

Manuscript: “OX40 signaling is involved in the autoactivation of CD4<sup>+</sup>CD28<sup>-</sup> T cells and contributes to pathogenesis of autoimmune arthritis”

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Rheumatology JC

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# Disclosures

- NIH
  - 2 T32 HL007457-36A1
  - UAB Division of Clinical Immunology and Rheumatology for pre and post-doctoral trainees
- Illumina
  - In-kind contribution
  - study epistasis between HLA risk alleles in RA

# Rationale for selection of article:

- Next talk is on Phase III trial of abatacept in PsA.
- “Regarding [synovial inflammation in PsA] agents targeting IL-23/IL-17 can achieve complete clearing of psoriatic skin lesions without a similar level of efficacy in the skin. We speculate that T-cell subsets driving pathology in the skin differ with respect to their expression of CD28 and hence their abatacept susceptibility”

# Rationale, continued

- So, I wanted to focus on a recent study involving CD4+ cells with
  - variable CD28 expression
  - related to an immune phenotype
  - Uses flow & other data
- Not expert on methods used in this manuscript
  - E.g. flow cytometry
  - Would love a dialectic on how data quality may impact the findings in this study.

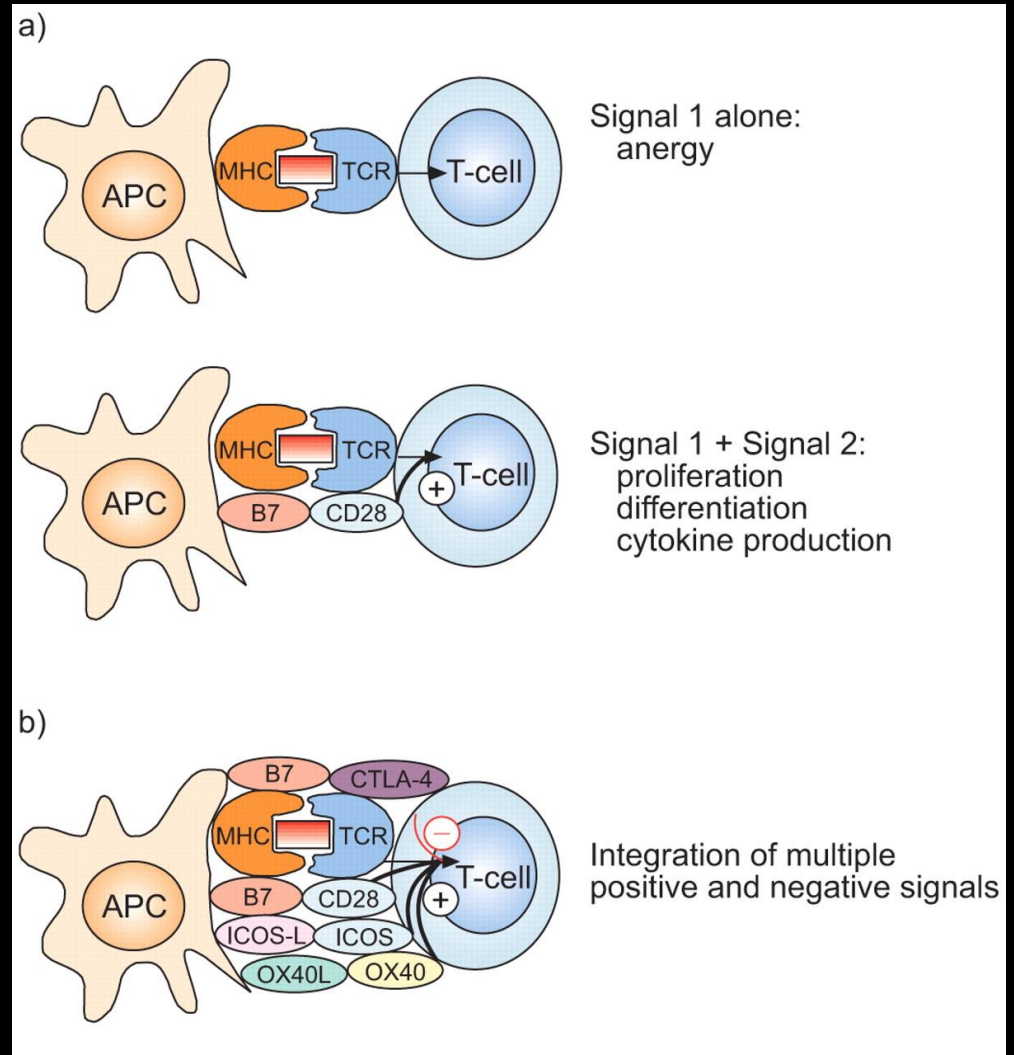
# Background

# T-cell co-stimulation

Signal I:  
TCR – MHC

Factors influencing Signal II:

CD28 – B7  
CTLA4 – B7  
CD40L – CD40  
OX40 – OX40L  
4BB-1 – CD137L

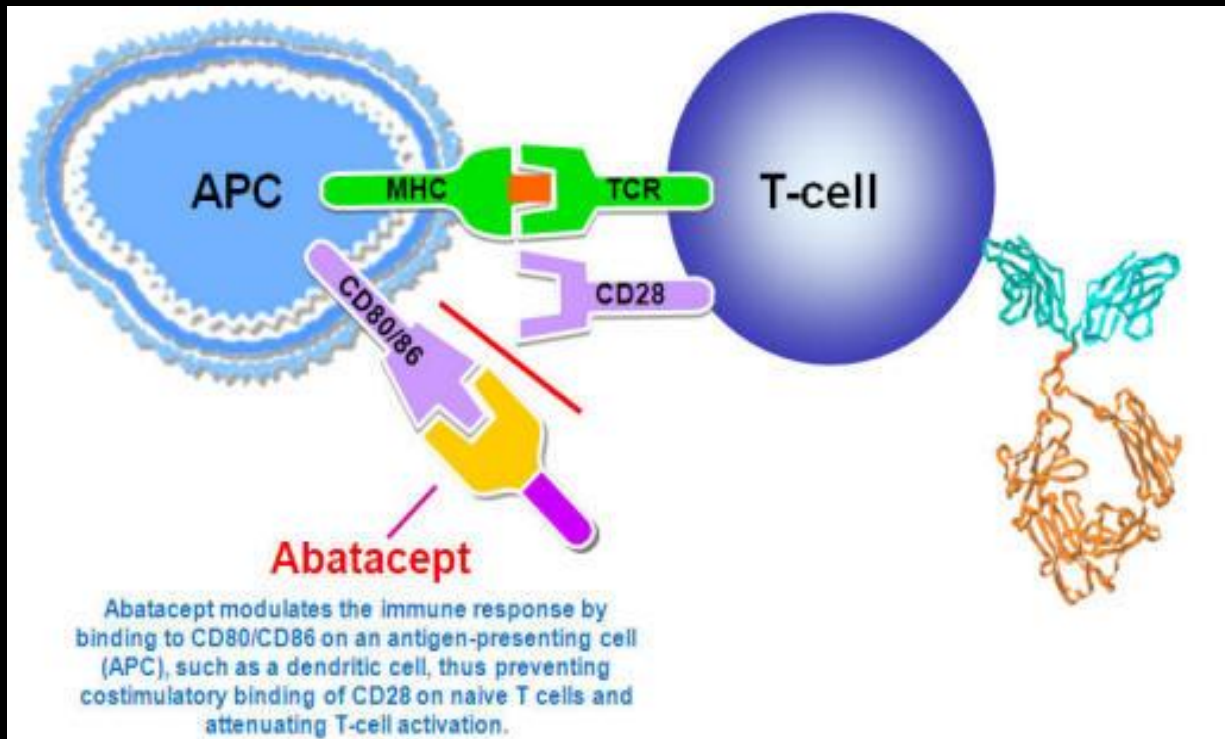
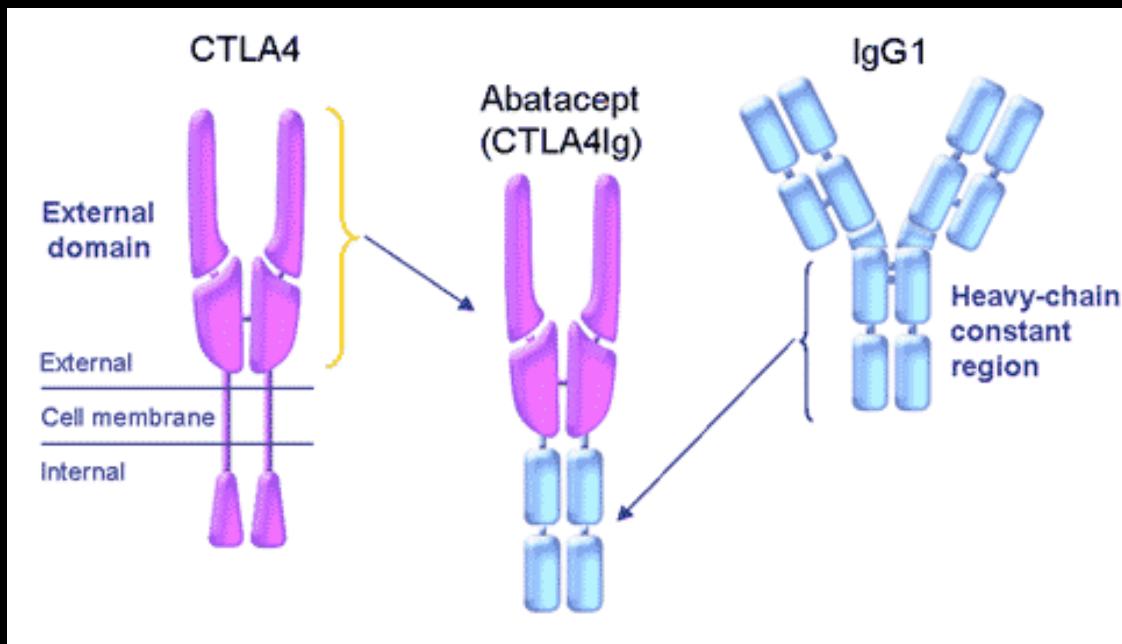


# CD28

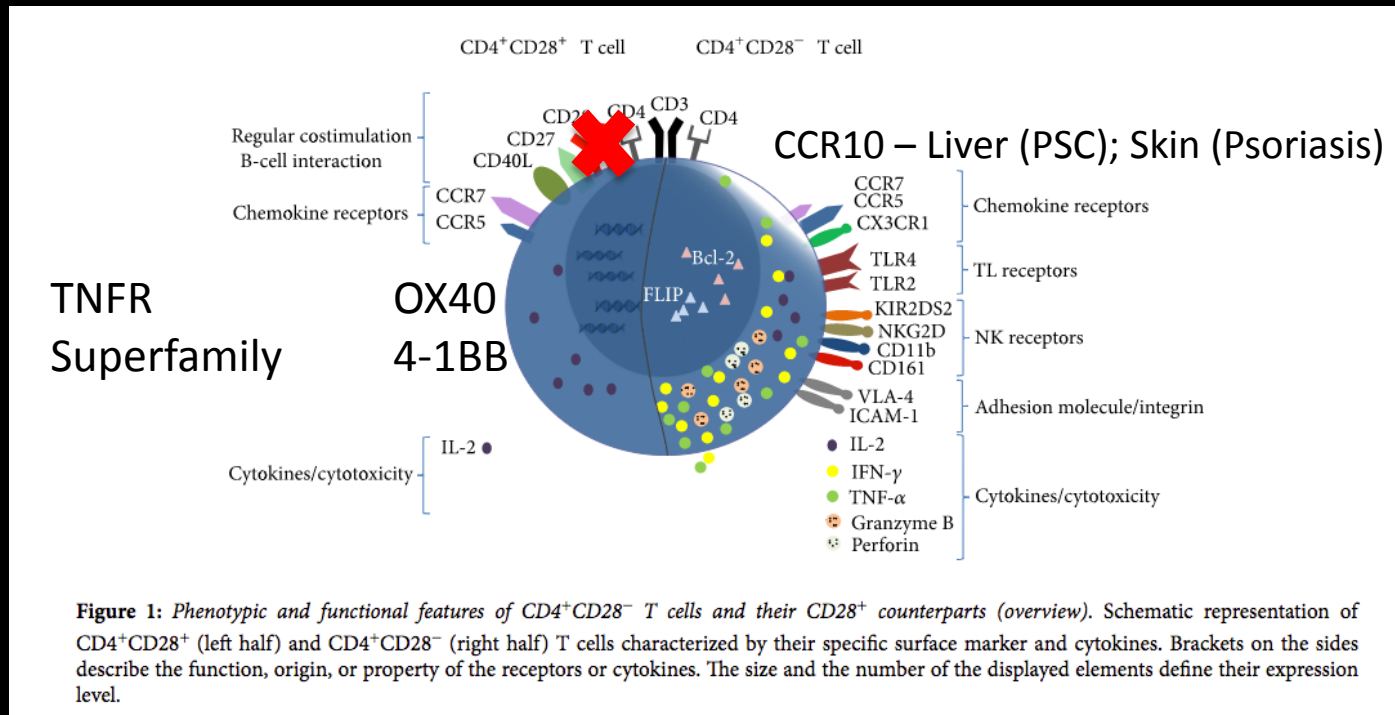
- Binds B7 (CD80/86) → costimulation
  - Survival
  - IL-2 production
  - Metabolic Activity
  - Clonal Expansion

# Abatacept (Orencia)

- CTLA4Ig ...
- Extracellular domain of CTLA4
- Fc portion of Ig
- Binds CD80/86, preventing CD28 binding



# CD4<sup>+</sup>CD28<sup>-</sup> T-cell characteristics



Described in RA ~20YA

More common in >65YOA; chronic disease states

Express variety of markers not often found tog.

Chemokine markers, including those related to tissue invasion (differ by disease state)

IL-2 secretors, but apparently independently of CD28

High amounts of IFN $\gamma$ , perforin, granzyme (“cytotoxic Thelper 1”)

Produce high amounts of TNF $\alpha$  → may help keep CD28lo.

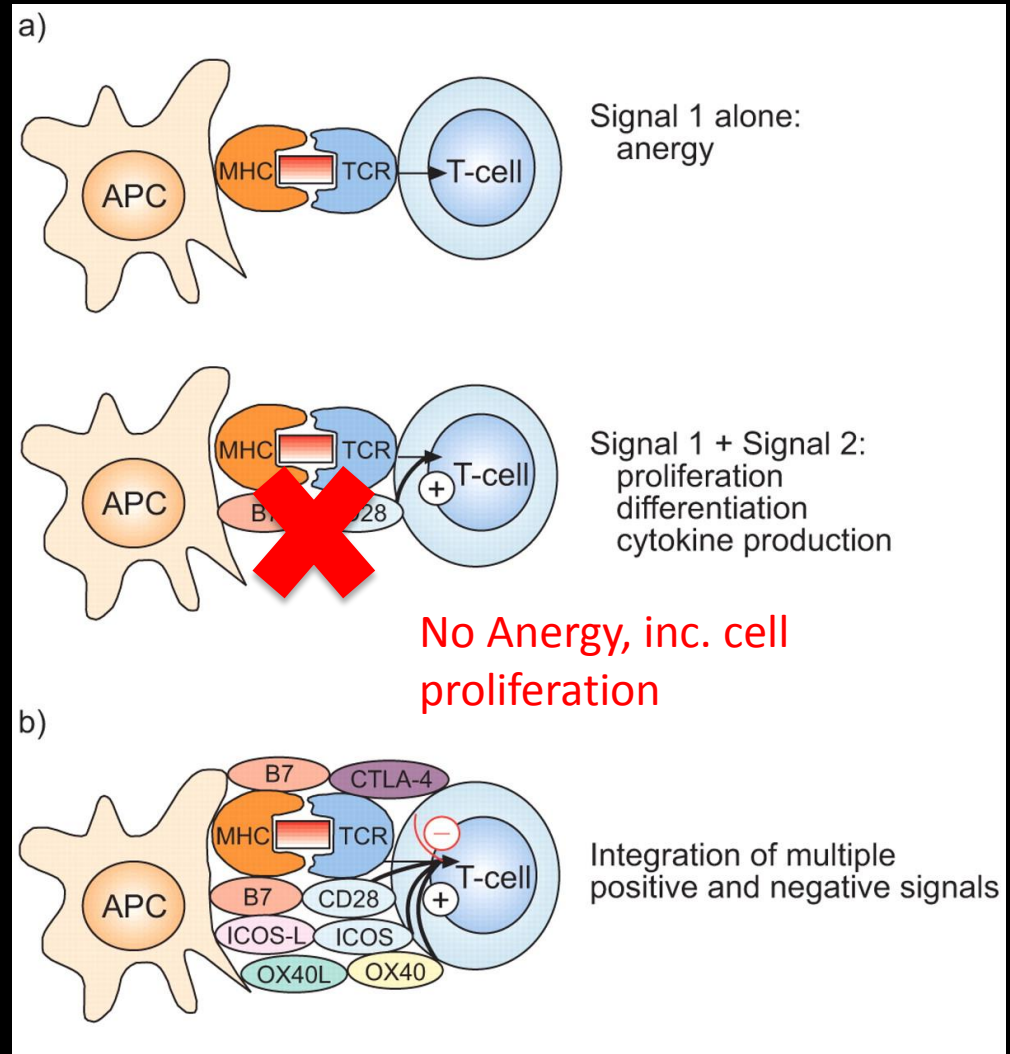
# CD4+CD28- cells show increased proliferation?!

Shortened telomere length  
No CD28 / abnormal Lack of costimulation, yet

Increased cell proliferation, and  
decreased apoptosis

CD28 → IL-2 production, but ...

??? → high IL-2 → FLIP → Fas-FasL inhibition → interacts with Caspase8 and 10 → decreased apoptosis.



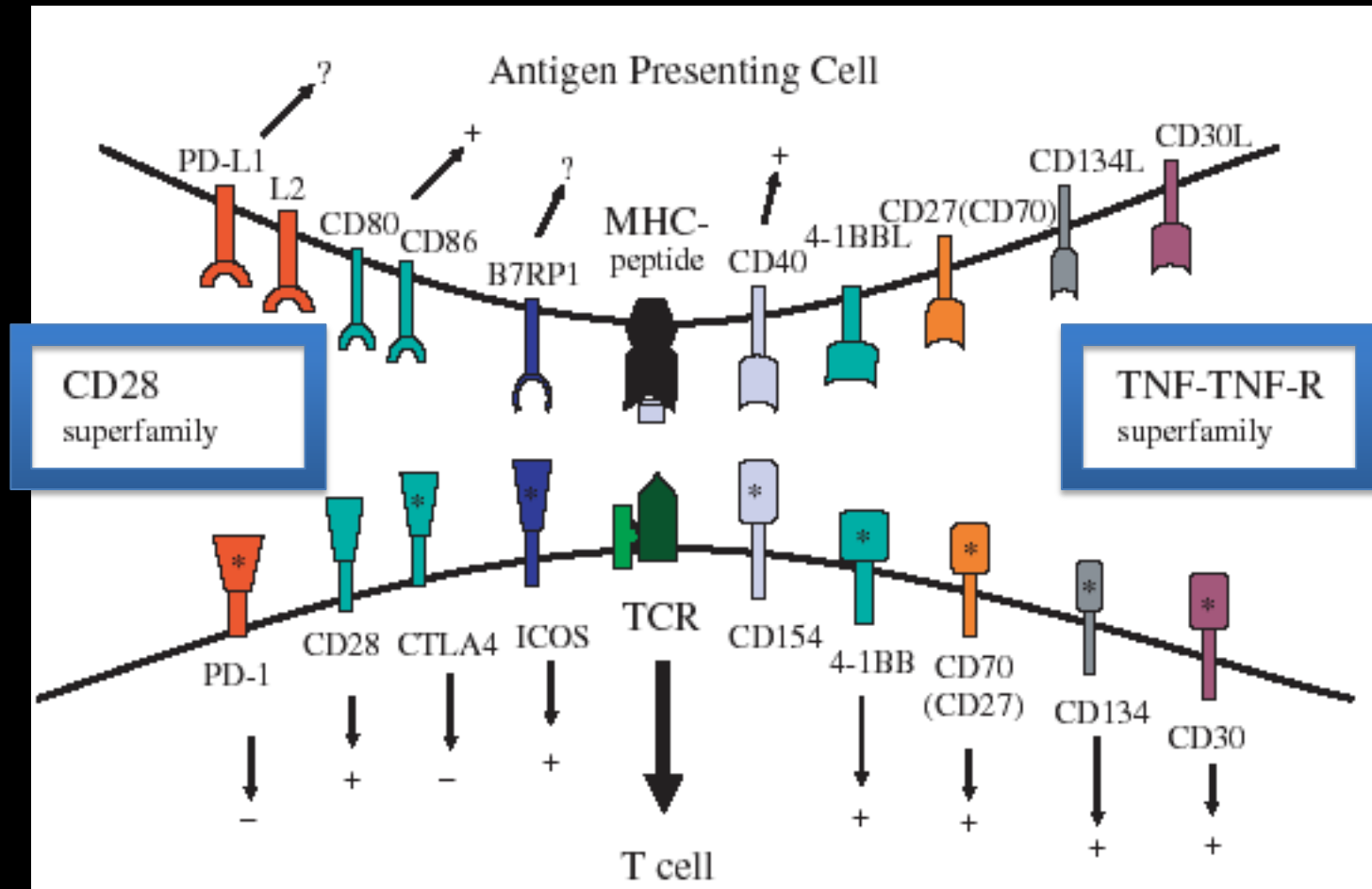
# Costimulation independent T cell activation??

- Early paper
- Markovic-Plese, Martin. *CD4<sup>+</sup>CD28<sup>-</sup> costimulation-independent T cells in multiple sclerosis*. J Clin Invest.

# Lessons from transplant bio

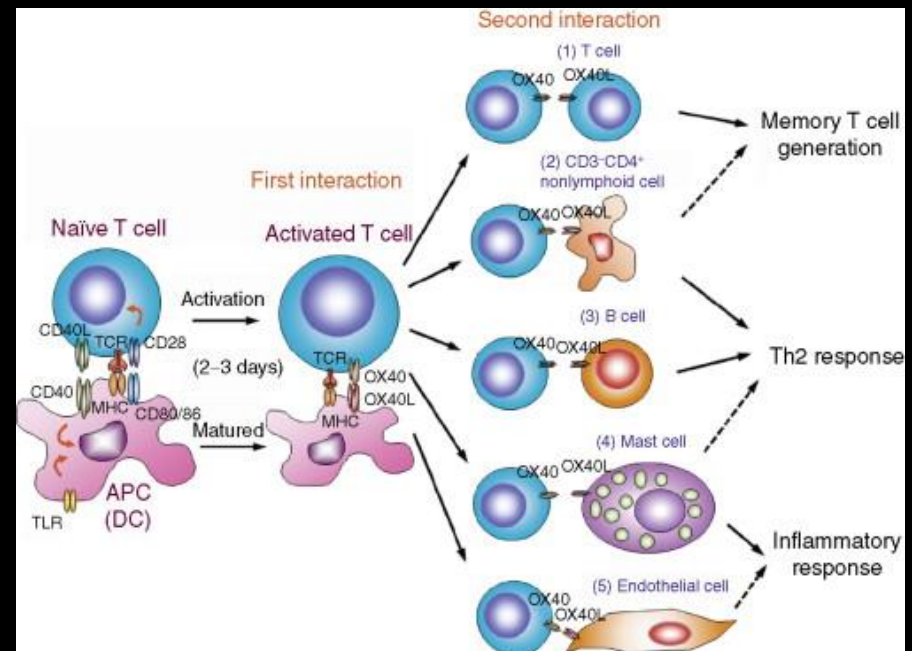
- Not necessarily costimulation independent
- Acute graft rejection
  - There are definitely ways of communicating with NK, effector T, memory cells independently of CD28/B7 ... can still get AGR
  - CD154/CD40L – CD40 another major route
  - 4-1BB – 4-1BBL underpin cytotoxic T cell production in graft rejection.
  - OX40 – CD134L

# Redundant (or hierachical?) pathways subserve costimulation



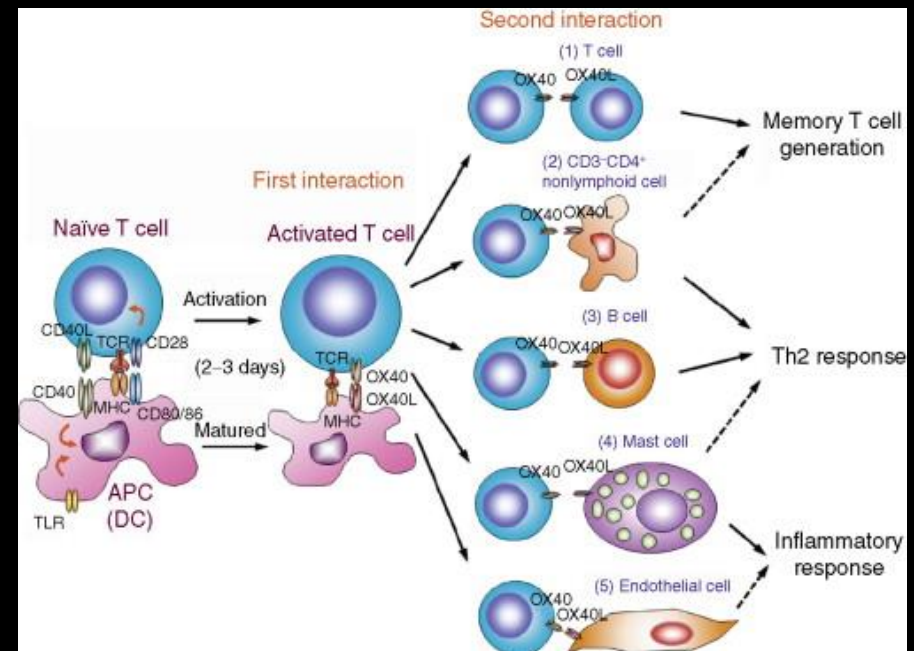
# OX40 & T-cell activation

- *CD134*; *TNFRSF4*
- TNFR family protein
- Expressed predominantly on **activated** T-cells
  - CD44+
- Ligand is *OX40L* (*CD252*)
  - APCs
- Blockade of *OX40*-*OX40L* binding improves CIA mice



# OX40 & T-cell activation, continued

- Seems to be able to drive Th1 or Th2 response
- Cytokine production, including signals to NK and NKT
- Effector T expansion and survival
- Appear to drive alloimmune (nonself from same species) Tcell stimulation



## Part II: study

Jiang et al. OX40 signaling is involved in the autoactivation of CD4+CD28– T cells and contributes to the pathogenesis of autoimmune arthritis. Arthritis Research & Therapy. 2017.

<https://doi.org/10.1186/s13075-017-1261-9>

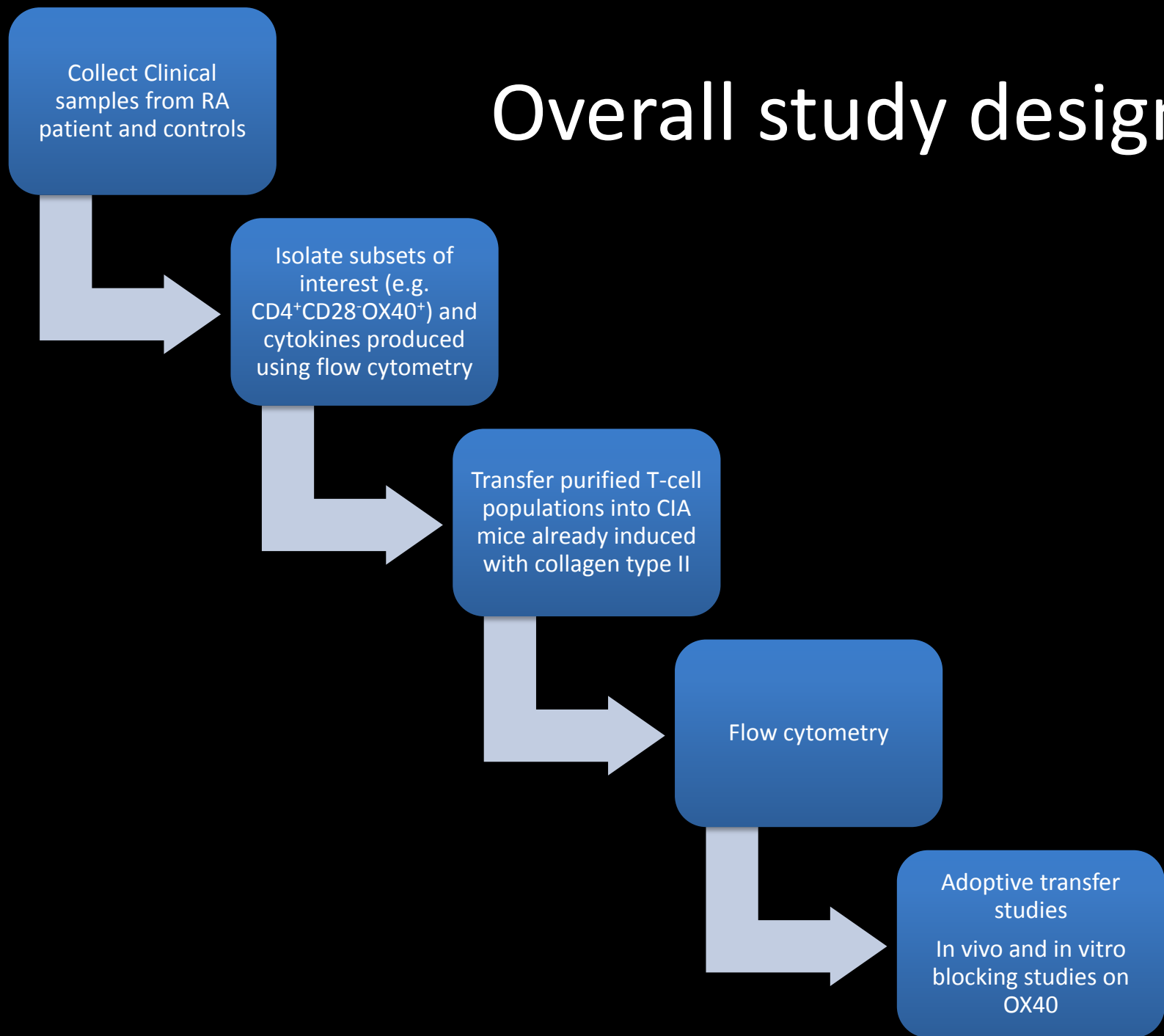
# Methods (Human Subjects)

- 71 RA (2010 ACR/EULAR)
- 44 sex & age matched OA patients
- 47 healthy volunteers
- DAS28 for disease activity – separated into 3 strata or in remission.
- 9 had been given MTX
- No other DMARDs w/in 1 year prior
- PB & SF collected after IC

# Methods (CIA mice)

- Male DBA/1 mice 8-10 wk
- D0 - Given 200ug bovine collagen type 2 & CFA
- D14 – reimmunized with CFA
- Scored 0-4 for degree of swelling in paw/wrist
- Grouped into Acute or Chronic CIA (A-CIA vs C-CIA).
  - Comments?
- D28 - Adoptive transfer and blocking studies
- D35 – dexamethasone given IP (0.5mg; 2mg; PBS)

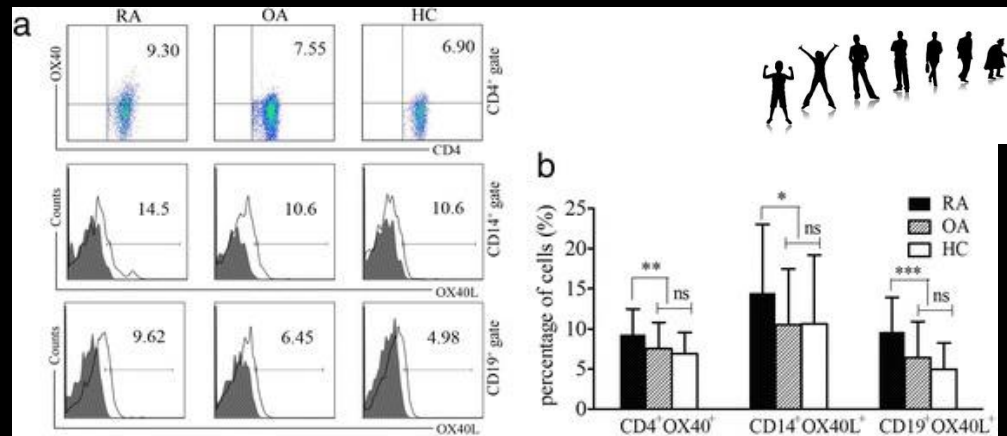
# Overall study design



# Fig 1: Differential expression of OX40 and OX40L in RA (1a, 1c) & CIA mice (1b)

A - PB of RA vs HC

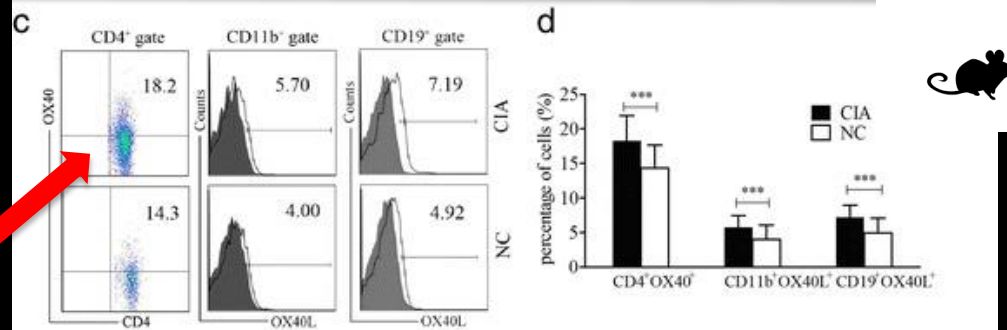
Top CD4<sup>+</sup>OX40<sup>+</sup>,  
Mid CD14<sup>+</sup>OX40L<sup>+</sup>  
Bot CD19<sup>+</sup>OX40L<sup>+</sup>



CD4 - T  
CD11b - NK,  
Macrophage, PMN  
CD14 - monocytes  
CD19 - B cells

C - Spleen in CIA

Top CIA  
Bot HC

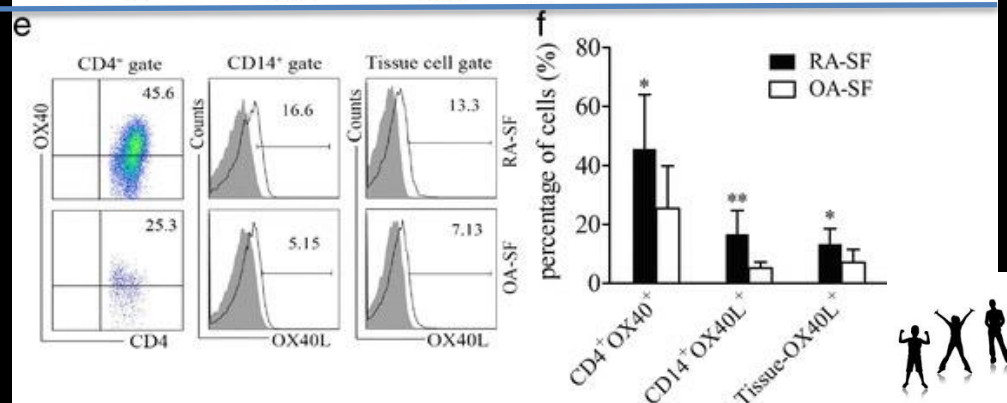


Comments

- Gates Non-spec?
- Gating (CD4 vs 44)
- Decades
- Naïve T (CD28-)

E - Synovial Tissue

Top RA SF  
Bot OA SF



# Figure 2

A,B,C - % CD4+ T cells in RA PB:

A – CD28-

B – CD28-OX40+

C - CD28+OX40+

Flow panels?

D – SF (RA vs OA) (same markers as A-C)

E – Quantification in RA vs OA.

F, G, H % splenic T cells in CIA and NC (same markers as A-C)

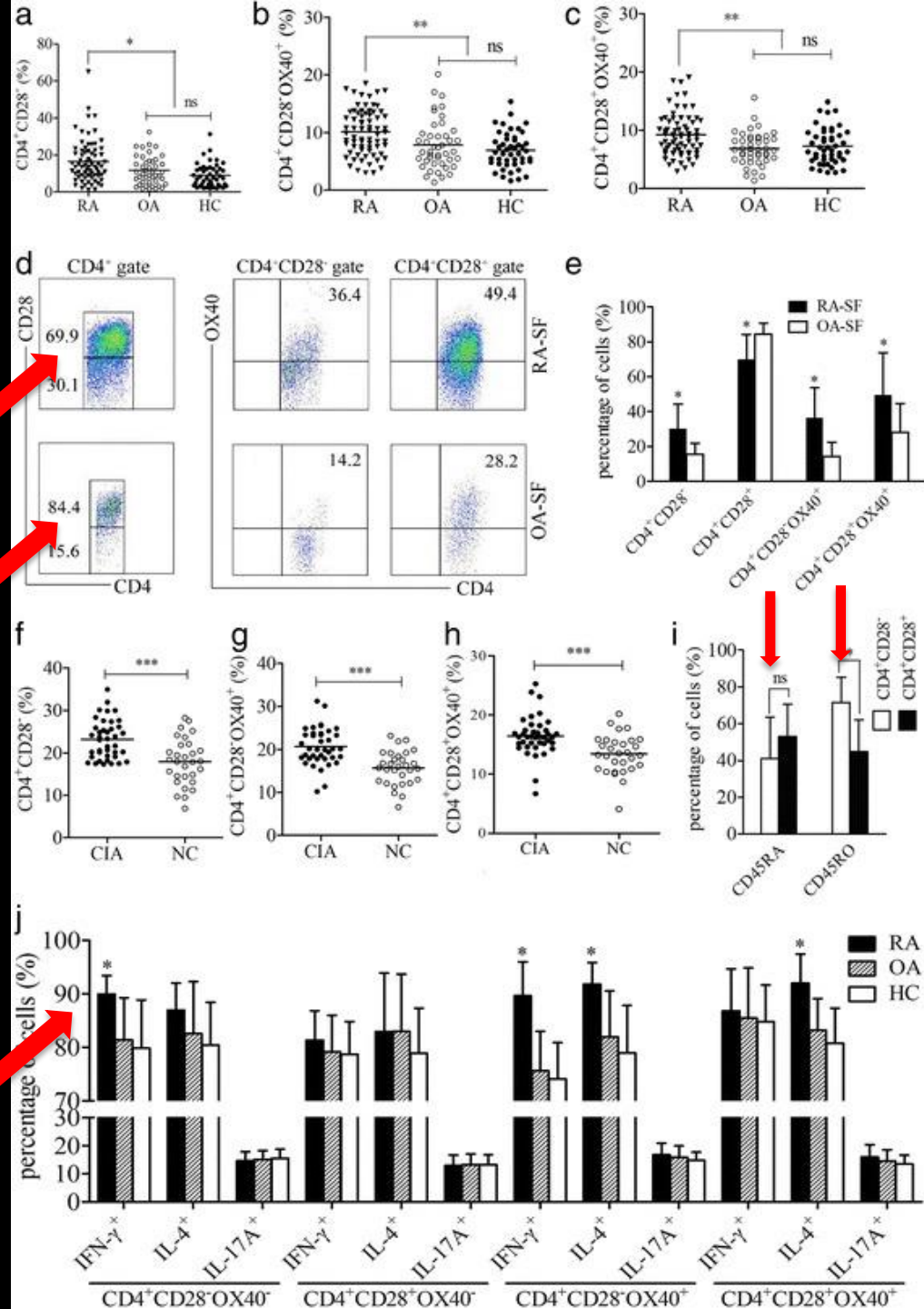
Flow panels?

I – CD45RA and CD45RO expression differs between CD4+CD28- and CD4+CD28+ populations in RA PB

- CD45RA – naïve; CD45RO – memory
- % look inherently incorrect to me

J – Cytokine production for gamma, IL-4, and IL-17A in PB samples from RA and control

- IFN $\gamma$  and IL4 with naïve cells?

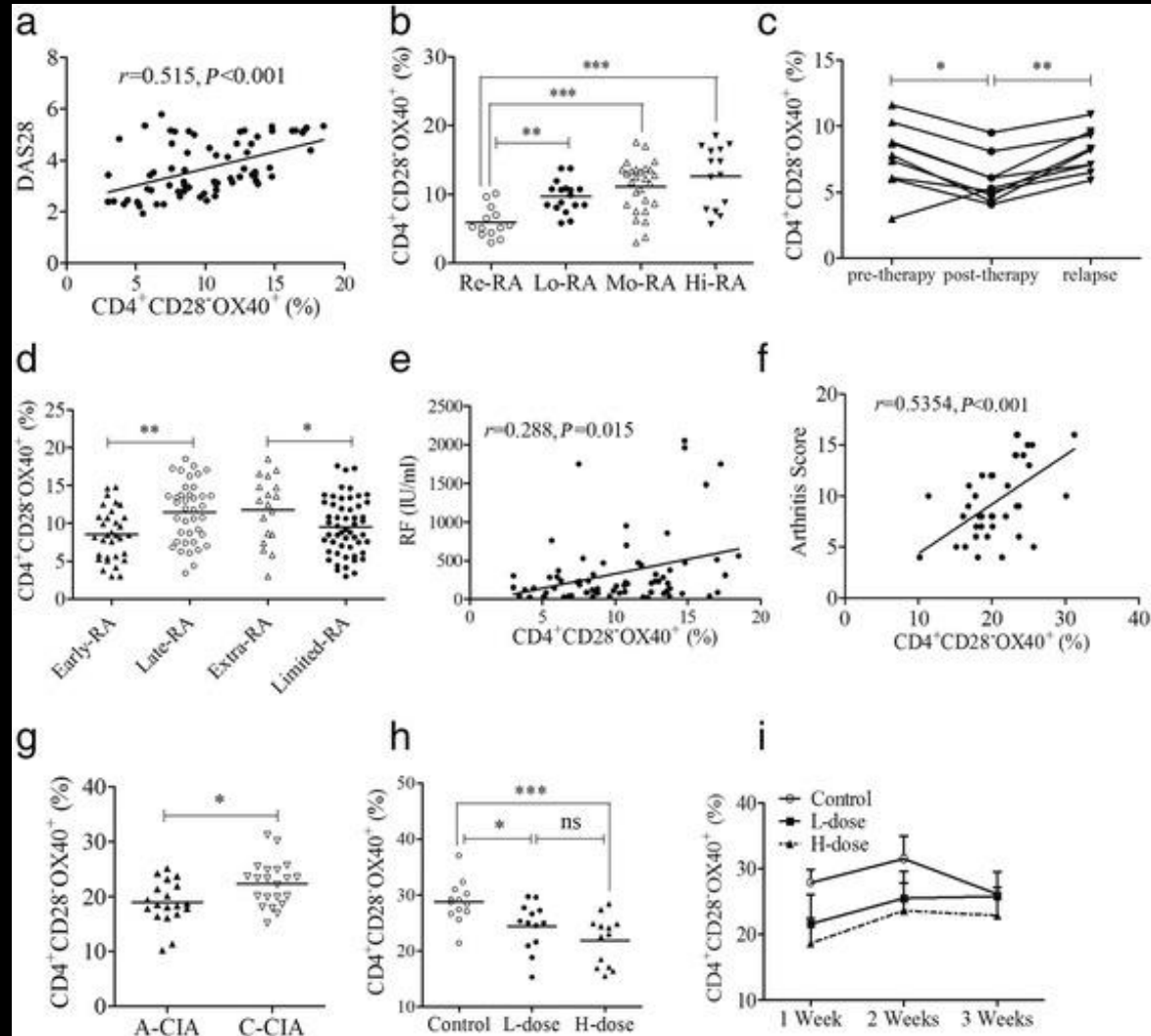


# Figure 3 – CD4<sup>+</sup>CD28<sup>-</sup>OX40<sup>+</sup> T-cells correlate with RA clinical indicators

A – DAS28  
 B – RA Sev (EULAR)  
 C – MTX tx response  
 D – RA Stage (Huizinga 2002)  
 E – RF titer  
 F – CIA mouse arthritis score  
 G – A-CIA and C-CIA % (Thornton 2000)  
 H – Dex dose or PBS (DD)  
 I – Change in subset frequency over time (NS)

R values range 0.29 – 0.53

Citations found in Jiang et al.



# Fig 4 – Adoptive Transfer of T cell subsets into CIA mice

D28 → sac → spleen → purify → AT into D0

A – CD28+OX40-

B – CD28-OX40+

C – CD28+OX40+

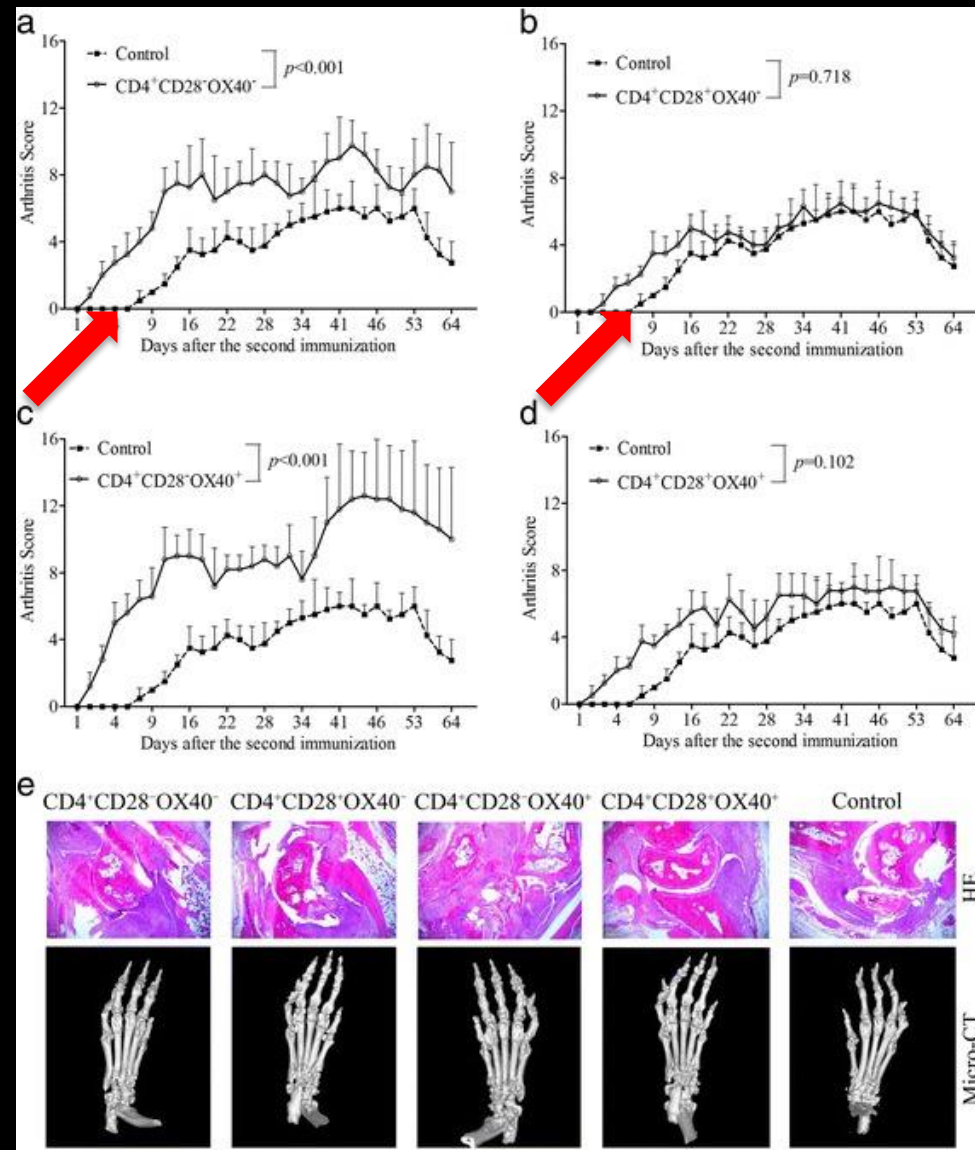
D – CD28-OX40-

Arthritis began earlier and had much much higher arthritis scores in CD4+ CD28- OX40+

compared to “control”

E – (top) H&E @ 200x of ankle sections

(bot) micro-computed tomographic analysis of ankles of animals from A-D.

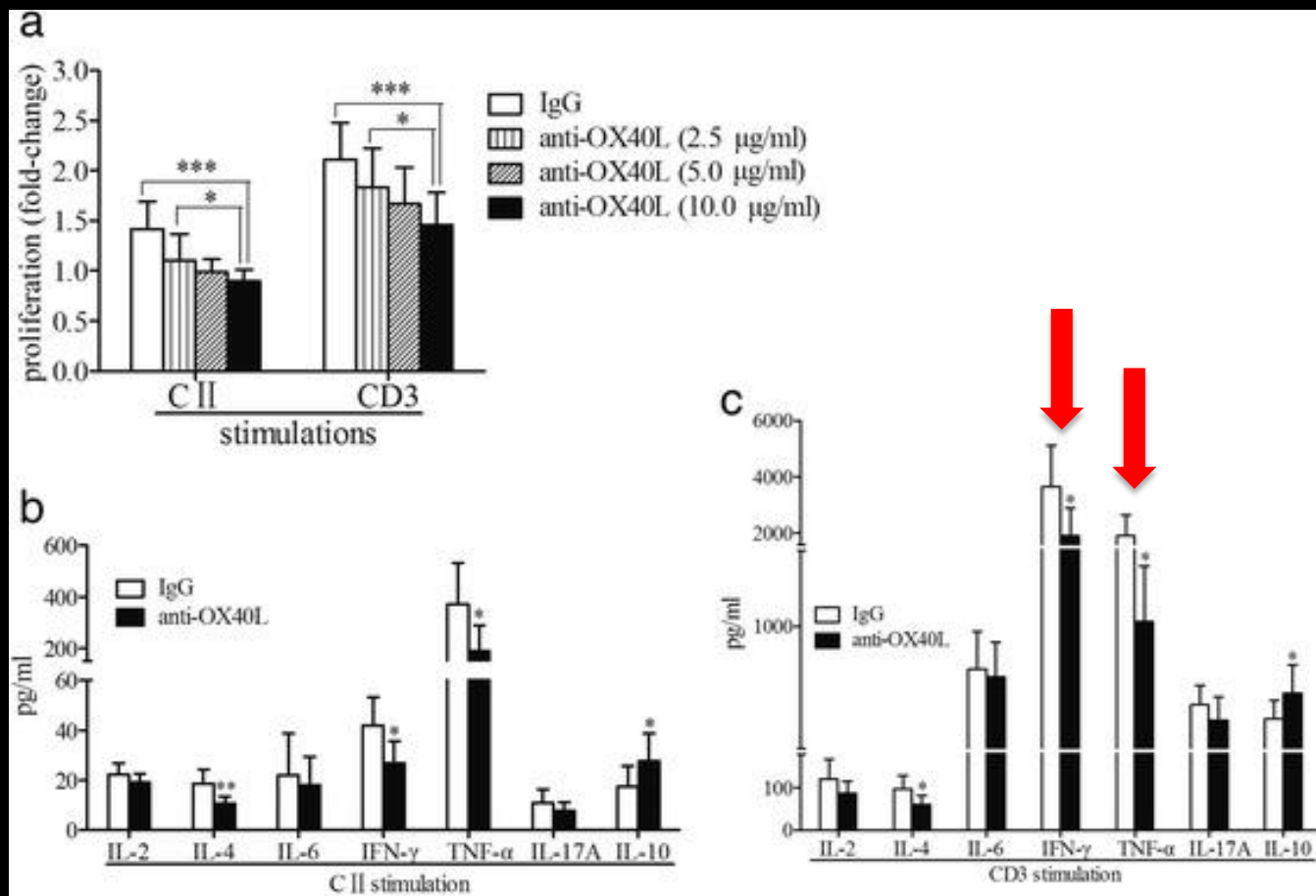


# Figure 5 – *In vitro* blocking of OX40L

- A OX40L blocking Ab **reduces** cell proliferation (fold change compared to IgG controls) **on splenocytes**
- B,C OX40L blocking Ab **reduces** cytokine secretion of CII or CD3-stimulated **splenocytes**, respectively (pg/ml)

D28  
Sac  
Spleen  
96 well plate  
Anti-CII or anti-CD3 mAb  
Stimulate  
Rat IgG used for control

AT studies vs blocking studies (see next slide)



# Figure 6 – OX40/40L blockade *in vivo*

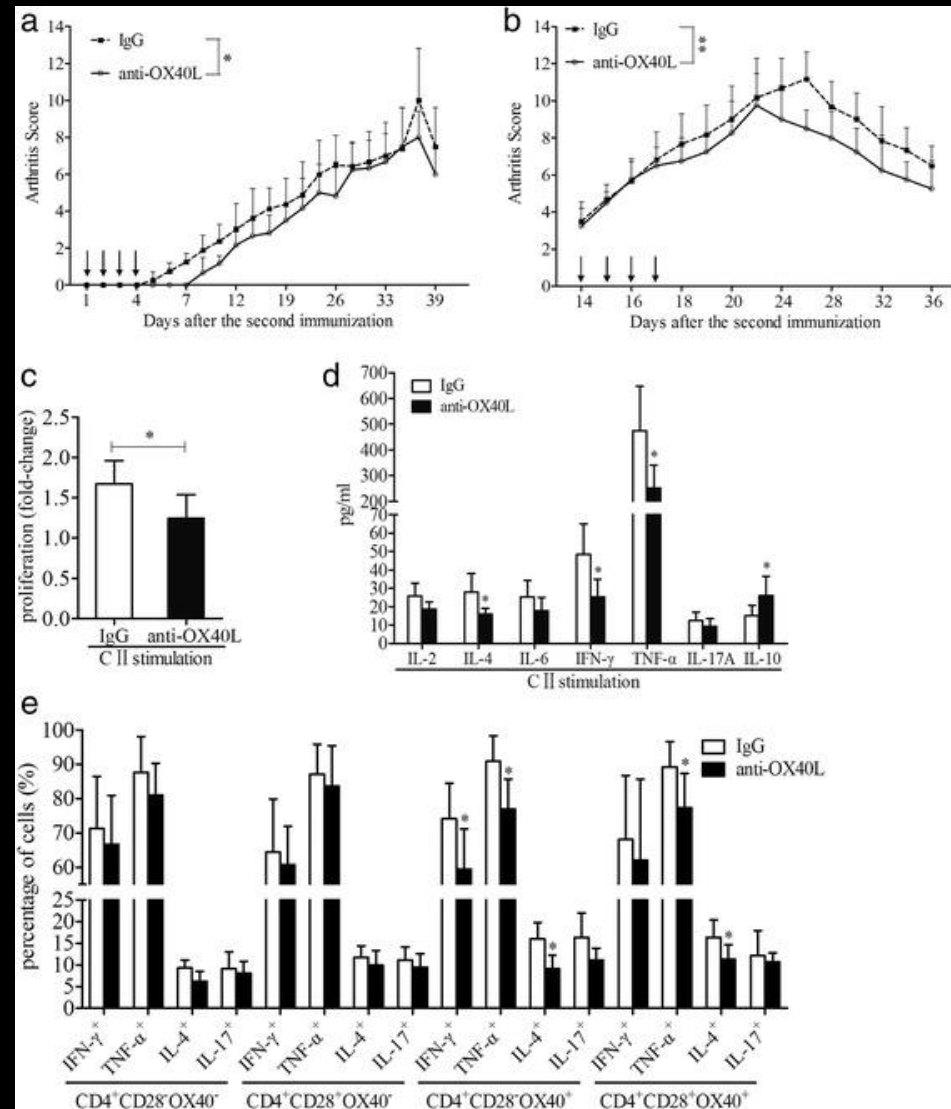
## s.p. 2<sup>nd</sup> immunization

A,B OX40 blockade on D1 and D14 delays arthritis onset and scores of CIA mice compared with IgG controls, *in vivo*.

C Decreases proliferative index after CII stimulation

D Decreases cytokine Secretion a/f CII stimulation

E Decreases cytokine secretion in CD4<sup>+</sup>CD28<sup>-</sup>OX40<sup>+</sup> cells, but not others.



# Blocking studies on unpurified populations

- In Blocking studies (in vitro and in vivo) T cells were not purified from mononuclear splenocytes (cf. adoptive transfer studies).
- This raises a variety of technical issues.
  - Example: Mphage, DC, PMN, present
    - Have FcR
    - FcR binds heavy chain of Ab.
    - This type of interaction could be counted as a positive event rather than the desired idiotypic binding
    - Could account for such high cytokine production levels
  - Too many cells per well → influences MIF
  - Etc.

# Discussion

CD4+CD28-OX40+ cells are clinicopathologically significant cell type

OX40 expression level was independent of CD28 level, which fits with other studies; could also present some rationales as to why CD28 expression could be driven down

I'd like to see the findings replicated in particular a little more carefully w/ respect to the flow data and the blocking studies.

I'd also like to see cohort characteristics granted what we know of age-dependency of this T-cell subset from other studies

I'd like to see some different comparisons made than what they make in particular in Fig 4.

Thanks for your attention!