MEDICAL GENOMICS LABORATORY

Next-Gen Sequencing and Deletion/Duplication Analysis of *NF2* Only (NF2-NG)

Ordering Information

Acceptable specimen types:

- Fresh blood sample (3-6 ml EDTA; no time limitations associated with receipt)
- Saliva (OGR-575 DNA Genotek; kits are provided upon request)
- DNA (extracted from lymphocyte cells; a minimum volume of 25μL at 3μg; O.D. of 260:280nm ≥1.8; must be extracted in a CLIA or equivalent certified lab)
- Flash frozen tumor sent on dry ice
- Fresh tumor or affected tissue biopsy, immersed in sterile culture media (PBS/RPMI)

Turnaround time:

30 working days for blood, saliva, or DNA; 40 working days for fresh/frozen tumor

Price, CPT codes, and Z code:

\$800 for blood, saliva, or DNA (USD – institutional/self-pay);

\$1,500 for fresh/frozen tumor (USD – institutional/self-pay);

CPT: 81406, 81405

Z code: ZB6AA

Candidates for this test:

Patients with bilateral vestibular schwannomas with or without other typical NF2-associated features (congenial cataracts, ependymoma, facial weakness, etc).

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Specimen shipping and handling:

- Please find acceptable specimen type above.
- All submitted specimens must be sent at room temperature. DO NOT ship on ice.
- Specimens must be packaged to prevent breakage and absorbent material must be included in the package to absorb liquids in the event that breakage occurs. Also, the package must be shipped in double watertight containers (e.g. a specimen pouch + the shipping company's diagnostic envelope).
- To request a sample collection kit, please visit the website or email medgenomics@uabmc.edu to complete the specimen request form.
- Please contact the MGL (via email at medgenomics@uabmc.edu, or via phone at 205-934-5562) prior to sample shipment and provide us with the date of shipment and tracking number of the package so that we can better ensure receipt of the samples.

Required forms:

- Test Requisition Form
- Form for Customs (for international shipments)

Note: Detailed and accurate completion of this document is necessary for reporting purposes. The Medical Genomics Laboratory issues its clinical reports based on the demographic data provided by the referring institution on the lab requisition form. It is the responsibility of the referring institution to provide accurate information. If an amended report is necessary due to inaccurate or illegible documentation, additional reports will be drafted with charge.

Requests for testing may not be accepted for the following reasons:

- No label (patients full name and date of collection) on the specimens
- No referring physician's or genetic counselor's names and addresses
- No billing information

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DNA samples must be extracted in a CLIA or equivalent certified lab

For more information, test requisition forms, or sample collection and mailing kits, please

call: 205-934-5562.

Disorder Background

Neurofibromatosis type 2 is an autosomal dominant disorder characterized by bilateral

vestibular schwannomas with associated symptoms of tinnitus, hearing loss and balance

dysfunction. Almost all affected individuals develop bilateral vestibular schwannomas by 30

years of age. Affected individuals may also develop schwannomas of other cranial and

peripheral nerves, meningiomas, and ependymomas. NF2 is an autosomal dominant disorder

with a frequency of 1:33-40,000 births in all populations. About 50% of patients are due to a de

novo variant, where neither parent has signs of the disorder. The offspring of an affected

individual have a 50% risk of inheriting the altered NF2 gene. NF2 is the only gene in which

pathogenic variants are known to cause neurofibromatosis 2, however there is significant

overlap between NF2 and Schwannomatosis.

Test Description

The NF2-only by NGS involves sequencing as well as deletion/duplication analysis of the entire

coding NF2 region using MLPA. The test uses an extensively customized and optimized set of

Agilent HaloPlex capture probes, followed by sequencing of overlapping amplicons within the

regions of interest using 300bp paired-end Illumina sequencing chemistry. Each coding exon

plus ~50bp of flanking intronic sequence are simultaneously sequenced. 5' and 3' untranslated

sequences are not included.

The average coverage is >1600x with 100% of the NF2 coding region ≥350x. This allows for

detection of very low-level mosaicism by sequencing (as low as 3% of the alleles in 100% of the

coding region with >95% confidence). Variant and copy number calls are made using a unique

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bioinformatics pipeline detecting all types of variants including single nucleotide substitutions,

indels, and frameshifts caused by deletion/duplication up to 112bp. Deletion/duplication

analysis for NF2 using MLPA is included in this test, as such variants are a part of the variant

spectrum for these conditions.

Variant detection rate in leukocytes is >90% in non-founder NF2 patients. Variants detected

include truncating variants (nonsense, frameshift, splicing variants), missense variants, multi-

exon deletions or duplications and total gene deletions.

In about 25-30% of founders (simplex cases, patients with unaffected parents), variants are not

detected in blood lymphocytes as a result of somatic mosaicism. Only variants with mosaicism

levels greater than 10% can be detected in lymphocyte DNA (Evans et al, 2007). Identification of

the majority of mosaic variants requires testing of tumor tissue (Evans et al, 2007). As RNA is

most often degraded in available tumor material, a DNA-based comprehensive analysis is

applied.

REFERENCES available on website.

Other related testing options:

Meningiomatosis/Multiple Meningioma Panel by Next-Gen Sequencing (MEN-NG)

Schwannomatosis/Multiple Schwannoma Panel by Next-Gen Sequencing (SCH-NG)

• NF2 gDNA Sequencing and Delection/Duplication Analysis on Tumor Block (NF24)

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