "procalcitonin" with keywords for different autoimmune diseases. Thus, the search phrase was as follows: "Procalcitonin AND (systemic lupus erythematosus OR rheumatoid arthritis OR juvenile rheumatoid arthritis OR Felty's syndrome OR autoimmune disease OR lupus vasculitis OR multiple sclerosis OR Sjögren's syndrome OR mixed connective tissue disease OR ankylosing spondylitis OR reactive arthritis OR psoriatic arthritis OR Wegener's granulomatosis OR Churg-Strauss syndrome OR polyarteritis nodosa OR microscopic polyangitis OR microscopic polyangitis OR giant cell arteritis OR Takayasu arteritis OR Henoch-Schönlein purpura OR Behçet's disease OR relapsing polychondritis OR autoimmune thyroiditis OR Kawasaki disease OR amyloidosis OR sarcoidosis OR hereditary periodic fever syndromes OR myasthenia gravis OR Lambert-Eaton myasthenic syndrome OR primary biliary cirrhosis OR bullous pemphigoid OR autoimmune hepatitis OR primary sclerosing cholangitis OR Crohn's disease OR diabetes OR autoimmune hemolytic anemia OR idiopathic thrombopenic purpura OR Goodpasture's syndrome OR pernicious anemia OR Hashimoto thyroiditis). We identified additional

references by cross-checking the bibliographies of retrieved full-text articles.

Study selection and data extraction. We included studies that evaluated procalcitonin alone or in comparison with other laboratory markers (such as CRP) to diagnose bacterial infection in patients with autoimmune diseases and that provided sufficient data for construction of a 2 × 2 contingency table. Two of the authors (JYW and PHY) independently assessed all titles and abstracts to assure that the inclusion criteria were satisfied. Full-text articles were retrieved if any of the authors considered the abstract suitable. The 2 authors then independently examined the full text of the retrieved studies and assessed the suitability for inclusion. Discrepancies between the 2 authors were resolved by ascertaining assessments from additional authors (SSC and CCL), and the decision for inclusion was made by consensus.

The 2 original authors independently extracted data from each selected study. These included overall characteristics of the study (first author, country, language, and year of publication), characteristics of the patients (age range, clinical setting, population parameters, disease type), availability of data required for reconstruction of a 2×2 table (number of participants, sensitivity, specificity, number of cases), and characteristics of the procalcitonin test (cutoff level), study settings, definitions of outcomes, and method used to document infection (microbiologic or clinical). The quality of selected studies was assessed using the Quality Assessment of Diagnostic Accuracy Studies (QUADAS) criteria (16). We

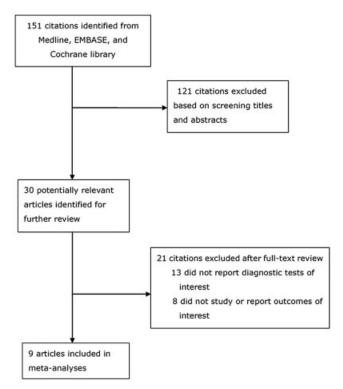


Figure 1. Procedure used to identify published studies of biomarkers in febrile patients with autoimmune diseases.

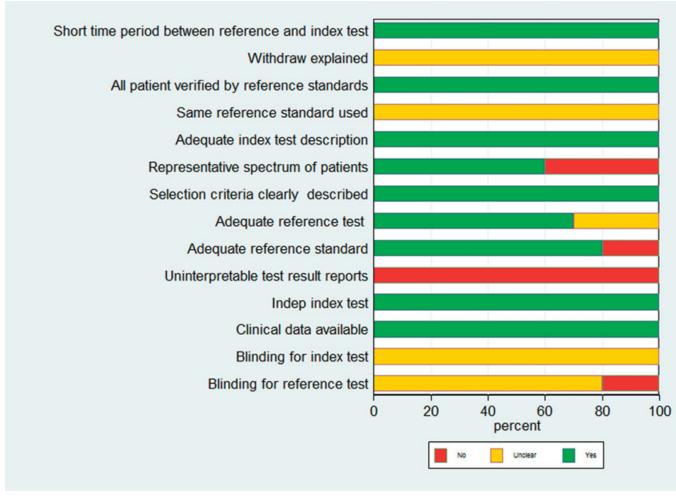


Figure 2. Quality Assessment of Diagnostic Accuracy Studies (QUADAS) criteria for the included studies.

used data with the highest Youden index ([sensitivity + specificity] - 1) for meta-analysis and sensitivity analysis (17). Data with the highest Youden index, rather than the highest sensitivity, were used.

Data preparation and statistical analysis. We used the bivariate model for diagnostic meta-analysis to obtain weighted overall estimates of the sensitivity and specificity of procalcitonin for discriminating fever related to bacterial infection from other causes of fever in patients with autoimmune disease (18). The bivariate approach assumes a bivariate distribution for the logit-transformed sensitivity and specificity. This approach accounts for study size and adjusts for the negative correlation of sensitivity and specificity of the index test that may arise from the use of different thresholds in different studies. A summary receiver operating characteristic (ROC) curve was constructed to summarize the true-positive and false-positive rates of different diagnostic studies (19). Overall sensitivity, specificity, and 95% confidence intervals (95% CIs) were calculated based on the binominal distributions of the true-positive and true-negative rates.

We also calculated I^2 statistics to quantify the extent of variation among studies (i.e., heterogeneity) (20). An I^2 value of >50% was considered an indication of significant heterogeneity. We also performed a conventional diagnostic odds ratio (OR) meta-analysis. Unadjusted data were used exclusively in all meta-analyses. We used a linear regression of log odds ratio on the inverse root of effective sample size as a test for funnel plot asymmetry in diagnostic meta-analyses. A non-zero slope is suggestive of a significantly small study bias (P < 0.05). We defined cutoff value and underlying disease as a priori clinical and design study characteristics that were potentially relevant covariates. All statistical analyses were conducted using Stata version 11.0 software, notably with the midas and metandi commands. All statistical tests were 2-sided. P values less than 0.05 were considered significant.

RESULTS

Identification of studies and study quality. Our initial search yielded 151 citations. After exclusion of

Table 1. Characteristics of the 9 included studies in which biomarkers were used to assess febrile patients with autoimmune diseases*

Author, year.		Prevalence		PCT testing	Cutoff		Severity of	PCT	T	CRP	Y.
country (ref)	Disease(s)	(no.)	Biomarker	system	value(s)†	Outcome(s)	infection	Sens.	Spec.	Sens.	Spec.
Eberhard et al, 1997, Germany (6)	Systemic auto- immune diseases	0.04 (397)	PCT	LumiTest‡	PCT ≥0.5	Microbio- logically documented infection	Systemic bacterial infection	100	84.0	NA	
Brunkhorst et al, 2000, Germany (4)	Systemic auto- immune diseases	0.30 (53)	PCT	LumiTest‡	PCT ≥0.5	Clinically documented infection, systemic	Systemic bacterial infection	100	84.0	NA A	
Martinot et al, 2005, France (7)	Acute arthritis	0.26 (42)	PCT, CRP	LumiTest‡	PCT ≥ 0.3 , CRP ≥ 50	Microbio- logically documented infectious	Generalized and local bacterial arthritis	72.7	93.5	100	40.0
Scire et al, 2006, Italy (8)	Systemic auto- immune diseases	0.45 (44)	PCT, CRP	PCT LIA (LumiTest)‡	PCT ≥ 0.5 , CRP ≥ 60	Microbio- logically documented infection	Systemic and local infection	75.0	75.0	95.0	8.30
Tamaki et al, 2008, Japan (10)	Systemic auto- immune diseases	0.29 (99)	PCT	SphereLight\$	PCT ≥0.53	Clinically and microbiologically documented infection	SIRS and non- SIRS	51.7	97.1	Y Y	
Chen et al, 2009, Taiwan (5)	Adult-onset Still's disease	0.32 (38)	PCT, CRP	VIDAS PCT assay¶	$PCT \ge 1.4,$ $CRP \ge 101$	Microbio- logically documented	Systemic and local infection	100	100	73.1	83.3
Suzuki et al, 2009, Japan (9)	Neuro- Behçet's disease	0.36 (11)	PCT	X Y	PCT ≥0.5	Microbio- logically documented bacterial CNS	Bacterial meningitis and brain abscess	0.06	0.06	NA	
Sleglova et al, 2010, Czech Republic (12)	Systemic auto- immune diseases	0.20 (212)	PCT, CRP	LumiTest‡	PCT ≥ 0.5 , CRP ≥ 20	Clinically and microbio-logically documented infection	Systemic and local infection	52.4	94.0	76.2	67.9
Joo et al, 2011, Korea (11)	Systemic auto- immune diseases	0.41 (79)	PCT, CRP	VIDAS PCT assay¶	PCT ≥0.09, CRP ≥71.8	Clinically documented infection	SIRS and non-SIRS	81.3	78.7	71.9	68.1

* Sens. = sensitivity; spec. = specificity; NA = not available; LIA = line immunoassay; SIRS = systemic inflammatory response syndrome; CNS = central nervous system. † Cutoff values for the procalcitonin (PCT) and C-reactive protein (CRP) assays were measured as ng/ml and mg/liter, respectively.

‡ Brahms Diagnostica.

§ Wako Pure Chemical Industries.

¶ bioMérieux.

Table 2. Summary of the subgroup analysis of the 9 included studies*

Publication bias, Egger's test I ² (95% CI) P value	0.0 (0.0–64.8) 0.021	0.0 (0.0–74.6) 0.049	65.9 (0.0–88.4) 0.082	56.2 (7.74–79.2) 0.401	71.2 (2.1–91.5) 0.044	68.3 (18.2–87.7) 0.788
Diagnostic OR (95% CI)	.84) 0.90 (0.85–0.93) 7.28 (5.10–10.38) 0.28 (0.18–0.40) 0.91 (0.88–0.93) 25.2 (15.4–41.5)	22.6 (13.3–38.3)	$0.81 \ (0.49 - 0.95) \ \ 0.86 \ (0.78 - 0.91) \ \ 5.83 \ (3.57 - 9.53) \ \ \ 0.22 \ (0.06 - 0.73) \ \ 0.88 \ (0.85 - 0.90) \ \ 27.0 \ (6.1 - 119.2) \ \ \ 65.9 \ (0.0 - 88.4)$.89) 0.88 (0.82–0.92) 6.19 (4.43–8.39) 0.27 (0.14–0.53) 0.90 (0.87–0.92) 25.6 (15.0–43.5) 56.2 (7.74–79.2)	$0.42\ (0.31-0.54)\ 0.95\ (0.89-0.98)\ 4.62\ (0.58-37.11)\ 0.52\ (0.38-0.72)\ 0.58\ (0.47-1.00)\ 8.85\ (0.98-79.66)\ 71.2\ (2.1-91.5)$	
AUROC (95% CI)	0.91 (0.88–0.93)	.84) $0.89 (0.82-0.94) 6.76 (4.47-10.20) 0.31 (0.20-0.48) 0.91 (0.88-0.93) 22.6 (13.3-38.3)$	0.88 (0.85-0.90)	0.90 (0.87–0.92)	0.58 (0.47–1.00)	.85) 0.56 (0.25–0.83) 1.76 (0.88–3.49) 0.41 (0.24–0.67) 0.81 (0.78–0.84) 5.06 (1.7–14.7)
Negative LR (95% CI)	0.28 (0.18–0.40)	0.31 (0.20–0.48)	0.22 (0.06–0.73)	0.27 (0.14-0.53)	0.52 (0.38–0.72)	0.41 (0.24–0.67)
Positive LR (95% CI)	7.28 (5.10–10.38)	6.76 (4.47–10.20)	5.83 (3.57–9.53)	6.19 (4.43–8.39)	4.62 (0.58–37.11)	1.76 (0.88–3.49)
Specificity (95% CI)	0.90 (0.85–0.93)	0.89 (0.82–0.94)	0.86 (0.78–0.91)	0.88 (0.82–0.92)	0.95 (0.89–0.98)	0.56 (0.25–0.83)
Sensitivity (95% CI)	0.75 (0.63–0.84)	0.73 (0.58–0.84)	0.81 (0.49–0.95)	0.76 (0.56–0.89)	0.42 (0.31–0.54)	0.77 (0.67–0.85)
No. of studies	6	9	4	7	ϵ	\$
Variable (refs.)	Procalcitonin Overall analysis (4-12)	Systemic autoimmune disease population (4,6,8,10–12)	AÒŚĎ excluded (4,6,7,9)	Cutoff 0.5 ng/ml (4–9,12)	Higher cutoff value (>0.5 ng/ml) (5,7,10) CRP	Overall analysis (5,7,8,11,12)

* 95% CI = 95% confidence interval; LR = likelihood ratio; AUROC = area under the receiver operating characteristic curve; OR = odds ratio; AOSD = adult-onset Still's disease; CRP = C-reactive protein.

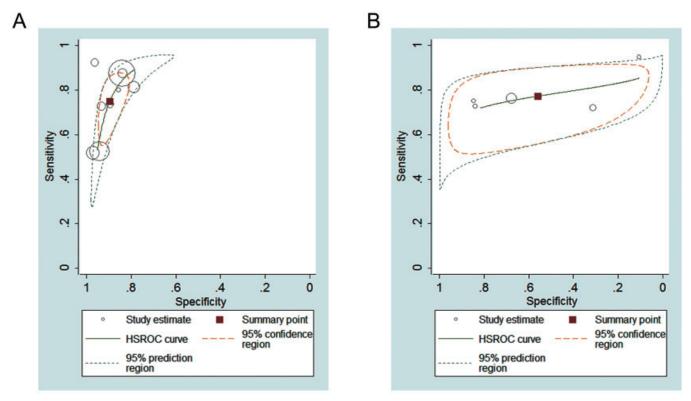


Figure 3. Hierarchical summary receiver operating characteristic (HSROC) analysis of procalcitonin (A) and C-reactive protein (B) for the detection of bacterial infection in febrile patients with autoimmune diseases.

certain publications based on review of the title and abstract, we retrieved 30 potentially relevant articles for full-text review. Twenty-one of these articles were ex-

cluded because related diagnostic tests or outcomes were not reported. A total of 9 citations were thus included in our final analysis (Figure 1). These studies

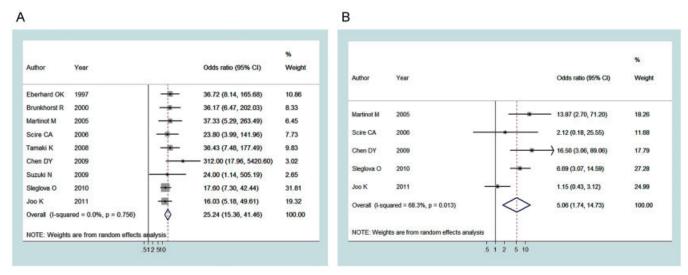


Figure 4. Forest plot of the diagnostic odds ratios of studies that used procalcitonin (A) and C-reactive protein (B) as markers for bacterial infection in febrile patients with autoimmune diseases. 95% CI = 95% confidence interval.

contained data on 975 patients in whom the procalcitonin level was measured and 330 patients in whom the CRP level was measured. The prevalence of bacterial infection was 18.6% among patients in whom procalcitonin was measured (181 of 975 patients) and 25.5% among patients in whom CRP was measured (84 of 330 patients).

Quality of included studies. We evaluated the quality of the 9 included studies, using the QUADAS criteria (Figure 2). All included studies were prospective, clearly described the selection criteria, and used the same reference criteria for all patients. The 9 included studies enrolled heterogeneous patient populations and may not be representative of the full spectrum of patients with autoimmune diseases. Some studies used clinically documented infection as an outcome definition but did not clearly describe whether clinical diagnosis was independent of the biomarker tests. None of the included studies described whether the physicians were blinded with regard to the index tests when they made the diagnosis of bacterial infection. In addition, none of the included studies explicitly provided explanations for withdrawal, and none showed uninterpretable results.

Study and population characteristics. Table 1 shows the characteristics of the 9 included studies. Six studies evaluated patients with various systemic autoimmune diseases, and 3 studies evaluated patients with a specific autoimmune disease. The outcome definitions were classified as microbiologically documented infection or clinically documented infection. Most of the studies used a LumiTest (Brahms Diagnostica) for procalcitonin measurement. Table 1 also shows the sensitivity and specificity of procalcitonin and CRP as biomarkers. When multiple sensitivity or specificity values at different cutoff values were reported in a study, we used the pair of values that maximized the Youden index.

Diagnostic accuracy indices. Table 2 shows the pooled sensitivity and specificity analysis for procalcitonin and CRP. Analysis of discrimination indicated that procalcitonin had a greater area under the ROC curve value (0.91 [95% CI 0.88–0.93]) compared with CRP (0.81 [95% CI 0.78–0.84]) (Figure 3). Procalcitonin had greater specificity than CRP, but CRP had greater sensitivity than procalcitonin. Procalcitonin had a high positive likelihood ratio (7.28 [95% CI 5.10–10.38]), making it an excellent rule-in test for the diagnosis of infectious complications in patients with autoimmune diseases. In contrast, CRP had a low positive likelihood ratio (1.76 [95% CI 0.88–3.49]) and was unsuitable as a rule-in test. Neither marker had a negative likelihood

ratio of <0.2, so neither was suitable as a rule-out test. The diagnostic ORs for procalcitonin and CRP were 25.2 (95% CI 15.4–41.5) and 5.06 (95% CI 1.7–14.7), respectively (Table 2 and Figure 4). There was substantial heterogeneity among the studies for CRP ($I^2 = 68.3\%$ [95% CI 18.2–87.7]) but not for procalcitonin ($I^2 = 0.0\%$ [95% CI 0.0–64.8]).

Subgroup analysis. We performed several subgroup analyses to examine the source of heterogeneity, and Table 2 shows specific test accuracy indices that may be of interest to clinicians. After restriction of analysis to studies with a cutoff value of 0.5 ng/ml, the negative likelihood ratio improved (0.27 [95% CI 0.14-0.53]), indicating that an infectious complication is unlikely when the serum level of procalcitonin is <0.5 ng/ml. Because the use of one standard cutoff point may lead to loss of diagnostic information, we sought to perform subgroup analysis with 2 sets of cutoff points, one with high sensitivity and one with high specificity. Three studies provided diagnostic accuracy indices on cutoff points of >0.5 ng/ml. Pooled results from these studies showed greatly improved specificity (0.95 [95% CI 0.89– 0.98]). Only 2 studies provided results on cutoff points of <0.5 ng/ml; therefore, we could not perform a subgroup analysis for low cutoff points. The diagnostic accuracy indices of procalcitonin in the 6 studies with mixed autoimmune disease populations did not differ significantly from the results of all 9 studies. Considering that procalcitonin expression may be elevated in some vasculitis syndromes and AOSD, we performed an additional analysis by excluding 5 studies containing patients with AOSD. The sensitivity of procalcitonin did improve (0.81 [95% CI 0.49-0.95]) among patients with autoimmune diseases other than AOSD.

DISCUSSION

We evaluated the use of procalcitonin and CRP as biomarkers to identify bacterial infection as the cause of fever in patients with autoimmune diseases, by performing a meta-analysis of 9 prospective studies. Our results indicate that procalcitonin had higher diagnostic value than CRP for differentiating between bacterial infection and disease flare in patients with autoimmune diseases. In particular, the area under the ROC curve values were 0.91 for procalcitonin and 0.81 for CRP. In general, procalcitonin was more specific than CRP as a marker of bacterial infection, and CRP was more sensitive than procalcitonin as a marker of bacterial infection. Procalcitonin had a high positive likelihood ratio, making it sufficient to serve as a rule-in biomarker. In

contrast, because CRP had a low positive likelihood ratio, CRP testing may lead to a high false-positive rate if used in isolation. Both markers had a suboptimal negative likelihood ratio and thus are not suitable for the exclusion of bacterial infection in febrile patients with autoimmune diseases.

This study provides evidence that measurement of procalcitonin is a clinically useful procedure for identification of the cause of fever in patients with autoimmune diseases. A previous meta-analysis that compared procalcitonin and CRP in the diagnosis of bacterial infections in hospitalized patients demonstrated that procalcitonin testing was more sensitive (88% versus 75%) and more specific (81% versus 67%) than CRP testing (21). A more recent meta-analysis that evaluated the expression of procalcitonin and CRP in patients after surgery or trauma showed that the area under the summary ROC curve for procalcitonin was better than that for CRP (22).

Our finding of a suboptimal sensitivity of procalcitonin (75%) in identifying bacterial infections in patients with autoimmune disease may be attributable to several factors. For instance, the studies we included used heterogeneous groups of patients as controls; some studies used patients with autoimmune disease flares, and others used healthy individuals. Previous observational studies have shown that procalcitonin expression may be elevated in the presence of several systemic vasculitis syndromes (granulomatosis with polyangiitis [Wegener's] [23], Kawasaki disease [14], AOSD [5], or Goodpasture's syndrome [24]), even in the absence of bacterial infection.

Different cutoff levels are needed to optimize the discriminative capability of procalcitonin for different autoimmune diseases. However, this requires a large study or studies of patients with the same autoimmune disease. Until the results of such studies are available, we recommend using a standard procalcitonin cutoff value of 0.5 ng/ml, which has reasonable sensitivity (76%) and specificity (88%).

Our finding of suboptimal sensitivity of procalcitonin in identifying bacterial infections in febrile patients with autoimmune disease may also be attributable to the imperfect ascertainment of outcomes of infectious complications. In particular, some of the analyzed studies used only clinical criteria to document infection. There is no gold standard test for bacterial sepsis, so an interventional trial that compares procalcitonin-guided treatment and conventional treatment, as previously reported for respiratory tract infection (25), will provide stronger evidence than that provided by the cur-

rent observational studies. Another factor that can explain the suboptimal sensitivity of procalcitonin may be the procalcitonin testing system used in the included studies. Five of the 9 included studies used a procalcitonin line immunoassay (LumiTest) with a reported functional sensitivity of 0.5 ng/ml (coefficient of variation 20%). Values of <0.5 ng/ml (a cutoff point that is still of clinical importance) may lack precision. Only 2 studies used VIDAS assays (bioMérieux), which have higher precision (functional sensitivity of 0.09 ng/ml), and none used the most advanced Kryptor procalcitonin assay (Brahms Diagnostica), which has a reported functional sensitivity of 0.06 ng/ml. With use of these highly sensitive assays, the performance of procalcitonin testing in differentiating bacterial infection from autoimmune disease flare may be further improved.

The current study has strengths and limitations. A major strength is that we reported likelihood ratios in addition to sensitivity, specificity, and the area under the ROC curve. In contrast to sensitivity and specificity, likelihood ratios are less likely to change with the prevalence of the disorder, can be compared among different studies, and can be used to calculate posttest probability for a target disorder (26). One of the major limitations of our study is that we included studies that examined patients with a single autoimmune disease, such as AOSD (5), neuro-Behçet's disease (9), and acute arthritis (7). This may have led to between-study heterogeneity. We decided to include these studies because they may have increased the precision of our metaanalysis results, and because our meta-regression indicated that the results of the overall analysis did not significantly differ when those subgroups were included.

In conclusion, the results of our meta-analysis indicate that procalcitonin has a higher diagnostic value than CRP for differentiating bacterial infection from disease flare in febrile patients with autoimmune disease. Analysis of the pooled data suggests that procalcitonin is a more specific indicator of bacterial infection than CRP, but that CRP is a more sensitive indicator of bacterial infection than procalcitonin. Thus, procalcitonin testing should not be used in isolation as a rule-out tool. Future studies should investigate whether a procalcitonin-guided treatment algorithm provides better outcome in patients with autoimmune disease who present with fever.

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AUTHOR CONTRIBUTIONS

All authors were involved in drafting the article or revising it critically for important intellectual content, and all authors approved the final version to be published. Dr. Chang had full access to all of the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

Study conception and design. Wu, S.-H. Lee, Shen, Yo, Cheng. Acquisition of data. Hsieh.

Analysis and interpretation of data. Chan, C.-C. Lee, Chang.

REFERENCES

- Zein N, Ganuza C, Kushner I. Significance of serum C-reactive protein elevation in patients with systemic lupus erythematosus. Arthritis Rheum 1979;22:7–12.
- Williams RC Jr, Harmon ME, Burlingame R, Du Clos TW. Studies of serum C-reactive protein in systemic lupus erythematosus. J Rheumatol 2005;32:454–61.
- 3. Assicot M, Gendrel D, Carsin H, Raymond J, Guilbaud J, Bohuon C. High serum procalcitonin concentrations in patients with sepsis and infection. Lancet 1993;341:515–8.
- Brunkhorst R, Eberhardt OK, Haubitz M, Brunkhorst FM. Procalcitonin for discrimination between activity of systemic autoimmune disease and systemic bacterial infection. Intensive Care Med 2000;26 Suppl 2:S199–201.
- Chen DY, Chen YM, Ho WL, Chen HH, Shen GH, Lan JL. Diagnostic value of procalcitonin for differentiation between bacterial infection and non-infectious inflammation in febrile patients with active adult-onset Still's disease. Ann Rheum Dis 2009:68:1074-5.
- Eberhard OK, Haubitz M, Brunkhorst FM, Kliem V, Koch KM, Brunkhorst R. Usefulness of procalcitonin for differentiation between activity of systemic autoimmune disease (systemic lupus erythematosus/systemic antineutrophil cytoplasmic antibodyassociated vasculitis) and invasive bacterial infection. Arthritis Rheum 1997;40:1250–6.
- Martinot M, Sordet C, Soubrier M, Puechal X, Saraux A, Liote F, et al. Diagnostic value of serum and synovial procalcitonin in acute arthritis: a prospective study of 42 patients. Clin Exp Rheumatol 2005;23:303–10.
- Scire CA, Cavagna L, Perotti C, Bruschi E, Caporali R, Montecucco C. Diagnostic value of procalcitonin measurement in febrile patients with systemic autoimmune diseases. Clin Exp Rheumatol 2006;24:123–8.
- Suzuki N, Mizuno H, Nezu M, Takai Y, Misu T, Kuroda H, et al. Procalcitonin might help in discrimination between meningeal neuro-Behcet disease and bacterial meningitis. Neurology 2009; 72:762–3.
- Tamaki K, Kogata Y, Sugiyama D, Nakazawa T, Hatachi S, Kageyama G, et al. Diagnostic accuracy of serum procalcitonin concentrations for detecting systemic bacterial infection in patients with systemic autoimmune diseases. J Rheumatol 2008;35: 114-9.
- 11. Joo K, Park W, Lim MJ, Kwon SR, Yoon J. Serum procalcitonin

- for differentiating bacterial infection from disease flares in patients with autoimmune diseases. J Korean Med Sci 2011;26: 1147–51.
- 12. Sleglova O, Dejmkova H, Uhrova J. Může stanoveníi séirovéiho prokalcitoninu pomoci v diferenciáilní diagnóize mezi infekcí a akutniìm zhoršeniìm u pacientů se systéi movýim autoimunitním onemocnením. Ceska Revmatologie 2010;18:4–11.
- Zycinska K, Wardyn KA, Zielonka TM, Tyszko P, Straburzynski M. Procalcitonin as an indicator of systemic response to infection in active pulmonary Wegener's granulomacytosis. J Physiol Pharmacol 2008;59 Suppl 6:839–44.
- Okada Y, Minakami H, Tomomasa T, Kato M, Inoue Y, Kozawa K, et al. Serum procalcitonin concentration in patients with Kawasaki disease. J Infect 2004;48:199–205.
- Leeflang MM, Deeks JJ, Gatsonis C, Bossuyt PM. Systematic reviews of diagnostic test accuracy. Ann Intern Med 2008;149: 889–97.
- Whiting P, Rutjes AW, Reitsma JB, Bossuyt PM, Kleijnen J. The development of QUADAS: a tool for the quality assessment of studies of diagnostic accuracy included in systematic reviews. BMC Med Res Methodol 2003;3:25.
- Schisterman EF, Perkins NJ, Liu A, Bondell H. Optimal cut-point and its corresponding Youden Index to discriminate individuals using pooled blood samples. Epidemiology 2005;16:73–81.
- Chu H, Cole SR. Bivariate meta-analysis of sensitivity and specificity with sparse data: a generalized linear mixed model approach. J Clin Epidemiol 2006;59:1331–2.
- Jones CM, Athanasiou T. Summary receiver operating characteristic curve analysis techniques in the evaluation of diagnostic tests. Ann Thorac Surg 2005;79:16–20.
- 20. Higgins JP, Thompson SG. Quantifying heterogeneity in a metaanalysis. Stat Med 2002;21:1539–58.
- Simon L, Gauvin F, Amre DK, Saint-Louis P, Lacroix J. Serum procalcitonin and C-reactive protein levels as markers of bacterial infection: a systematic review and meta-analysis. Clin Infect Dis 2004;39:206–17.
- Uzzan B, Cohen R, Nicolas P, Cucherat M, Perret GY. Procalcitonin as a diagnostic test for sepsis in critically ill adults and after surgery or trauma: a systematic review and meta-analysis. Crit Care Med 2006;34:1996–2003.
- Moosig F, Csernok E, Reinhold-Keller E, Schmitt W, Gross WL. Elevated procalcitonin levels in active Wegener's granulomatosis. J Rheumatol 1998;25:1531–3.
- Morath C, Sis J, Haensch GM, Zeier M, Andrassy K, Schwenger V. Procalcitonin as marker of infection in patients with Goodpasture's syndrome is misleading. Nephrol Dial Transplant 2007;22: 2701–4
- Schuetz P, Christ-Crain M, Thomann R, Falconnier F, Wolbers M, Widmer I, et al. Effect of procalcitonin-based guidelines vs standard guidelines on antibiotic use in lower respiratory tract infections: the ProHOSP randomized controlled trial. JAMA 2009;302:1059–66.
- Grimes DA, Schulz KF. Refining clinical diagnosis with likelihood ratios. Lancet 2005;365:1500–5.