

# Uric acid priming in human monocytes is driven by the AKT–PRAS40 autophagy pathway

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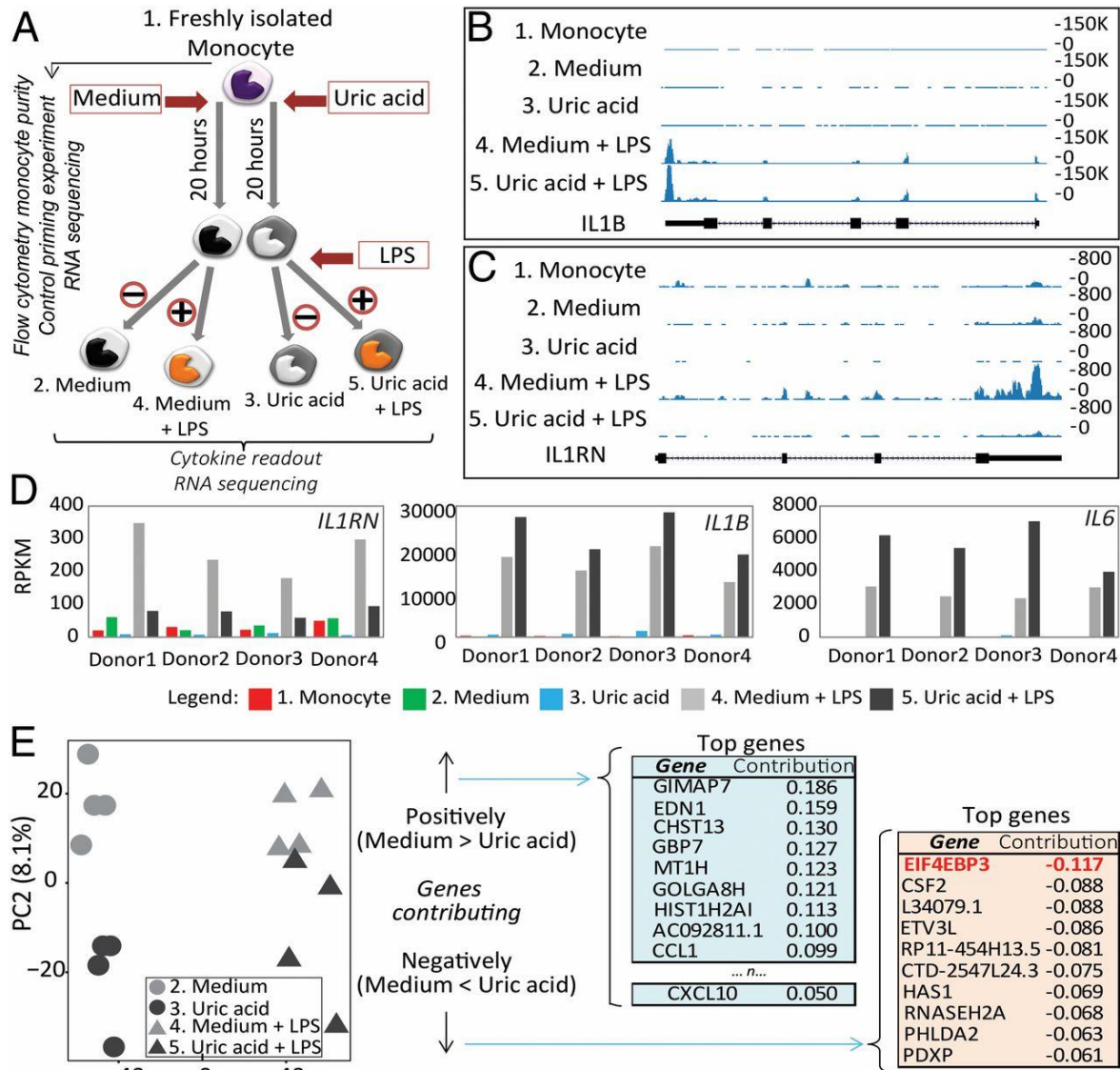
Contributed by Charles A. Dinarello, April 14, 2017 (sent for review December 22, 2016; reviewed by Patrick Durez and Pascal Richette)

Richard Reynolds  
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# Background

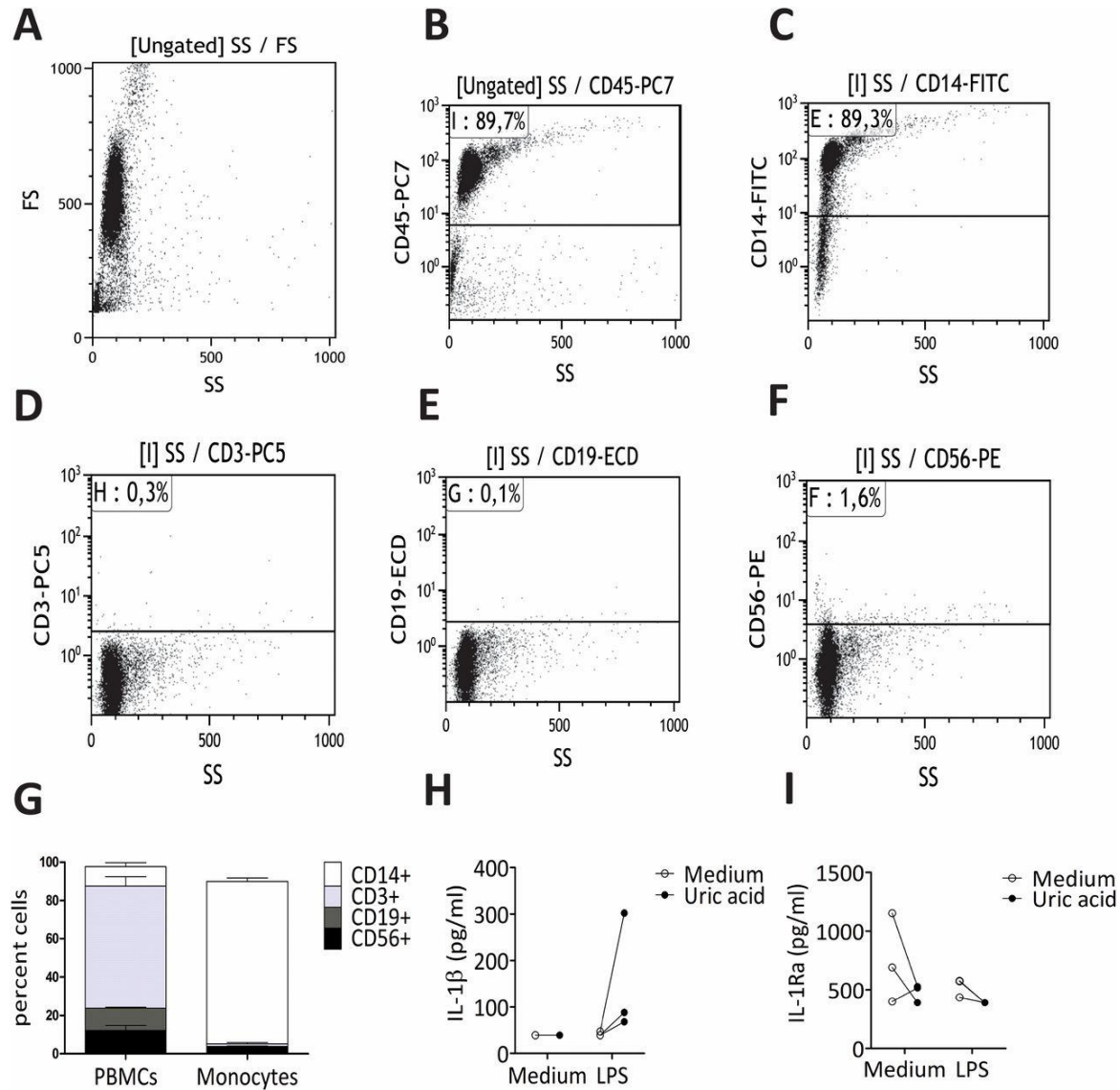
- Uric acid is the end product of purine metabolism in humans (and other hominoids).
- Hyperuricemia, UA levels  $> 6.8\text{mg/dL}$ , can cause gouty arthritis.
- MSU crystals bind nod-like receptors and cause production of proinflammatory IL-1 $\beta$ .
- Hyperuricemia also known to be associated with a number of diseases involving an inflammatory component, e.g., atherosclerosis.
- Previous work by this group demonstrated that soluble uric acid primes PBMCs by stimulating IL-1 $\beta$ , Il-6 production, and lowering production of anti-inflammatory IL-1Ra.
- Here the authors are exploring these results in more detail, trying to ascertain the mechanism by which soluble UA is upregulating cytokine production.
- Appears to involve elements of the autophagy system.

# Transcriptomic analysis in uric acid-primed monocytes.



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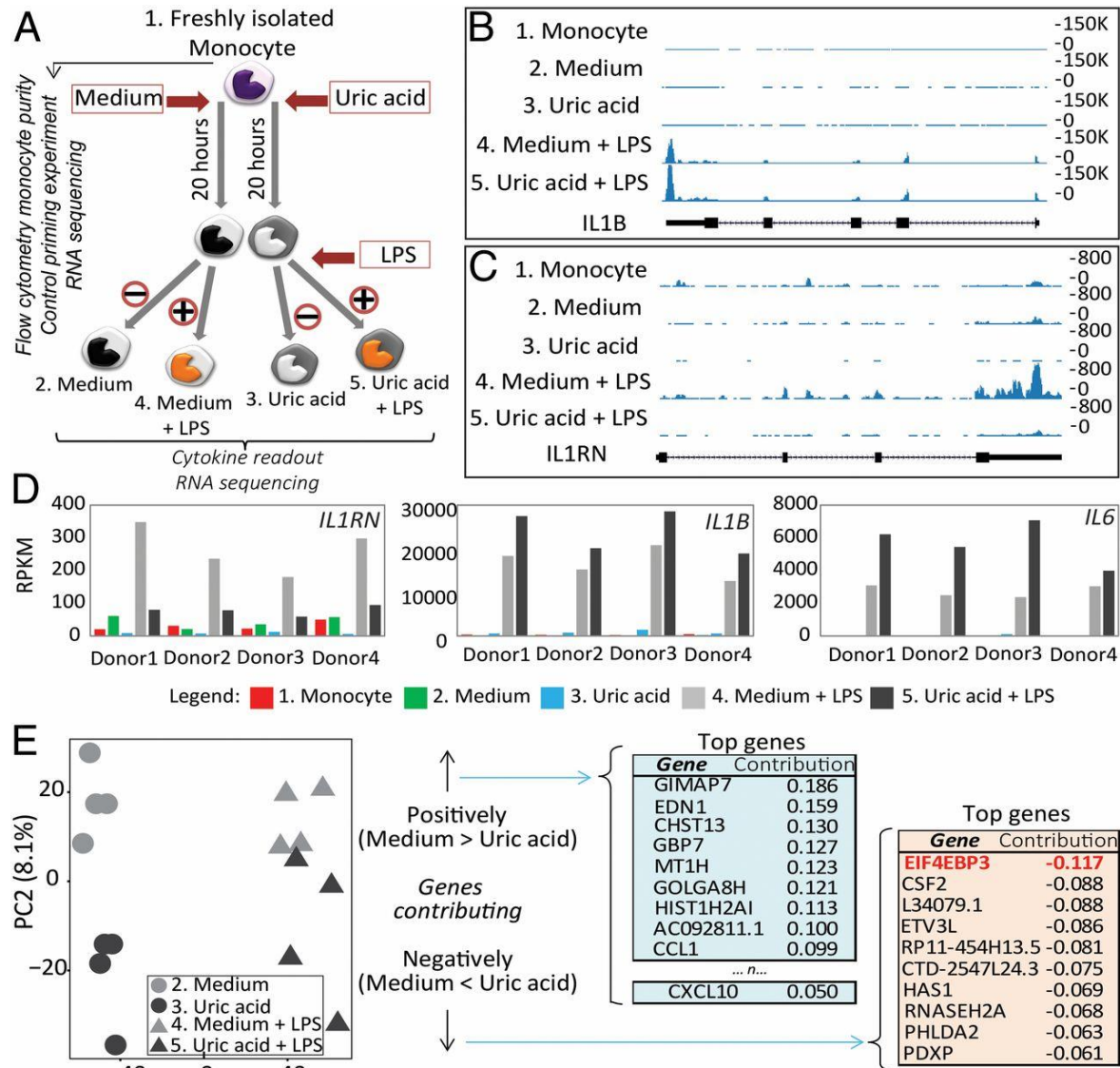
# Flow cytometry and cytokine control data for negatively selected monocytes.



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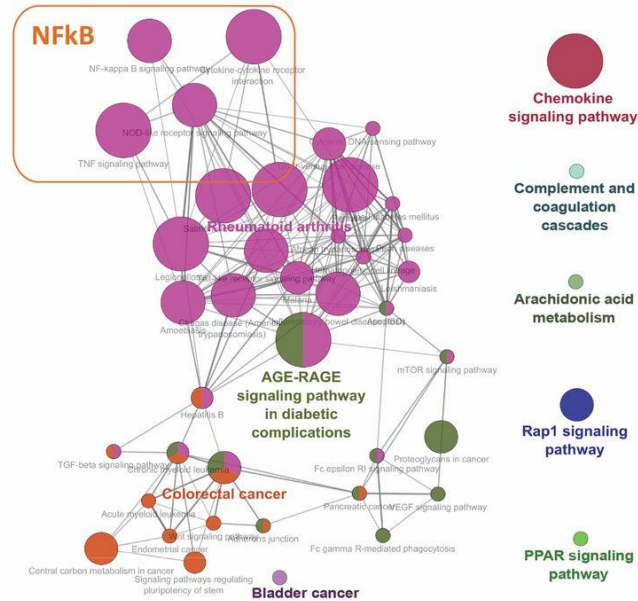
# Transcriptomic analysis in uric acid-primed monocytes.



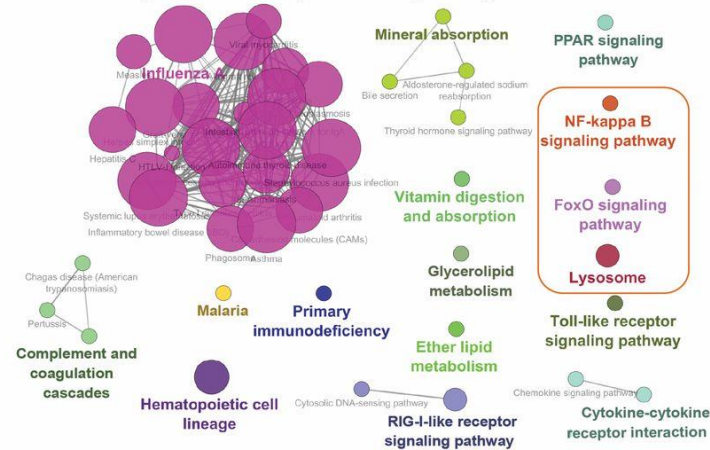
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# Pathway enrichment analysis was performed using significantly regulated genes and top 100 contributors to PC2 (Fig. 2E).

**A** KEGG Pathway enrichment analysis - Upregulated genes

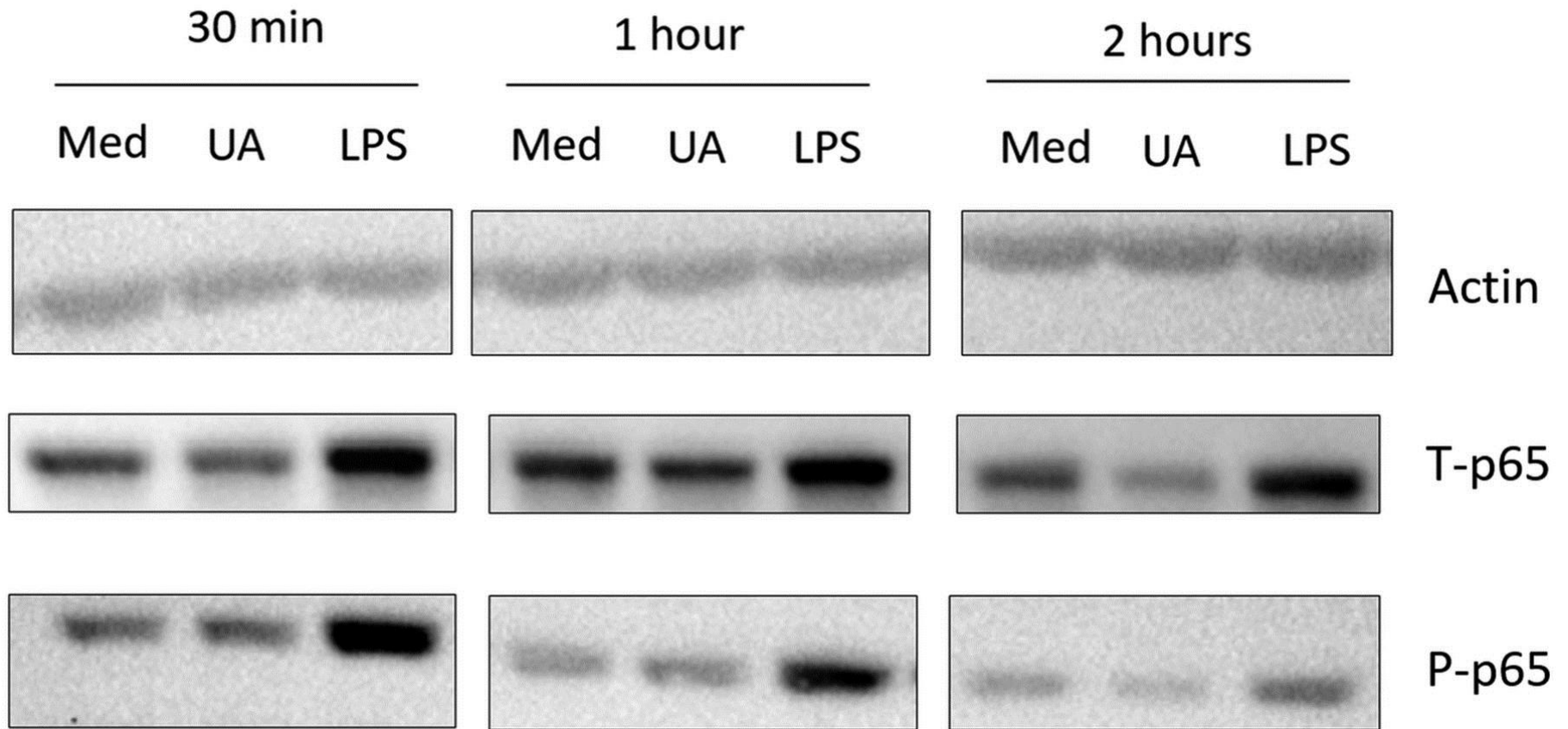


**B** KEGG Pathway enrichment analysis - Downregulated genes



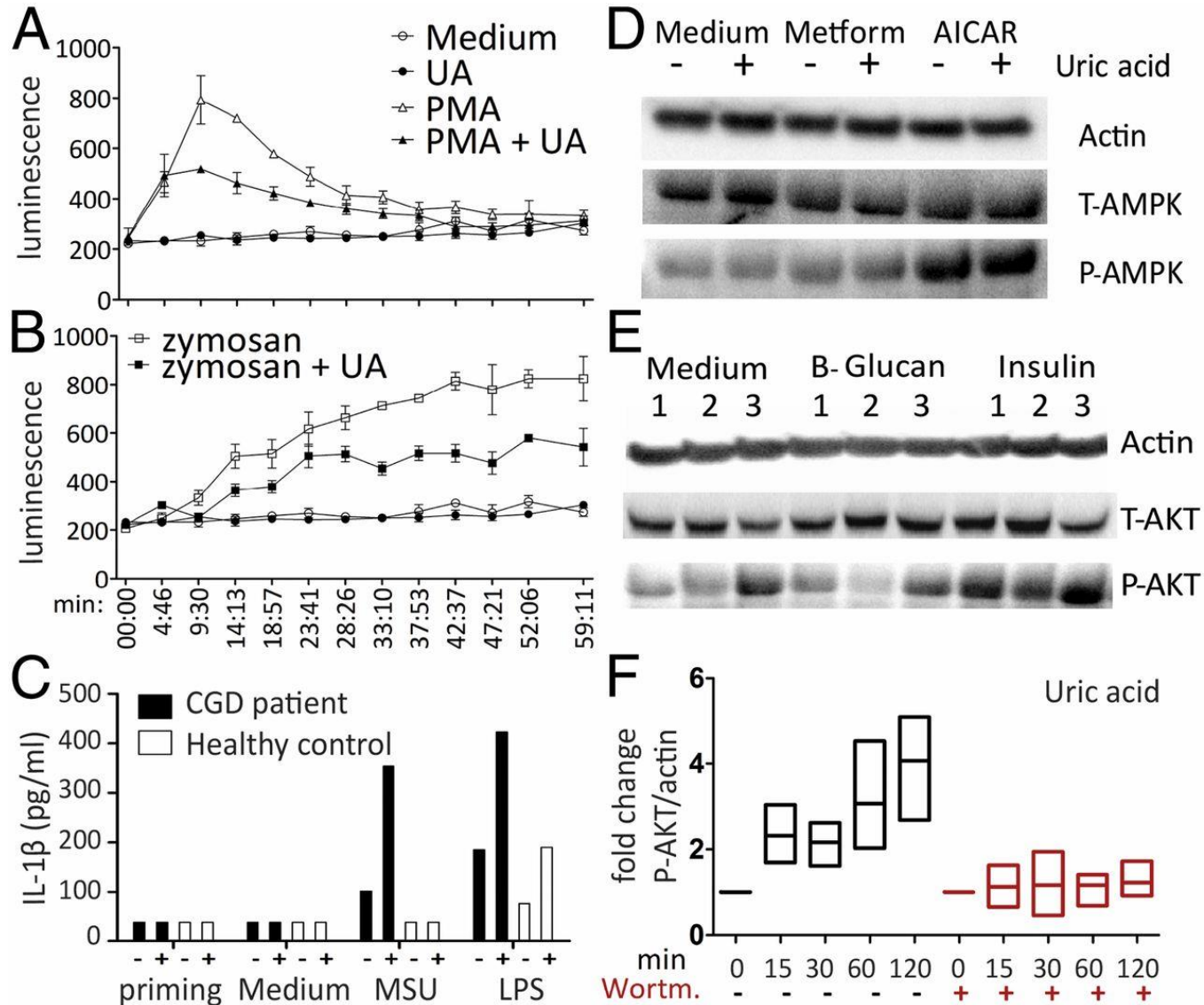
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Western blot of the p65 component of NF- $\kappa$ B in uric acid priming.



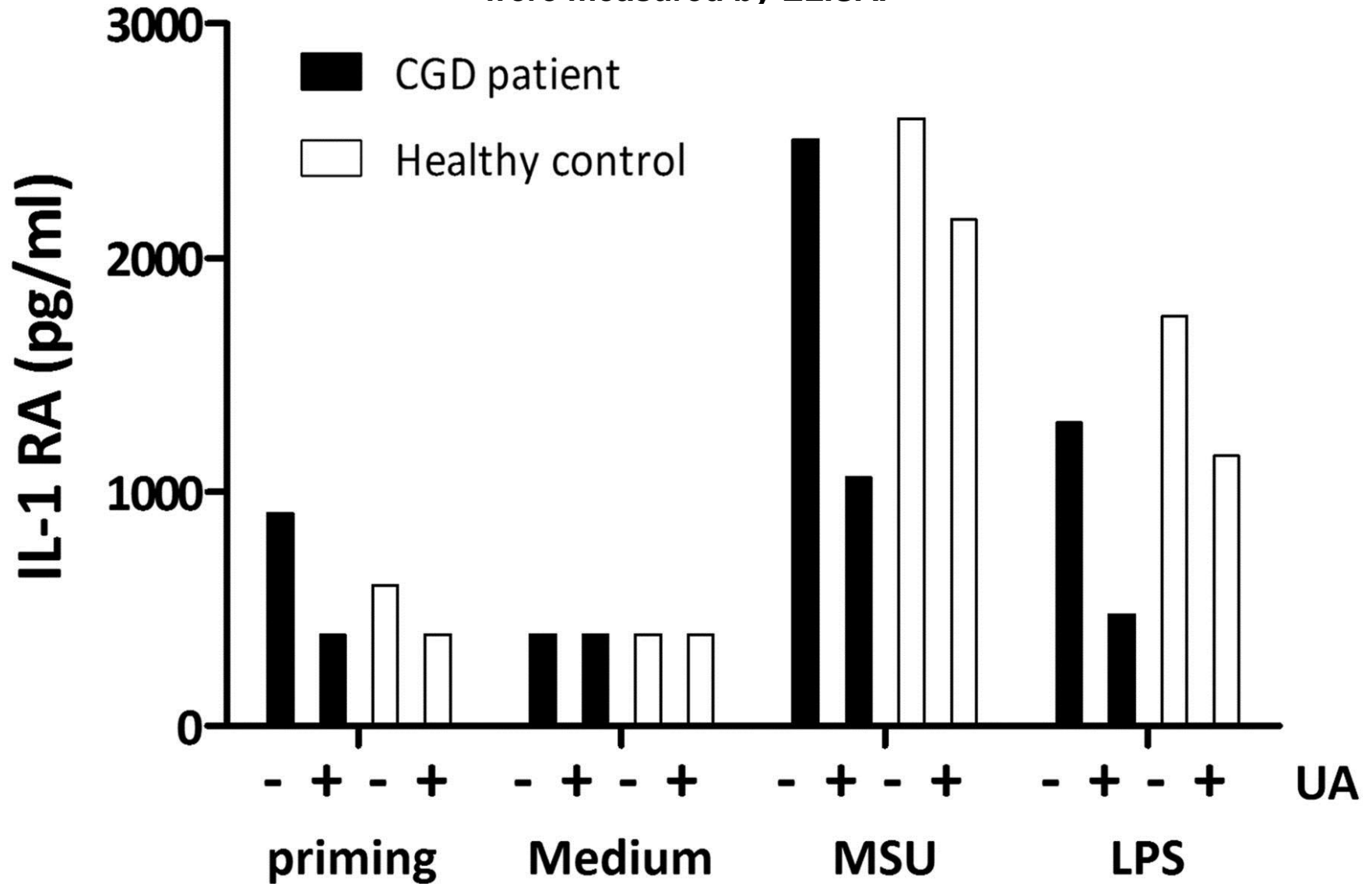
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# Phosphorylation of AKT and lack of ROS or AMPK induction by uric acid.



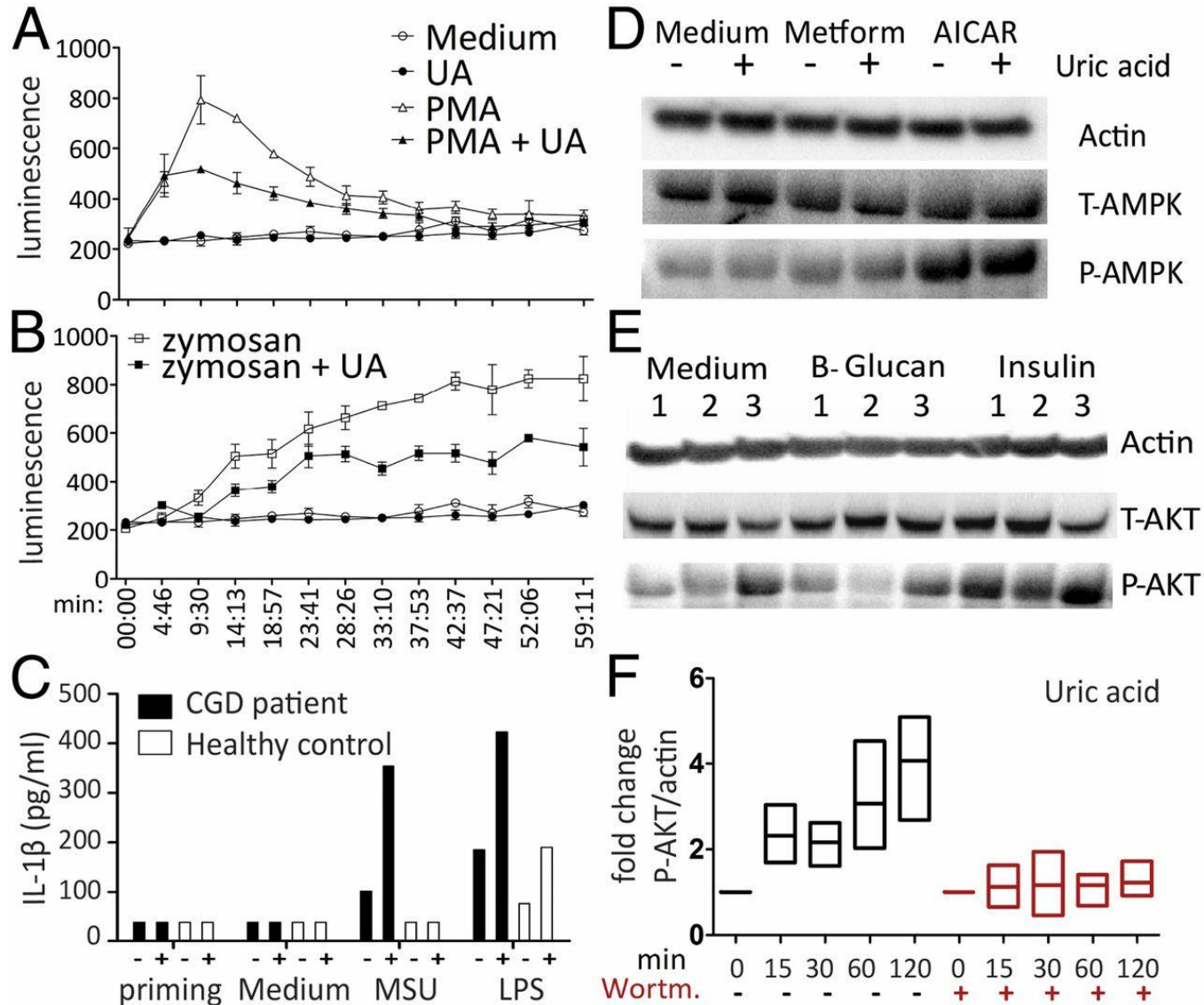
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PBMCs of a CGD patient and control were isolated, and  $0.5 \times 10^6$  cells were stimulated per well with 300  $\mu\text{g}/\text{mL}$  MSU or 10  $\text{ng}/\text{mL}$  LPS after priming with medium or uric acid, and IL-1RA levels were measured by ELISA.



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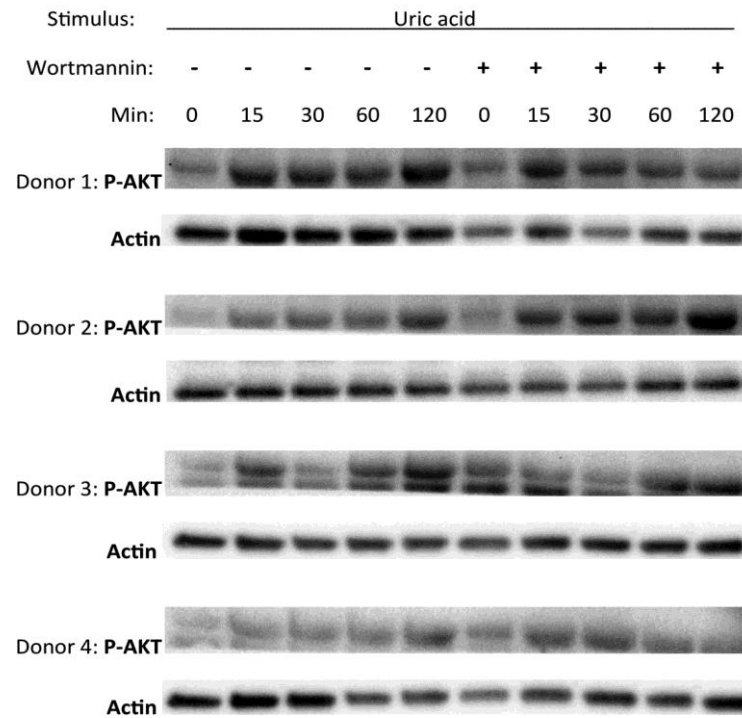
# Phosphorylation of AKT and lack of ROS or AMPK induction by uric acid.



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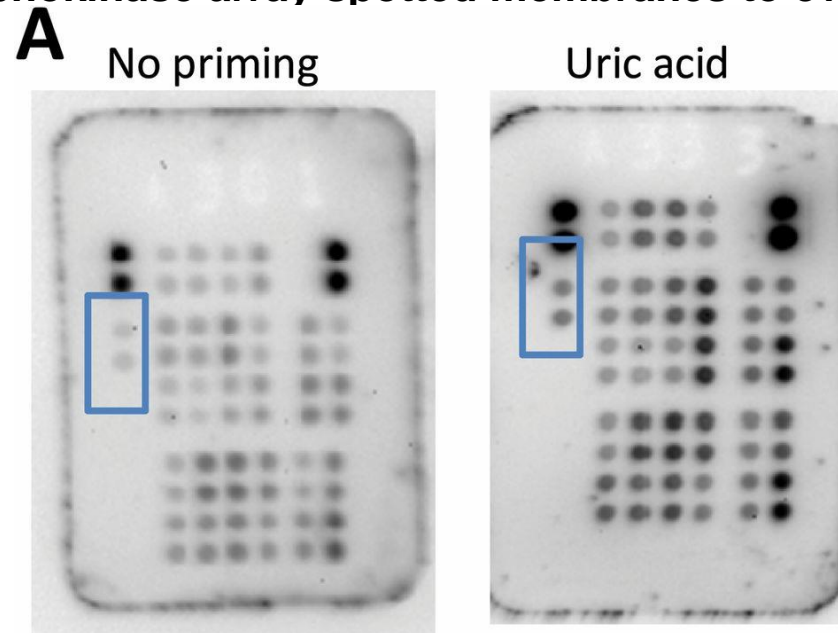
The 106 monocytes were treated with 50 mg/dL uric acid for increasing durations in the presence or absence of 100 nM wortmannin in four donors.

Uric acid		P-AKT/Actin			
Wortmannin	Min	Donor 1	Donor 2	Donor 3	Donor 4
-	0	1,00	1	1,00	1,00
-	15	1,97	1,69	3,04	2,58
-	30	2,62	1,95	1,61	2,47
-	60	2,25	2,03	3,44	4,54
-	120	3,44	2,68	5,06	5,10
+	0	1,00	1,00	1,00	1,00
+	15	1,63	1,51	0,66	0,72
+	30	1,94	1,65	0,46	0,62
+	60	1,24	1,34	1,41	0,69
+	120	1,28	1,72	0,92	0,99



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Negatively selected monocytes were primed with or without uric acid for 24 h, followed by 7 min of short exposure to high concentrations of 1  $\mu$ g/mL LPS. Cells were lysed and lysates were exposed to phosphokinase array spotted membranes to evaluate the downs...

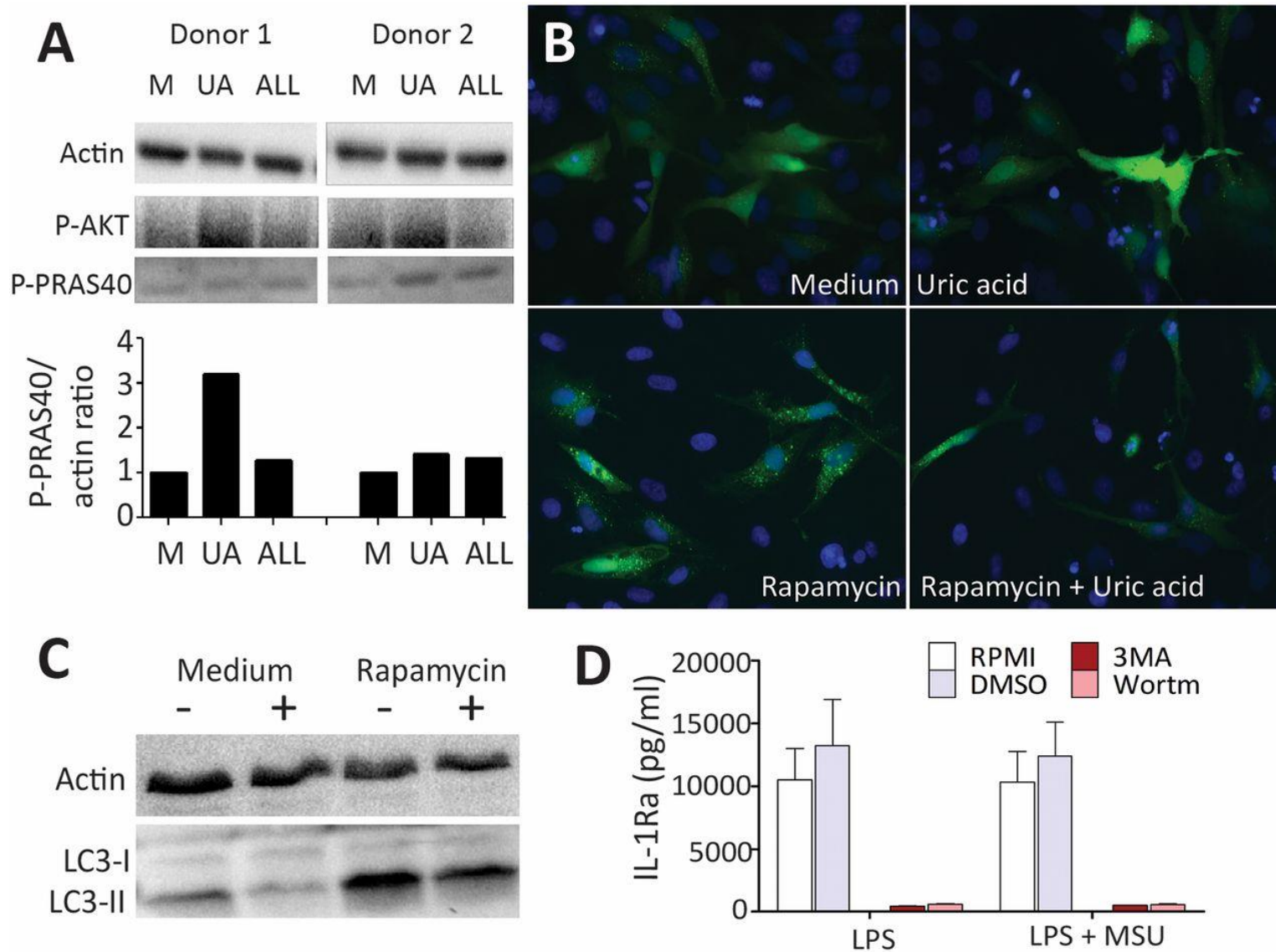


**B**

Kinase	Site	%change
<b>Fyn</b>	Y420	-77
<b>Hck</b>	Y411	-76
<b>p27</b>	T198	-73
<b>GSK-3 alpha/beta</b>	S21/S9	42
<b>ERK 1/2</b>	T202/Y204, T185/Y187	54
<b>PRAS40</b>	T246	72

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# Identification of PRAS40 as intermediate in AKT signal transduction to inhibit autophagy in uric acid-primed cells.

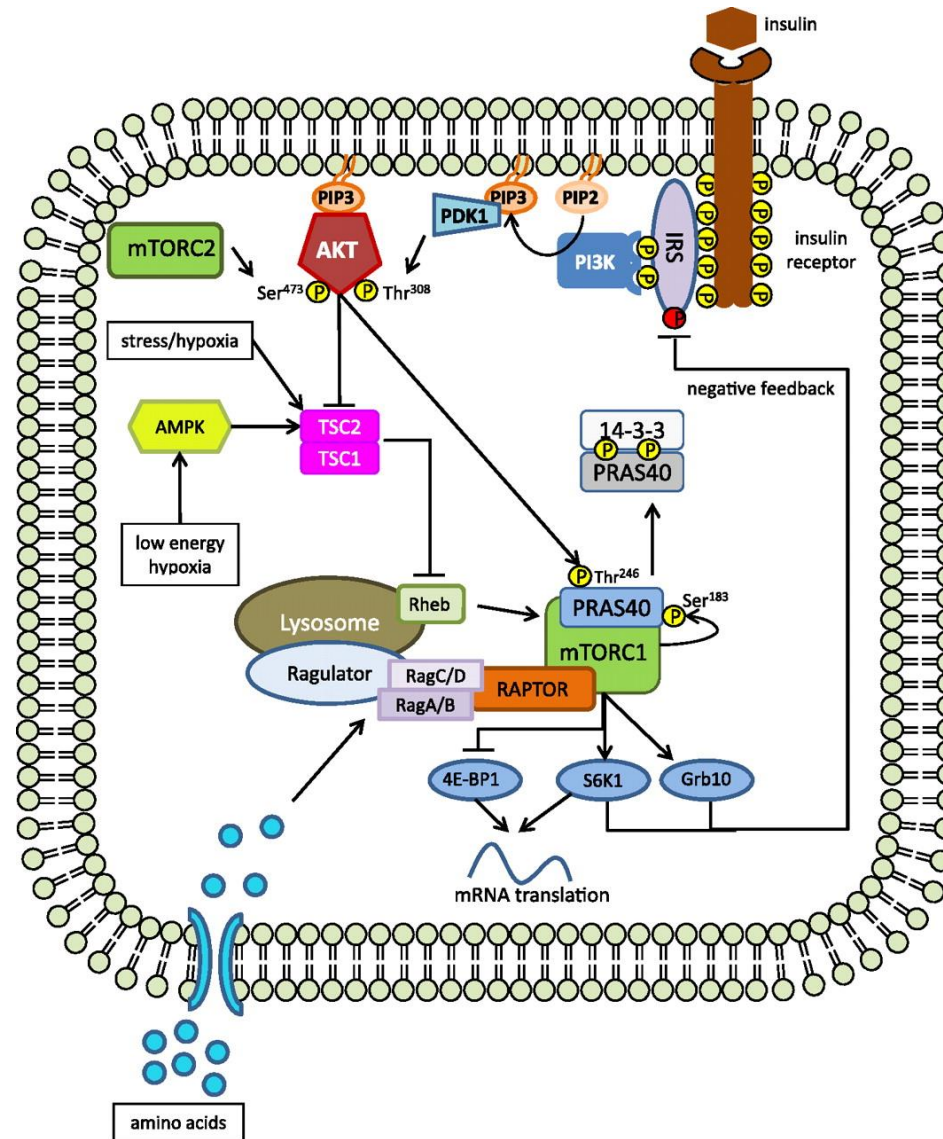


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# Take homes

- The authors have suggested a autophagy mechanism for the increased proinflammatory and decreased anti-inflammatory cytokine production which occurs in response to uric acid.
- Apparently the mechanism is via the AKT signaling pathway.
  - Pathway analysis implicated mTOR signaling (downstream of AKT).
  - A top hit from the RNA-seq experiment was overexpressed in uric acid treated cells and involved mTOR.
  - Uric acid exhibited modulatory effects on several components of the AKT pathway, i.e., AKT, PRAS40, autophagosome formation.
- Uric acid stimulated monocytes may inhibit autophagy.
  - This seems to be manifested by known downstream effects on increased production of IL-1 $\beta$ , and decreased anti-inflammatory IL1Ra.

# Regulation of PRAS40 phosphorylation by Akt and mTORC1.



Claudia Wiza et al. Am J Physiol Endocrinol Metab  
2012;302:E1453-E1460

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