

# Standardizing Risk Assessment for Treatment-Related Gonadal Insufficiency and Infertility in Childhood Adolescent and Young Adult Cancer: The Pediatric Initiative Network Risk Stratification System

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## Introduction

CONTINUED ADVANCEMENTS IN THERAPY in pediatric and adolescent oncology have led to increasing numbers of patients achieving long-term survival. Consequently, conveying the late complications of treatment has become a regular discussion topic at the time of diagnosis. This is particularly relevant for gonadotoxicity as fertility preservation interventions exist to mitigate this late effect. Studies of self-reported infertility in survivors have assessed rates of 46% in males and 13% in females.<sup>1,2</sup> Crucial to these discussions is providing an estimate of the gonadotoxic risk that a particular treatment carries. Over time and across many studies, it has been demonstrated that cancer patients who are exposed to surgery involving the reproductive system, radiation to gonadal tissue or the hypothalamus and chemotherapy that depletes reproductive germ cells risk impairment to their fertility.<sup>3–6</sup> Among chemotherapy agents, the classes of drugs most likely to cause impairment are alkylating agents and heavy metals.

Further fine tuning an estimate of risk for a given individual has been a desirable but difficult goal. Studies from which risk factors are derived are either small in number, represent the experience at a single institution, rely on self-report, and/or are retrospective in nature. The study population is often heterogeneous and outcome measures vary across studies making direct comparisons of data difficult. The true outcomes of interest, that is, the ability or inability to become pregnant or sire a pregnancy usually occurs many years after cancer treatment, when patients are often no longer in contact with their oncology physicians.<sup>7,8</sup> Therefore, most studies of reproductive potential in survivors of childhood cancer use surrogate measures for infertility such as lack of pregnancy, acute ovarian failure, premature ovarian insufficiency, diminished ovarian reserve, oligo/amenorrhea, and elevated gonadotropins in females and elevated follicle stimulating hormones, oligospermia, or

azoospermia in males.<sup>9–19</sup> In addition, the prevalence of infertility can only be determined by knowing the number of survivors who, during the same period, were attempting pregnancy.

Finally, for females specifically, gonadotoxic therapies can variably deplete a female patient's nonrenewable ovarian reserve, further complicating risk stratification. When marked depletion of the ovarian reserve occurs secondary to cancer therapy, ovarian insufficiency can develop quite soon after the completion of therapy. For those in whom partial depletion occurs, a reproductive window large enough to fulfill their goals for biologic children before the onset of premature ovarian insufficiency (POI) may exist. This window of time may also be an opportunity for fertility preservation post-treatment. In this context, the definition of risk for POI clearly has different implications and yet is extremely difficult to quantify. In men, temporary azoospermia can occur for variable periods of time post-therapy and preclude siring a pregnancy even if permanent infertility is not the ultimate clinical outcome.

A first step in remediating the difficulties in estimating risk is to establish a standardized risk stratification model based upon currently available data to serve as a new baseline moving forward in clinical practice and research. Utilizing such a risk stratification model in clinical practice and across research studies can provide a platform for uniform clinical counseling and facilitate collaboration in and comparisons across research studies. To this end, the Best Practice Committee of the Pediatric Initiative Network (PIN) of the Oncofertility Consortium has developed such a risk stratification model for future gonadal insufficiency/infertility based on cancer treatment exposure.

The process for creating this model began with the creation of a working group composed of 27 clinicians and researchers from 15 institutions (Table 1). Current stratification systems in use were compared, national and international guidelines for long-term care were reviewed,<sup>3–5</sup> and a pertinent

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TABLE 1. PARTICIPANTS IN THE RISK STRATIFICATION WORKING GROUP OF THE PEDIATRIC INITIATIVE NETWORK

| <i>Name</i>                  | <i>Specialty</i>                               | <i>Institution</i>  | <i>Role in fertility preservation</i>                                  |
|------------------------------|--|---|--|
| Antoinette Anazodo, MD       | Pediatric and adolescent oncology              | Sydney Children's Hospital, Prince of Wales Hospital; University of New South Wales                 | Service development, educator, champion, and expert                    |
| Leslie Appiah, MD            | Pediatric and adolescent gynecology            | The University of Colorado Denver and Children's Hospital Colorado Cancer and Blood Diseases Center | Director, fertility preservation and reproductive late effects program |
| Kari Bjornard, MD, MPH       | Pediatric hematology/oncology                  | St Jude Children's Research Hospital  | Oncology liaison to fertility clinic                                   |
| Karen Burns, MD              | Pediatric oncology                             | Cincinnati Children's Hospital Medical Center; University of Cincinnati College of Medicine         | Codirector, comprehensive fertility care and preservation program      |
| Brooke Cherven, PhD, MPH, RN | Pediatric and adolescent oncology              | Children's Healthcare of Atlanta, Emory University  | Nurse researcher in reproductive outcomes                              |
| Krista Childress, MD         | Pediatric and adolescent gynecology            | Children's Healthcare of Atlanta, Emory University  | Member- fertility preservation team                                    |
| Allison Close, MD            | Pediatric hematology oncology                  | Helen DeVos Children's Hospital, Michigan State University  | Leader of fertility preservation services                              |
| Daniel Green, MD             | Pediatric hematology oncology                  | St Jude Children's Research Hospital  | Clinical researcher in gonadal outcomes                                |
| Holly Hoefgen, MD            | Pediatric and adolescent gynecology            | Washington University School of Medicine  | Codirector, integrated care & fertility preservation program           |
| Yasmin Jayasinghe, PhD       | Pediatric and adolescent gynecology            | Royal Children's Hospital, University of Melbourne  | Clinical lead fertility preservation service                           |
| Lisa Klimpel, NP             | Pediatric, adolescent and young adult oncology | Children's Hospital of Orange County  | Fertility program nurse practitioner leader                            |
| James Klosky, PhD            | Pediatric psychology                           | Children's Healthcare of Atlanta, Emory University  | Member-fertility preservation team                                     |
| Mary Langevin, NP            | Pediatric oncology                             | Children's Minnesota  | Codirector AYA oncofertility/preservation program                      |
| Jennifer Levine, MD MSW      | Pediatric hematology oncology                  | Weill Cornell Medicine  | Director, fertility preservation program and director of sSurvivorship |
| Veronica Gomez-Lobo, MD      | Pediatric and adolescent gynecology            | Eunice Kennedy Shriver National Institute of Child Health and Human Development                     | Director of pediatric and adolescent obstetrics/gynecology             |
| Lillian Meacham, MD          | Pediatric endocrinologist                      | Children's Healthcare of Atlanta, Emory University  | Director of the fertility preservation program                         |
| Molly Moravek, MD            | Reproductive endocrinology and infertility     | University of Michigan  | Director fertility preservation program; IVF medical director          |
| Leena Nahata, MD             | Pediatric endocrinologist                      | Nationwide Children's Hospital  | Director of fertility and reproductive health program                  |
| Kyle Orwig, PhD              | Researcher, reproductive medicine              | University of Pittsburgh  | Director, fertility preservation program of UPMC                       |
| Olivia Prebus, RN            | AYA oncology                                   | Cook Children's Medical Center  | Oncofertility Nurse Navigator  |
| Megan Pruett, NP             | Pediatric endocrine nurse practitioner         | Children's Healthcare of Atlanta  | Member- fertility preservation team                                    |
| Erin Rowell, MD              | Pediatric surgery                              | Ann & Robert Lurie Children's Hospital  | Director, fertility and hormone preservation and restoration program   |
| Jill Samis, MD               | Pediatric endocrinology                        | Ann & Robert H. Lurie Children's Hospital of Chicago  | Member - fertility and hormone preservation and restoration program    |
| Amanda Saraf, DO             | Pediatric oncology                             | Riley Hospital for Children   | Director of oncofertility  |

(continued)

TABLE 1. (CONTINUED)

| Name                     | Specialty                         | Institution   | Role in fertility preservation  |
|--------------------------|-----------------------------------|---|---|
| Hanna Valli-Pulaski, PhD | Researcher                        | University of Pittsburgh  | Fertility preservation Program coordinator  |
| Stacy Whiteside, NP      | Pediatric and adolescent oncology | Nationwide Children's Hospital  | Nurse practitioner/patient navigator fertility & reproductive health program        |
| Mary Zelinski, PhD       | Researcher                        | Oregon National Primate Research Center; Oregon Health & Science University | Ovarian tissue Cryopreservation research; Consultant to fertility preservation team |

literature search was performed. The new proposed stratification guidelines were presented to the working group first by email and then discussed by conference call. Revisions were made in an iterative manner until consensus was obtained. Given the lack of adequate studies with self-reported infertility as an outcome, the percentage chance of infertility could not be assigned to the risk categories but instead terms describing the levels of risk were used: "minimally increased risk," "significantly increased risk," and "high level of increased risk." Female risk stratification takes into account the different risk based on prepubertal vs pubertal status at the time of exposure to alkylators and radiation (Fig. 1). The male risk stratification does not vary based on age at exposure<sup>13</sup> (Fig. 2).

#### Perspective: Clinical Benefits of a Standardized Risk Stratification System

Providing an accurate assessment of risk to fertility is important for patient knowledge in the provision of informed consent. Current guidelines recommend that all levels of risk—from low to high—be conveyed to patients as part of comprehensive cancer care. Decisions to pursue fertility preservation require a patient-centered process involving the medical team that weighs the pros and cons of such interventions. This balance of pros and cons varies by risk for infertility, age, gender, and diagnosis among other factors. In general, the pros of pursuing fertility preservation are to mitigate the risk of permanent long-term gonadal

damage. The drawbacks of pursuing fertility preservation interventions include unnecessary interventions in low risk situations, delaying the start of cancer fighting therapy, and cost as these interventions are expensive and often not covered by insurance.

Patients who have already progressed through puberty have standard of care options to preserve their fertility through cryopreservation of sperm, oocytes, or embryos. Recently, ovarian tissue cryopreservation has been deemed no longer experimental particularly in postpubertal females.<sup>20</sup> Male patients who have not yet gone through puberty have the choice of participating on a research protocol to freeze testicular tissue. The ability to provide accurate and consistent information will help providers work with families to make the best decision for their situation. Together they must weigh the risk of the procedure and any potential delay to cancer fighting therapy with the risk of permanent long-term gonadal damage.

This risk stratification system will allow clinicians to provide information to patients and families in a way that is comprehensive, uniform, and up to date across institutions. It allows for standardization of risk assessments in an area of medicine that continues to evolve. In addition, this standardization of the risk assessment across pediatric/AYA oncology institutions will provide continuity of care from diagnosis through survivorship, as patients often geographically relocate for school, work, or family. This will allow for consistent information regardless of institution should the patient and/or family need to transfer care.

|                                    |              |             | Minimally Increased Risk | Significantly Increased risk | High level of Increased risk   |
|------------------------------------|--------------|-------------|--------------------------|------------------------------|--|
| Alkylators<br>CED* gm/m2           |              | Prepubertal | CED < 8                  | 8-12                         | > 12   |
|                                    |              | Pubertal    | CED < 4                  | 4-8                          | >8   |
| Heavy Metal                        |              |             | Cisplatin<br>Carboplatin |                              |  |
| Hematopoietic Stem Cell Transplant |              |             |                          |                              | Alkylator +/-Total<br>body irradiation<br>Myeloablative and<br>Reduced intensity<br>regimens |
| Radiation<br>exposure              | Ovary        | Prepubertal |                          | < 15 Gy                      | ≥ 15 Gy  |
|                                    |              | Pubertal    |                          | < 10 Gy                      | ≥ 10 Gy  |
|                                    | Hypothalamus |             | 22-29.9                  | > 30-39.9 Gy                 | > 40 Gy  |

**FIG. 1.** Female level of risk for gonadal failure/infertility above that for the general population. \*CED.<sup>6</sup> CED, cyclophosphamide equivalent dose.

**FIG. 2.** Male level of risk for gonadal failure/infertility above that for the general population. \*CED.<sup>6</sup> CED, cyclophosphamide equivalent dose; ^RPLND, retroperitoneal lymph node dissection.

|                                    |              | Minimally Increased Risk | Significantly Increased risk | High level of Increased risk  |
|------------------------------------|--------------|--------------------------|------------------------------|---|
| Alkylators<br>CED* gm/m2           |              | CED < 4                  |                              | CED ≥ 4   |
| Hematopoietic Stem Cell Transplant |              |                          |                              | Alkylator +/-Total body irradiation<br>Myeloablative and Reduced intensity regimens |
| Heavy Metal mg/m2                  |              | Cisplatin<br>Carboplatin | Cisplatin > 500              |   |
| Radiation Exposure                 | Testicular   | 0.2-0.6Gy                | 0.7-3.9 Gy                   | ≥ 4.0 Gy  |
|                                    | Hypothalamus | 26-29.99                 | > 30-39.9 Gy                 | > 40 Gy   |
| Surgery                            |              |                          | RPLND^                       |   |

### Perspective: Research Benefits of a Standardized Risk Stratification System

The field of fertility preservation continues to evolve with new methodologies originating as research studies. Since prepubertal patients do not produce oocytes or sperm, the options currently offered to these patients for fertility preservation are cryopreservation of ovarian tissue or testicular tissue. Ovarian tissue cryopreservation in prepubertal females and testicular tissue cryopreservation are considered investigational. Although the actual harvest of these tissues (typically a unilateral oophorectomy or a testicular biopsy) is not *per se* experimental, the cryopreservation and utilization of these tissues for future reproduction remain experimental, particularly for prepubertal patients. Active research in many laboratories is moving the field forward quickly.<sup>21–26</sup> Since the goal of harvest is ultimately the clinical use of the tissue to restore fertility, these interventions should be performed under the regulatory oversight of an Institutional Review Board (IRB), a Clinical Ethics and Novel Technologies Committee, or other appropriate regulatory body. The purview of this oversight is to determine whether the risks of infertility for a given group of patients are significant enough to justify intervention and that the benefits of preserving tissue for future use outweigh the risks of the surgical procedure to retrieve the tissue. An evidence-based risk stratification system helps to establish the tipping point between risk and benefit, which is necessary to identify and counsel patients who are eligible for experimental fertility preservation.

It is also necessary to continuously assess gonadal insufficiency and infertility outcomes for those patients treated on older treatment protocols as well as the infertility risks posed by each new treatment paradigm. The risk stratification system provides a benchmark for comparison that can be tested in preclinical animal models or in patient-focused clinical studies. Now that discussions of risk for infertility at the time of diagnosis have become more standardized and the fertility preservation community is well connected, collaborative research to combine patient data and proactively track reproductive outcomes is possible. Multi-institutional collaborations are necessary to represent the diversity of patients,

diagnoses, and treatments with sufficient numbers to draw statistically significant conclusions about fertility risks.

### Summary

The risk stratification model developed and presented here summarizes the currently available data for infertility risk in the pediatric and adolescent cancer population. Given the limitations already outlined, it is recognized as being both incomplete and imperfect. It is designed, however, as a critical start to creating a more complete and nuanced risk stratification system. We advocate the utilization of this model to promote uniform counseling, comparisons across research studies, and collaboration on infertility projects. In this manner, the data that evolve from the next generation of research studies will take us to the next level of understanding risk.

### Author Disclosure Statement

No competing financial interests exist.

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### References

1. Wasilewski-Masker K, Seidel KD, Leisenring W, et al. Male infertility in long-term survivors of pediatric cancer: a report from the childhood cancer survivor study. *J Cancer Surviv.* 2014;8(3):437–47.
2. Barton SE, Najita JS, Ginsburg ES, et al. Infertility, infertility treatment, and achievement of pregnancy in female survivors of childhood cancer: a report from the Childhood Cancer Survivor Study cohort. *Lancet Oncol.* 2013;14(9):873–81.
3. Children's Oncology Group Long-Term Follow-Up Guidelines for Survivors of Pediatric, Adolescent and Young Adult Cancers. Accessed April 16, 2020 from: [www.survivorshipguidelines.org](http://www.survivorshipguidelines.org)
4. van Dorp W, Mulder RL, Kremer LC, et al. Recommendations for premature ovarian insufficiency surveillance for

- female survivors of childhood, adolescent, and young adult cancer: a report from the International Late Effects of Childhood Cancer Guideline Harmonization Group in Collaboration With the PanCareSurFup Consortium. *J Clin Oncol*. 2016;34(28):3440–50.
5. Skinner R, Mulder RL, Kremer LC, et al. Recommendations for gonadotoxicity surveillance in male childhood, adolescent, and young adult cancer survivors: a report from the International Late Effects of Childhood Cancer Guideline Harmonization Group in collaboration with the PanCareSurFup Consortium. *Lancet Oncol*. 2017;18(2):e75–e90.
  6. Green DM, Nolan VG, Goodman PJ, et al. The cyclophosphamide equivalent dose as an approach for quantifying alkylating agent exposure: a report from the Childhood Cancer Survivor Study. *Pediatr Blood Cancer*. 2014;61(1):53–67.
  7. Antal Z, Sklar CA. Gonadal function and fertility among survivors of childhood cancer. *Endocrinol Metab Clin North Am*. 2015;44(4):739–49.
  8. van den Berg M, van Dulmen-den Broeder E, Overbeek A, et al. Fertility studies in female childhood cancer survivors: selecting appropriate comparison groups. *Reprod Biomed Online*. 2014;29(3):352–61.
  9. Green DM, Zhu L, Wang M, et al. Effect of cranial irradiation on sperm concentration of adult survivors of childhood acute lymphoblastic leukemia: a report from the St. Jude Lifetime Cohort Study. *Hum Reprod*. 2017;32(6):1192–201.
  10. Levine JM, Whitton JA, Ginsberg JP, et al. Nonsurgical premature menopause and reproductive implications in survivors of childhood cancer: a report from the Childhood Cancer Survivor Study. *Cancer*. 2018;124(5):1044–52.
  11. Chemaitilly W, Li Z, Krasin MJ, et al. Premature ovarian insufficiency in childhood cancer survivors: a report from the St. Jude Lifetime Cohort. *J Clin Endocrinol Metab*. 2017;102(7):2242–50.
  12. Chow EJ, Stratton KL, Leisenring WM, et al. Pregnancy after chemotherapy in male and female survivors of childhood cancer treated between 1970 and 1999: a report from the Childhood Cancer Survivor Study cohort. *Lancet Oncol*. 2016;17(5):567–76.
  13. Green DM, Liu W, Kutteh WH. Cumulative alkylating agent exposure and semen parameters in adult survivors of childhood cancer: a report from the St. Jude Lifetime Cohort Study. *Lancet Oncol*. 2014;15(11):1215–23.
  14. Bresters D, Emons JA, Nuri N, et al. Ovarian insufficiency and pubertal development after hematopoietic stem cell transplantation in childhood. *Pediatr Blood Cancer*. 2014;61(11):2048–53.
  15. Romerius P, Ståhl O, Moëll C, et al. High risk of azoospermia in men treated for childhood cancer. *Int J Androl*. 2011;34(1):69–76.
  16. Green DM, Nolan VG, Kawashima T, et al. Decreased fertility among female childhood cancer survivors who received 22–27 Gy hypothalamic/pituitary irradiation: a report from the Childhood Cancer Survivor Study. *Fertil Steril*. 2011;95(6):1922–7, 1927 e1.
  17. Green DM, Kawashima T, Stovall M, et al. Fertility of male survivors of childhood cancer: a report from the Childhood Cancer Survivor Study. *J Clin Oncol*. 2010;28(2):332–9.
  18. Sklar CA, Mertens AC, Mitby P, et al. Premature menopause in survivors of childhood cancer: a report from the childhood cancer survivor study. *J Natl Cancer Inst*. 2006;98(13):890–6.
  19. Chemaitilly W, Mertens AC, Mitby P, et al. Acute ovarian failure in the childhood cancer survivor study. *J Clin Endocrinol Metab*. 2006;91(5):1723–8.
  20. Practice Committee of the American Society for Reproductive Medicine. Fertility preservation in patients undergoing gonadotoxic therapy or gonadectomy: a committee opinion. *Fertil Steril*. 2019;112(6):1022–33.
  21. Gassei K, Valli-Pulaski H, Close AG, et al. Male fertility preservation: current options and advances in research. In: Woodruff TK, Shah DK, Vitek WS (Eds). *Textbook of oncofertility research and practice*. Switzerland: Springer Nature; 2019.
  22. Medrano JV, Andrés MDM, García S, et al. Basic and clinical approaches for fertility preservation and restoration in cancer patients. *Trends Biotechnol*. 2018;36(2):199–215.
  23. Moravek MB, Appiah LC, Anazodo A, et al. Development of a Pediatric Fertility Preservation Program: a report from the Pediatric Initiative Network of the Oncofertility Consortium. *J Adolesc Health*. 2019;64(5):563–73.
  24. Braye A, Tournaye H, Goossens E. Setting up a cryopreservation programme for immature testicular tissue: lessons learned after more than 15 years of experience. *Clin Med Insights Reprod Health*. 2019;13:1179558119886342.
  25. Armstrong AG, Kimler BF, Smith BM, et al. Ovarian tissue cryopreservation in young females through the Oncofertility Consortium's National Physicians Cooperative. *Future Oncol*. 2018;14(4):363–378.
  26. Fayomi AP, Peters K, Sukhwani M, et al. Autologous grafting of cryopreserved prepubertal rhesus testis produces sperm and offspring. *Science*. 2019;363(6433):1314–1319.

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